PAPER NAME AUTHOR

Effect of NR-g-cellulose coupling agent i nto NR-cellulose composite dispersibility and its physical

Hani Handayani

WORD COUNT CHARACTER COUNT

2667 Words 15283 Characters

PAGE COUNT FILE SIZE

7 Pages 313.4KB

SUBMISSION DATE REPORT DATE

Feb 20, 2023 4:16 PM GMT+7 Feb 20, 2023 4:17 PM GMT+7

11% Overall Similarity

The combined total of all matches, including overlapping sources, for each database.

- 9% Internet database
- 5% Submitted Works database
- 0% Publications database

Excluded from Similarity Report

- Crossref database
- · Bibliographic material
- · Manually excluded sources

- Crossref Posted Content database
- Cited material
- · Manually excluded text blocks

Micro-fibrillated Cellulose Fabrication from Empty Fruit Bunches of Oil Palm

Marcelinus Christwardana^{1,a}, Aniek Sri Handayani^{1,b,*}, Shirley Savetlana^{2,c}, Riana Herlina Lumingkewas^{3,d}, Mochamad Chalid^{4,e}

- Department of Chemical Engineering, Institut Teknologi Indonesia, Jl. Raya Puspitek Serpong, South Tangerang, 13514 Indonesia
- ² Department of Mechanical Engineering, Faculty of Engineering, Universitas Lampung, Indonesia 35145
- ³ Department of Civil Engineering, Institut Teknologi Indonesia, Jl. Raya Puspitek Serpong, South Tangerang, 13514 Indonesia
 - ⁴ Department of Metallurgical and Materials Engineering, Faculty of Engineering, Universitas Indonesia, Depok, West Java, 16424 Indonesia

amarcelinus@iti.ac.id, baniek.handayani@iti.ac.id, cShirley.savetlana@eng.unila.ac.id, driana.herlina@iti.ac.id, eMchalid@gmail.com

* corresponding author: Aniek Sri Handayani

Keywords: Micro Fibrillated Cellulose, oil palm empty fruit bunch, DSC, TGA, SEM

Abstract. Micro-fibrillated celluloses (MFCs) are made from oil palm empty fruit bunches (EFB). EFB is processed through several stages of the process, including washing, alkalization, and bleaching to remove impurities, lignin, and hemicellulose. Each treatment stage was characterized by differential scanning calorimeter (DSC) and thermogravimetric (TGA) analysis. Morphological analysis was characterized using Scanning Electron Microscope (SEM). The process results show that MFC has an average length and thickness of 450 and 80 microns for coarse fibers respectively, averaging 50 and 5 microns for fine fibers, respectively. Fibrillation fibers appear on the surface of fibers which are treated using alkalization and bleaching processes. The TGA results showed a decrease in weight occurred at a temperature of 40 to 109 °C for the first stage of the heating process and at a temperature of 247 to 382 °C for the second stage. The decrease in fiber weight is caused by evaporation of water content and degradation of cellulose compounds at each stage. The glass transition temperature of MFC was obtained at 236 °C. The thermal stability of cellulose from fibers treated using alkalization and bleaching processes proved the formation of cellulose crystals. Removal of lignin and hemicellulose is shown by the absorption of O-H and C-C bonds in FTIR spectroscopy. From these results, it is stated that micro-fibrillation cellulose is formed well through a series of processes given.

17ntroduction

Cellulose is one of the most important renewable materials for human life. Cellulose is crystalline, hard fibrous which is widely used in the textile, paper, sanitary wares, and packaging industries [1]. Cellulose-based materials can be used with the presence or absence of lignin and hemicellulose.

At present, cellulose had been produced using mechanical [2], Biological [3], and chemical method [4]. Chemical methods prefer to be used in cellulose fabrication because the product yields are higher than mechanical methods that require much energy. Cellulose production using biological methods is less preferred because of the long process. Making cellulose by chemical methods can eliminate lignin and hemicellulose quickly so that the fibrillation process runs faster to cellulose

fibrillation. The fibrillation process on cellulose through this chemical method can also provide different characteristics and is better for cellulose fibrillation produced.

Microfibril cellulose generally has a fibrous cellulose structure as thick as 2-10 nm with a length of several tens of microns, varying depending on the origin [5]. Usually, MFC consists of aggregates of cellulose microfibrils [6]. Cellulose microfibrils undergo transverse division along the amorphous region due to treatment, both mechanical, biological, and/or chemical. This transverse division makes the MFC aggregate have strong bonds between the components [7].

indonesia is one of the largest palm oil producers in the world. The palm oil industry produces waste, one of which is oil palm empty bunches (EFB). According to Pattanamanee et al., [8], EFB waste produced globally reaches 40 million tons. So far, EFB is only used for fertilizers because it has high organic and inorganic content [9]. In addition, EFB is also used as fuel through an incineration process [10]. Seeing this, the utilization of EFB waste is still not high. EFB contains much fiber has excellent potential to be developed into micro-fibrillated cellulose material, which has a higher economic value than as a raw material for fertilizer and combustion processes.

In addition to EFB, several factories have developed as raw materials to make MFC. Yuanita, et al. [11] investigated the manufacture of MFCs from palm oil fibers. Kumar et al. [12] have researched the production of MFC from bagasse and straw fiber. Meanwhile, Ismojo et al. [13] have made MFC using sorghum raw materials. Therefore, the above has reinforced the estimate that EFB can be used in making MFC.

In this study, MFC was made from EFB fibers through several stages, including the alkalization and bleaching process. The alkalization process is used to break down lignin and hemicellulose and to remove impurities. This process also aims to increase resistance to water so that the fiber has hydrophobic properties. After the alkalization process, the fiber goes through a bleaching process which aims to (i) whiten the color of MFC and (ii) fibrillate the surface of the cellulose fiber. Scanning Electron Microscopy (SEM) was used to examine EFB fiber morphology before and after treatment and to prove the formation of MFC. Thermogravimetric (TGA) and Differential Scanning Calorimeter (DSC) analyzes were carried out to prove cellulose crystal formation and removal of lignin and hemicellulose as impurities, as well as Fourier-transform infrared spectroscopy (FTIR) analysis.

Materials and Method

Materials. Empty fruit bunches of oil palm waste were obtained from an oil palm plantations in Ciseeng, West Java, Indonesia. Sodium hydroxide and hydrogen peroxide were purchased from Merck (Darmstadt, Germany).

Micro-fibrillated Cellulose Fabrication. Before use, oil palm empty bunches fibers were dried under the sun (29-32 °C) for three days. After drying, the fiber was broken down and crushed using a crusher then filtered with a 40 mesh sieve until the average size was 3 mm. The alkalization process was carried out by immersing 1 g of crushed fiber in 35 mL of 4% sodium hydroxide solution at 70 °C for 3 hours while continuing to stir for the product to be homogeneous. The bleaching process was carried out by dipping the alkaline fiber into hydrogen peroxide at room temperature for two hours, and then the fiber was dried under 50 °C overnight to produce micro-fibrillation cellulose.

Characterization

scanning Electron Microscopy (SEM). SEM was used to analyze the morphology of MFC, which was formed after treatment with alkalization and bleaching. SEM micrographs were taken using a Field-Emission Scanning Electron Microscope (FE-SEM) FEI Inspect F50 (Oregon, USA). The sample was prepared by dispersing dry MFC powder on a two-sided conductive adhesive tape and coated with carbon to make it more conductive.

Thermal Stability Analysis. Simultaneous Thermal Analysis ²TA-6000 from Perkin Elmer (Massachusetts, USA) was used for DSC-TGA analysis simultaneously of the initial fiber, after alkalization, and after bleaching, under nitrogen flow at a rate of 10 mL/min. It was calibrated against melting points and fusion enthalpies of indium and silver. Samples (9-10 mg) were heated from 40 to 500 °C at a rate of 10 °C/min.

Fourier Transform Infra-Red (FTIR) Spectroscopy. Spectrum TwoTM Infrared Spectrometer Perkin Elmer (Massachusetts, USA) was used to determine the chemical bonding of treated EFB micro-fibrillated cellulose.

4 Results and Discussion

Morphology Analysis. Figure 1 shows the micrographs of the fibers before processing and after being processed through alkalization and bleaching. In untreated fibers, the fiber surface was relatively smooth, and some impurities are still found. The average fiber length was around 500 microns for both crude fiber and fine fiber. Whereas in fibers treated with alkalization and bleaching to form MFC, the surface of the fiber looks different where cellulose fibers are clearly visible on crude fibers. Lignin and hemicellulose then dissolved during the alkalizing and bleaching process. Coarse fiber had an average length of 400 microns, and fine fibers had a length between 50-100 microns. This proves that fiber treatment using alkalization and bleaching can reduce fiber size.

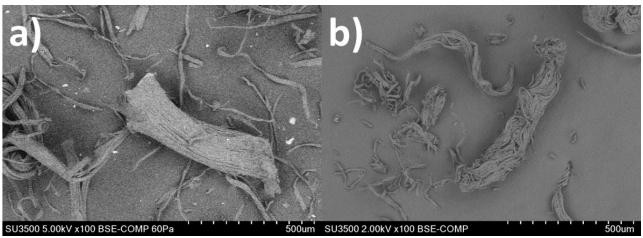


Figure 1. SEM images of a) un-treated fiber and b) fiber which treated using alkalization and bleaching process.

Thermal Stability Analysis. Investigating the thermal properties of each treatment process in MFC production from oil palm empty bunches is essential to do. Figure 2a shows the TGA for fiber samples (i) before treatment, (ii) after alkalization, and (iii) after bleaching. All TGA curves show a decrease in weight in two stages. In the fiber sample before treatment, the weight loss from 40 to 136 °C (first stage) indicating the drying process that causes evaporation of water contained in the sample. Then the weight loss dramatically at temperatures around 220-347 °C (the second stage) due to the degradation process of the components such as cellulose, hemicellulose, lignin at the same time. In fiber samples after the alkalization process, the first stage of weight loss occurred from 40 °C until the temperature reached 116 °C, and the second stage occurred at 237-357 °C. Whereas in the fiber sample after the bleaching process, the first stage of weight loss occurred until temperature achieved 109 °C and the second stage at 247-382 °C. The increasing decomposition temperature after the alkalization and bleaching process was caused by the crystallization of cellulose, which has better thermal stability than the initial fiber [14,15]. The crystalline of cellulose

causes fibers through the process of treatment (alkalization and bleaching) to have high thermal stability compared to alkalinized fiber, and fiber without treatment [16].

The DSC thermograms from heating fiber samples (i) before treatment, (ii) after alkalization, and (iii) after bleaching are shown in Figure 2b. Thermograms of fibers after alkalization process had Tg of 238 °C, higher than fibers without treatment and fiber after the bleaching process which had Tg around 235 and 236 °C, respectively. All thermograms had two melting peaks. In general, the melting peak occurs due to melting and recrystallization process of the crystalline phenomenon during the heating process [17]. In the initial fiber (without treatment), the second peak appeared around 340 °C. In the fiber after the alkalization process and after bleaching, the second peak appeared when the temperature was 353 and 369 °C, respectively. The interaction between cellulose crystals can slow down the melting process of cellulose.

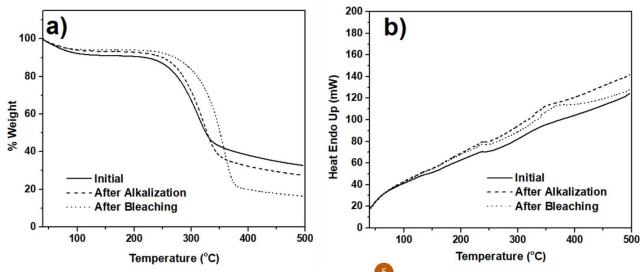


Figure 2. a) TGA and b) DSC curves of MFC samples; the neating rate of 10 °C/min

FTIR Analysis. The width of the relative absorbance peak at 2895.88 cm⁻¹ occurs because of the presence of CH_2 groups in the MFC sample [18]. This band also represent the crystallinity degree of cellulose. While high and sharp peak intensity at 1029.24 cm⁻¹ showed an increase in cellulose content which has a skeletal ring C-O-C pyranose. The peak at 892.60 cm⁻¹ was small but sharp, representing the glycosidic deformation of C_1 -H, which was related to β-glycoside bond between anhydroglucose units in cellulose [19].

A prominent peak at 1639.08 cm⁻¹ seen in the spectrum illustrates the presence of esters of the carboxylic and p-coumaric groups or acetyl groups and uronate esters of lignin and/or hemicellulose [20]. The peak at 1510.02 cm⁻¹ represented the aromatic C-C stretch of the lignin aromatic ring [21]. The small absorbance at 1639.08 cm⁻¹ and about 1510.02 cm⁻¹ showed that hemicellulose, pectin, and lignin in oil palm empty bunches fibers were effectively removed by leaving cellulose as the main component. The peaks at 3334.49 cm⁻¹ were related to O-H bonds which shows the degradation of hydrogen bonds between lignin and carbohydrates or with cellulose.

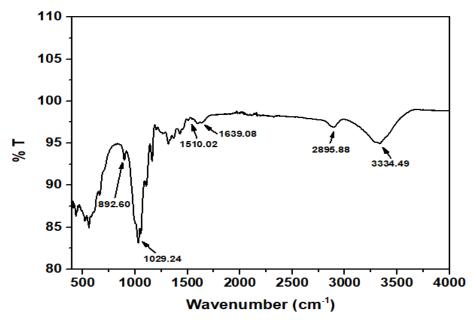


Figure 3. FTIR spectra of cellulose sample after alkalization and bleaching process

Summary

Chemical treatments such as alkalization and bleaching process had been carried out for the fabrication of micro-fibrillated cellulose from oil palm empty bunches. The morphology of the fiber changes drastically after undergoing an alkalization and bleaching process in which lignin and hemicellulose are removed and make fibrillated fibers clear. Fibers treated with alkalization and bleaching have better thermal stability than the initial fibers without treatment, and an alkalization treatment with degradation temperature was 247-382 °C. This proves the presence of cellulose crystals will slow down the decomposition of fiber material. Two peaks were also seen at temperatures 236 and 369 °C during DSC analysis, also proved the existence of the produced cellulose crystals. FTIR spectra strengthen the analysis of the formation of cellulose crystals and the elimination of lignin and hemicellulose from micro-fibrillated cellulose produced from these oil palm empty bunches, where the O-H and C-C bonds had relatively small peak spectrum intensities.

Acknowledgment

This work was carried out with the financial support of Competitive Research Grant from Kemenristek- Dikti 2019 with the contract number: 43/AKM/MONOPNT/2019.

References

- [1] A. M. Adel, A. A.El-Gendy, M. A. Diab, R. E. Abou-Zeid, W. K. El-Zawawy, A. Dufresne, Microfibrillated cellulose from agricultural residues. Part I: Papermaking application. Industrial Crops and Products, 93 (2016) 161-174.
- [2] H. Liimatainen, J. Sirviö, A. Haapala, O. Hormi, J. Niinimäki, Characterization of highly accessible cellulose microfibers generated by wet stirred media milling. Carbohydrate Polymers, 83 (2011) 2005-2010.

- [3] A. M. Adel, Z. H. A. El–Wahab, A. A. Ibrahim, M. T. Al–Shemy, Characterization of microcrystalline cellulose prepared from lignocellulosic materials. Part I. Acid catalyzed hydrolysis. Bioresource technology, 101 (2010) 4446-4455.
- [4] A. M. Adel, Z. H. A.El-Wahab, A. A. Ibrahim, M. T. Al-Shemy, Characterization of microcrystalline cellulose prepared from lignocellulosic materials. Part II: Physicochemical properties. Carbohydrate Polymers, 83 (2011) 676-687.
- [5] H. A. Krässig, Cellulose-structure. Accessibility and Reactivity. Gordon and Breach Science Publishers, Yverdon, 1993.
- [6] A. J. Svagan, M. A. Azizi Samir, L. A. Berglund, Biomimetic polysaccharide nanocomposites of high cellulose content and high toughness. Biomacromolecules, 8 (2007) 2556-2563.
- [7] B. G. Rånby, Fibrous macromolecular systems. Cellulose and muscle. The colloidal properties of cellulose micelles. Discussions of the Faraday Society, 11 (1951) 158-164.
- [8] W. Pattanamanee, Y. Chisti, W. Choorit, Photofermentive hydrogen production by Rhodobacter sphaeroides S10 using mixed organic carbon: effects of the mixture composition. Applied energy, 157 (2015) 245-254.
- [9] E. Derman, R. Abdulla, H.Marbawi, M. K. Sabullah, Oil palm empty fruit bunches as a promising feedstock for bioethanol production in Malaysia. Renewable energy, 129 (2018) 285-298.
- [10] J. W. Park, J. Heo, H. V. Ly, J. Kim, H. Lim, S. S. Kim, Fast pyrolysis of acid-washed oil palm empty fruit bunch for bio-oil production in a bubbling fluidized-bed reactor. Energy. 179 (2019) 517-527
- [11] E. Yuanita, J. N. Pratama, J. H. Mustafa, M. Chalid, Multistages preparation for microfibrillated celluloses based on Arenga pinnata "ijuk" fiber. Procedia Chemistry, 16 (2015) 608-615.
- [12] A. Kumar, Y. S. Negi, V. Choudhary, N. K. Bhardwaj, Characterization of cellulose nanocrystals produced by acid-hydrolysis from sugarcane bagasse as agro-waste. Journal of Materials Physics and Chemistry, 2 (2014) 1-8.
- [13] Ismojo, P. H. Simanulang, A.Zulfia, M. Chalid, Preparation of micro-fibrillated cellulose from sorghum fibre through alkalization and acetylation treatments. In IOP Conference Series: Materials Science and Engineering, 223 (2017) 012057.
- [14] U. J. Kim, S. H. Eom, M. Wada, Thermal decomposition of native cellulose: influence on crystallite size. Polymer Degradation and Stability, 95 (2010) 778-781.
- [15] M. Poletto, V. Pistor, M. Zeni, A. J. Zattera, Crystalline properties and decomposition kinetics of cellulose fibers in wood pulp obtained by two pulping processes. Polymer Degradation and Stability, 96 (2011) 679-685.
- [16] A. Mandal D.Chakrabarty, Isolation of nanocellulose from waste sugarcane bagasse (SCB) and its characterization. Carbohydrate Polymers, 86(2011) 1291-1299.
- [17] R. Masirek, Z. Kulinski, D. Chionna, E. Piorkowska, M. Pracella, Composites of poly (L-lactide) with hemp fibers: Morphology and thermal and mechanical properties. Journal of applied polymer science, 105 (2007) 255-268.
- [18] R. D. Kalita, Y. Nath, M. E. Ochubiojo, A. K. Buragohain, Extraction and characterization of microcrystalline cellulose from fodder grass; Setaria glauca(L) P. Beauv, and its potential as a drug delivery vehicle for isoniazid, a first line anti tuberculosis drug. Colloids and Surfaces B: Biointerfaces, 108 (2013) 85–89.

- [19] A. Alemdar, M. Sain, Isolation and characterization of nanofibers from agricultural residues-Wheat straw and soy hulls. Bioresource Technology, 99 (2008) 1664–1671.
- [20] E. M. Maafi, F. Malek, L. Tighzert, P. Dony, Synthesis of polyurethane and characterization of its composites based on alfa cellulose fibers. Journal of Polymer Environment, 18 (2010) 638–646.
- [21] H. Nadji, P. N. Diouf, A. Benaboura, Y. Bedard, B. Riedl, T. Stevanovic, Comparative study of lignins isolated from Alfa grass (Stipa tenacissima L.). Bioresource Technology, 100 (2009) 3585–3592.

11% Overall Similarity

Top sources found in the following databases:

• 9% Internet database

- 0% Publications database
- 5% Submitted Works database

TOP SOURCES

The sources with the highest number of matches within the submission. Overlapping sources will not be displayed.

Universiti Teknologi MARA on 2016-12-28 Submitted works	1%
ojs.jmolekul.com Internet	1%
jos.unsoed.ac.id Internet	<1%
hindawi.com Internet	<1%
mdpi.com Internet	<1%
core.ac.uk Internet	<1%
idoc.pub Internet	<1%
ncbi.nlm.nih.gov Internet	<1%
eprints.utar.edu.my Internet	<1%

10	repository.iti.ac.id Internet	<1%
11	intechopen.com Internet	<1%
12	kompozit.org.tr Internet	<1%
13	researchgate.net Internet	<1%
14	University of Stellenbosch, South Africa on 2019-08-02 Submitted works	<1%
15	coek.info Internet	<1%
16	ijtech.eng.ui.ac.id Internet	<1%
17	vdoc.pub Internet	<1%
18	Universiti Malaysia Perlis on 2023-01-31 Submitted works	<1%

Excluded from Similarity Report

- Crossref database
- Bibliographic material
- Manually excluded sources

- Crossref Posted Content database
- Cited material
- Manually excluded text blocks

EXCLUDED SOURCES

scientific.net

Internet

1%

EXCLUDED TEXT BLOCKS

Micro-fibrillated Cellulose Fabrication from Empty Fruit Bunches of OilPalmMarcel...

www.researchgate.net

Micro-fibrillated celluloses (MFCs) are made from oil palm empty fruit bunches (E...

www.researchgate.net

AcknowledgmentThis work was carried out with the financial support of Competiti...

repository.iti.ac.id

1 Department of Chemical Engineering, Institut Teknologi Indonesia, Jl. Raya

ejournal.undip.ac.id

Department of

ejournal.undip.ac.id

4

humaniora.journal.ugm.ac.id