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Bioassay-Guided Separation Approach for Characterization of New Antibacterial Fractions from the Stem Roots Extracts of *Archidendron jiringa*

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Abstract: Infectious diseases caused by bacteria has become a global health issue, especially antibacterial drug resistance. The most serious concern with antibacterial resistance is that some bacteria became resistant to almost all antibacterial drugs, which makes them less effective. *Archidendron jiringa* is one of the most potent medicinal plants to be developed as a new source of antibacterial components. In current study, based on the antibacterial assay-guided approach, the separation of bioactive fractions of *A. jiringa* stem roots was carried out through several stages including isolation, fractionation, and characterization. The stages of isolation of secondary metabolites were conducted by gradually extraction followed by fractionation using chromatographic methods. The antibacterial potential of extract was evaluated by the disc diffusion and microdilution methods employing the resazurin assay against one Gram-negative resistant bacteria, *Escherichia coli*, and one Gram-positive bacteria, *Bacillus subtilis*. Among three extracts obtained, ethyl acetate and methanol extracts demonstrated to be the most significant antibacterial effects, while no antibacterial activity was shown on the n-hexane extract. The fractionation of ethyl acetate extract led to the isolation of the most bioactive fractions (E₂8₁5 and E₂8₁6) with the MIC's values ranging of 12.5-25 µg/mL for both resistant bacteria. Due to the low amount, only the fraction E₂8₁6 was subjected to analysis by ¹H-NMR spectroscopy. The results exhibited that the bioactive fraction was obtained as a mixture of at least three major constituents. However, the purification of the bioactive fraction is required, to further clarify the antibacterial compound that can be utilized as a new promising antibacterial agent. The bioassay-guided separation approach and the dye resazurin as an indicator of the growth of bacteria are applied for the first time for the phytopharmacological investigation from this plant. The present study represented the most effective method for subsequent finding and isolation of potential novel antibacterial constituents from *A. jiringa* stem roots, in particular against the multi-drug resistant strains.

Keywords: *A. jiringa*, antibacterial assay, bioassay-guided separation, medicinal plant

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INTRODUCTION

Infectious disease is a serious global health problem, and every year, it causes deaths of 13 million people worldwide, especially in developing countries like Indonesia (1, 2). WHO (World Health Organization) data show that viral, bacterial, fungal and parasitic infections are the biggest cause of death for the world's population. The use of antibiotics in the prevention of infectious diseases is the only solution (3). However, continuous use of antibiotics raises new problems for health, especially the resistance of microorganisms that causes infection (1, 2). Bacteria are the most abundant organisms and widespread in living things. In terms of distribution and number, most bacteria may cause disease in humans and other living things (3). *B. subtilis* and *E. coli* are the most common pathogenic bacteria in humans; *E. coli* is a Gram negative bacterium found in the large intestine of humans, and is a major cause of diarrheal diseases, especially in infants and children (1-3). The mortality rate from diarrhea in Indonesia is still around 7.4%, while the mortality rate due to persistent diarrhea is higher at 45%. In 1,000 inhabitants, 200-374 subjects experience diarrhea, with 60%-70% of whom are children under the age of 5 (4).

Bacterial resistance to antibiotics is a problem that has not been resolved until now (3, 5, 6). The *E. coli* resistants to chloramphenicol group of antibiotics and amoxicillin have long been reported (2, 3, 7, 8). The greater the percentage of bacterial resistance to an antibiotic state that bacteria are no longer susceptible to these antibiotics. Various studies to overcome the bacterial resistance were carried out, but no effective reports exist. Thus, investigating new antibacterial substances that are still active and selective is necessary. The search for antibiotic sources from natural ingredients is still the main trend for researchers. The ability of bioactive compounds of natural materials as a healing medium is estimated because of the content of secondary metabolites, including terpenoids, steroids, coumarin, flavonoids, and alkaloids (10-18).

One of the plants that has not been studied intensively in Indonesia is the family of Fabaceae (18, 19). The Fabaceae family demonstrates quite interesting bioactivity such as antibacterial (18, 19), antituberculosis (20), antioxidants (21), anticancer (22), and antimalarial (18, 23). The jengkol plant (*A. jiringa* (Jack) I. C. Nielsen), which belongs to the Fabaceae family, is commonly used by the Indonesian people as traditional medicine (18). Jengkol leaves are efficacious as medicine for eczema, scabies, sores, and ulcers, and the skin of the fruit is used as ulcerative medicine. Several studies were carried out on jengkol plants, including the leaves, fruit peel, and seeds (18, 19, 21, 24). The phytochemical screening has been done on jengkol fruit peel, seeds, bark, and leaves extracts (24). Based on this screening, from the parts of jengkol contain alkaloids, steroids, triterpenoids,

glycosides, saponins, flavonoids, and tannins. However, research on the stem roots of the jengkol plant was never performed.

In our ongoing investigation for new lead constituents from medicinal plants, we elaborated the bioactive secondary metabolites of *A. jiringa* stem roots and assayed their antibacterial activity. The aims of the study were to obtain antibacterial active fractions from the stem roots of *A. jiringa*, based on the bioassay-guided separation approach through their antibacterial property.

EXPERIMENTAL SECTION

Plant Materials

Samples of the stem roots of *A. jiringa* were assembled on January 25, 2018 from Unila's Housing area at Gedongmeneng District, Bandar Lampung, Lampung Province, Indonesia. The plant specimens (NV6/NRGD/2018) were identified at the Herbarium Bogoriense, LIPI Bogor, Indonesia.

General Experimental Procedures

TLC was performed on silica gel 60 GF₂₅₄ plates (Merck; 0.25 mm) and sprayed with staining reagent Ce(SO₄)₂. Column chromatography (CC) was made on silica gel (Kieselgel 60, 70-230 mesh ASTM; Merck) and Sephadex LH-20. ¹H NMR spectrum was measured in acetone-d₆ (TMS as an internal standard), on an NMR Agilent 500 MHz spectrophotometer (Agilent Technologies, JNM-ECZ500R/S1) or Bruker 500 MHz spectrometer. Finally, *Microplate Reader* Hospitex-Italy was used to measure the absorbance resulted on the resazurin assay, while UV spectra were performed using an Eppendorf BioSpectrometer kinetic instrument.

Bacterial Strains and Biochemicals

Ampicillin and chloramphenicol were purchased from Sigma Aldrich, and resazurin sodium salt was purchased from Sigma Aldrich. Gram-positive bacteria, *B. subtilis* ITBCCB148, was obtained from the Microbiology and Fermentation Technology Laboratory, Bandung Institute of Technology, Gram-negative bacteria, *E. coli* UNIATCC25922, nutrient agar broth, disposable sterile petri dishes (Idealcare), disposable syringe, micropipette tips, sterile tissue culture 96-well plates.

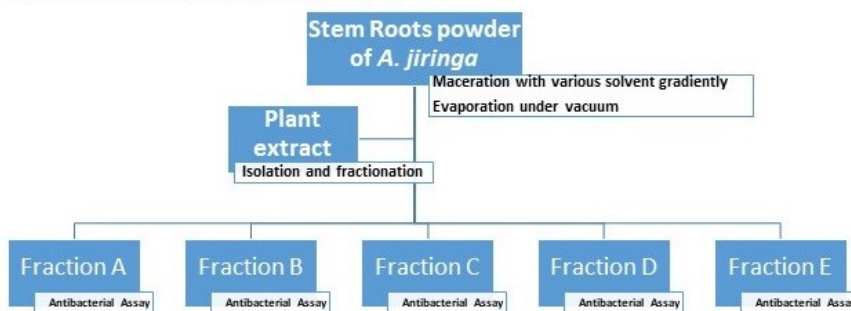
Preparation of Extracts

Fresh chopped stem roots of *A. jiringa* (2.5 kg) were cleaned by rinsing under running tap water to remove soil and dirt. The samples were dried in an open space for three weeks, and the air-dried roots are finally ground into a powder form. The powdered air-dried stem roots (1.5 kg) were extracted with a polar gradient polarity of solvent using the maceration technique. The solvent used at this stage starts from the solvent which exhibits the lowest polarity, *n*-hexane, followed by ethyl acetate (EtOAc), and finally with a high polarity organic solvent, methanol. Each extraction was conducted three times in each type of solvent. Before changing the type of solvent, the extract residue is first air

dried for at least three days or until the residue is dry/free of solvent. The filtrates obtained from maceration obtained from polar gradient solvents are then separated from the residue by ordinary filtration. The filtrates are then concentrated under reduced pressure using a vacuum rotary evaporator to yield *n*-hexane (3.6 kg), EtOAc (55.8 kg), and

methanol (67.2 kg) extracts, respectively. Using agar disc diffusion and microdilution methods, each extract obtained was subjected to an assay of its antibacterial activity. The most active extract was further isolated and fractionated through the bioassay-guided separation approach. The general research flowchart can be seen in Figure 1.

Extraction and Isolation Part



Bioassay Guided Fractionation Part

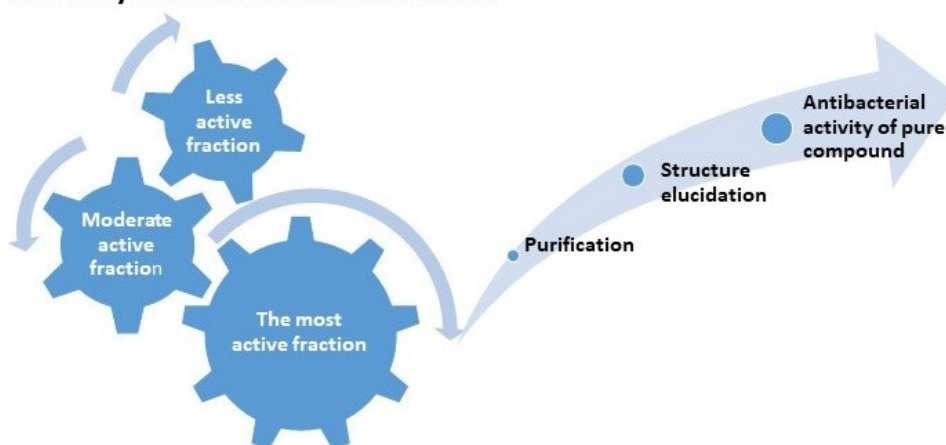


Figure 1. Research flow chart of bioassay guided separation.

Bioassay-guided Separation of Ethyl Acetate Extract

Based on the antibacterial activity results of extracts, the EtOAc extract (55.8 g) was selected to be isolated and fractionated further using the VLC method on silica gel (35-70 Mesh). The column was eluted with a stepwise gradient polarity of the solvent systems, including *n*-hexane-EtOAc (100%-0% of *n*-hexane), EtOAc-acetone (100%-0% of EtOAc), and acetone-MeOH (100%-0% of acetone), affording 23 fractions (200 mL each). According to their chemicals profile analyzed by TLC and their proton NMR spectrum, these fractions were grouped and combined into seven primary fractions, E₂1 (fr.1-10), E₂2 (fr.11), E₂3 (fr.12), E₂4 (fr.13), E₂5 (fr.14-15), E₂6 (fr.16-19), E₂7 (fr.20-23) (Figure 2). All fractions E₂1-E₂7 were tested for their antibacterial property separately using disc diffusion and microdilution methods. Among all fractions tested, three fractions exhibited antibacterial

activity against both bacterial strains with quite similar MIC's values, therefore fraction E₂2 (439.5 mg) was subjected to further fractionation due to the simplest chemical profile on its TLC. Fraction E₂2 was redissolved in acetone and then purified with CC on silica gel G-60 (35-70 Mesh) using *n*-hexane/isopropyl alcohol with the ratio volumes of 70/30, 60/40, 50/50, and 40/60, generating six major subfractions, E₂2a (132.0 mg), E₂2b (8.4 mg), E₂2c (9.0 mg), E₂2d (47.0 mg), E₂2e (2.0 mg), and E₂2f (50.3 mg). Using microdilution methods, all subfractions were screened against both bacterial strains tested. The MIC's values and optical density (OD) means of bioactive subfractions were described on Table 2. Among them, two subfractions (E₂2e and E₂2f) performed the most antibacterial activity against *B. subtilis* and *E. coli*, with the MIC's values ranging of 12.5-25 µg/mL. Only subfraction E₂2f was selected to be analyzed further by ¹H-NMR spectroscopy as exhibiting sufficient quantity.

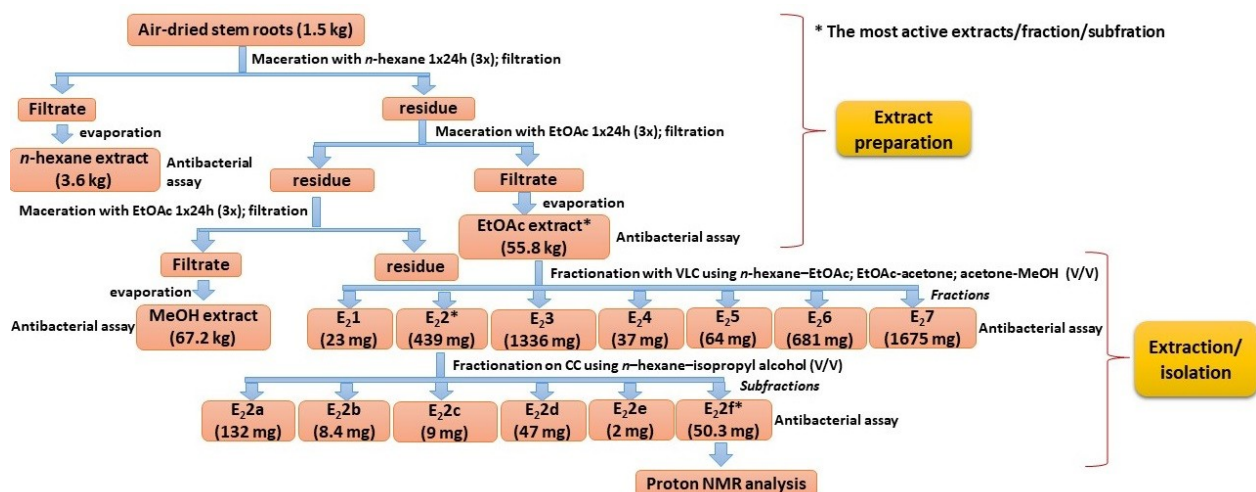


Figure 2. Scheme of bioassay guided separation of EtOAc extract subfractions.

Phytochemical Screening

The phytochemical screening was performed for triterpenes/steroids, alkaloids, flavonoids, and saponins, by using the standard procedures (25). The formation of precipitate or the color intensity was applied for analytical response of screening results.

Evaluation of Antibacterial Activity

For the testing antibacterial activity, the microbial strains employed in the biological assay are gram positive bacteria, *Bacillus subtilis*. Gram negative bacteria, *Escherichia coli*, have been obtained from a stock culture from Hospital Abdul Muluk, Bandar Lampung, Indonesia. To maintain stock culture, original cultures are further stored at a low temperature in the refrigerator. Fresh cultures are used for testing antibacterial activity using disc diffusion assay and dilution methods.

Disc diffusion assay

The antibacterial activity of the stem roots extract (*n*-hexane, ethyl acetate, and methanol) were tested by the disc diffusion method (26) against pathogenic bacteria gram negative (*E. coli*) and gram positive (*B. subtilis*). In this method, freshly prepared agar media is dispensed into the sterilized petri-dish. The agar is allowed to solidify, and 100 μ L of the bacterial suspension was poured over the agar media and spread by a spreader or a rod. Ampicillin and chloramphenicol (30 μ g/dish) is used as a positive control, while methanol is used as a negative control. In each culture medium petri-dish, four disks were used, one disk of antibiotics, two disks separately for (*n*-hexane, ethyl acetate, and methanol) extracts, and one disk used as a control (methanol). The plates are sealed and incubated overnight at 37 $^{\circ}$ C in the incubator. Next, antibacterial activity is assigned by measuring the inhibition zone formed around the discs, and the diameter of zone of inhibition (mean of three replicates SD) as indicated by clear area was measured to determine the antibacterial activity.

The experiment is replicated three times to confirm the reproducibility.

Determination of MIC via Resazurin Assay

Resazurin assay was carried out in 96-well plates titration with some minor modification (27). In the complete nutrient broth, two-fold dilutions of plant extracts and antibiotics were prepared in the test wells. The final concentration (20 μ L of each bacterial suspension) was added to 180 μ L of antibiotics and plant extracts (30–0.02 mg/mL in sequence) contained in the culture medium, as well as the antibiotics concentrations of 0.06 mg/mL and 0.12 mg/mL for ampicillin and chloramphenicol, respectively. For the comparative study, control plates were prepared only with the culture medium and bacterial suspension. The plates were sealed and incubated for 12 hours at 37 $^{\circ}$ C for an additional 5 hours. At 1-hour intervals, plates were observed for a blue to pink and pink to colorless color change in living bacterial strains containing wells. Preliminary micro titer plate assay reveals that the fast decolonization of resazurin extract does not exhibit antibacterial potential. The bioactivity of extracts was screened, which shows that the extracts inhibit the dye reduction.

RESULTS AND DISCUSSION

The plant material was macerated using gradient polarity of solvent and afforded *n*-hexane (3.6 kg), EtOAc (55.8 kg), and methanol (67.2 kg) extracts. EtOAc extract performed the most bioactive extract against two bacterial strains, *E. coli* and *B. subtilis*, using agar disc diffusion and microdilution methods. Using the diffusion method, extracts of *n*-hexane, EtOAc, and methanol obtained from the extraction stage were each screened for antibacterial activity tests against *E. coli* and *B. subtilis* bacteria. Bioactivity test results in bacteria *E. coli* and *B. subtilis* were observed based on the diameter of inhibition zones, as tabulated in Table 1.

Table 1. The results of inhibitory zone of the extracts (14 mm) against *E. coli* and *B. subtilis*.

Concentration	<i>E. coli</i>			<i>B. subtilis</i>		
	<i>n</i> -hexane	EtOAc	MeOH	<i>n</i> -hexane	EtOAc	MeOH
0.3 mg/disc	-	10	8	-	8	7
0.5 mg/disc	-	11	9	-	9	8

The results of the antibacterial screening test on the three extracts by diffusion method showed that EtOAc extract exhibited better bioactivity, compared to *n*-hexane and methanol extracts. Furthermore, the repetition of the antibacterial activity test carried out using a dilution method in order to

determine the minimum inhibitory concentration (MIC) of each fraction. The results of the MIC test of the three extracts using the dilution method against *E. coli* and *B. subtilis* bacteria can be seen in Figure 3, while the optical density (OD) values can be seen in Table 2.

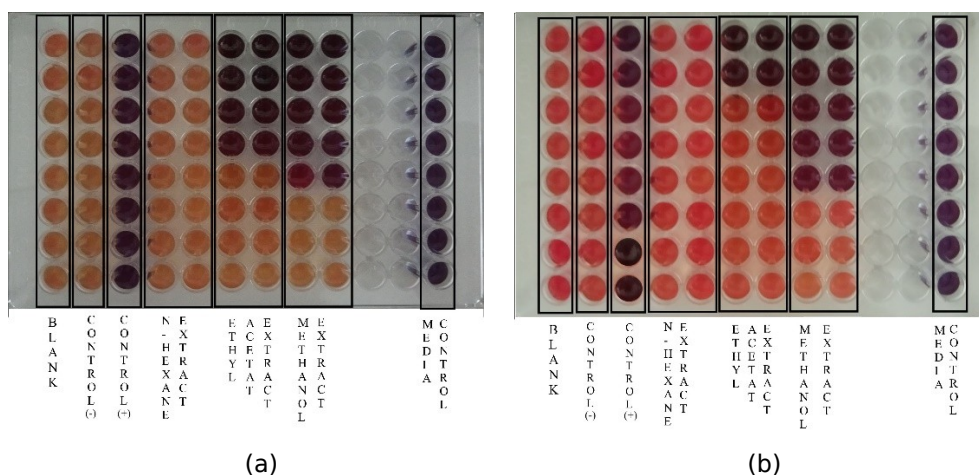


Figure 3. MIC testing of *n*-hexane, EtOAc, and methanol extracts using 96-well plates (a) on *E. coli* (b) bacteria on *B. subtilis* bacteria.

Table 2. The results of MIC test of *n*-hexane, EtOAc, and methanol extract using dilution method against *E. coli* and *B. Subtilis*.

No	Type of extract	Average of OD on <i>E. coli</i> *	Average of OD on <i>B. subtilis</i> *	MIC (µg/mL) on <i>E. coli</i>	MIC (µg/mL) on <i>B. subtilis</i>
1	<i>n</i> -hexane	0.76838	0.54231	-	-
2	EtOAc	1.88706	1.48556	12.5	50
3	Methanol	1.68644	1.66288	6.25	6.25
4	Positive control	1.773	1.48025	0.78	0.78
5	Negative control	0.74013	0.57575	-	-

The test results based on Table 2 show that semi-polar EtOAc extract and polar methanol demonstrate a minimum inhibitory concentration better than *n*-hexane extract. In this dilution antibacterial test, using visual observation is not sufficient to observe the presence or absence of bacterial growth; however, this tends to be subjective from each person's eyesight so the risk of

error is relatively greater. This occurs because the color test makes it difficult to observe, so absorbance values before and after incubation are used to help determine the presence or absence of bacteria. The wavelength used to measure the number of microbes is 600 nm, because cells in the mitochondria and cytoplasm absorb at that wavelength (28). After incubation for 18 hours and

measuring the OD, the addition of the reaction color was resazurin and was reincubated for 4 hours until the color changed. This color change occurred because resazurin exhibits a blue color that is not fluorescent, and it can be reduced to a fluorescent pink color. The change in color from blue to pink is an indicator of cell reduction. The color change of resazurin is carried out by enzymes in cells in the mitochondria and cytoplasm (27).

Based on the results of the antibacterial tests that were carried out, EtOAc extract was chosen for the fractionation and purification process because it exhibits the best antibacterial activity in inhibiting

bacterial growth to a minimum inhibitory concentration of 12.5 and 50 µg/mL, compared to extract *n*-hexane, which demonstrates no inhibition at all. The results of fractionation with the VLC process showed that a separation was present between semi-polar components and non-polar components, resulting in seven combined fractions, namely E₂1 (fr.1-10; 23 mg), E₂2 (fr.11; 439.5 mg), E₂3 (fr.12; 1,336 mg), E₂4 (fr.13), E₂5 (fr.14-15), E₂6 (fr.16-19), E₂7 (fr.20-23; 1,675 mg). All these fractions were screened for antibacterial activity using the agar diffusion method, and the results obtained were tabulated in Table 3.

Table 3. The diameter of inhibition zone (mm) of VLC-1 and VLC-2 fractions against *E.coli* and *B. subtilis*

Concentration	<i>E. coli</i>				<i>B. subtilis</i>			
	E ₂ 1	E ₂ 2	E ₂ 3	E ₂ 7	E ₂ 1	E ₂ 2	E ₂ 3	E ₂ 7
0.3 mg / disk	7	10	8	7	7	10	8	7
0.5 mg / disk	7	11	8	8	7	12	9	8

Using a dilution method to determine the MIC value, each antibacterial active fraction obtained above was then re-tested for bioactivity. The results of the

MIC test can be seen in Figure 4, and the average OD of each fraction can be seen in Table 4.

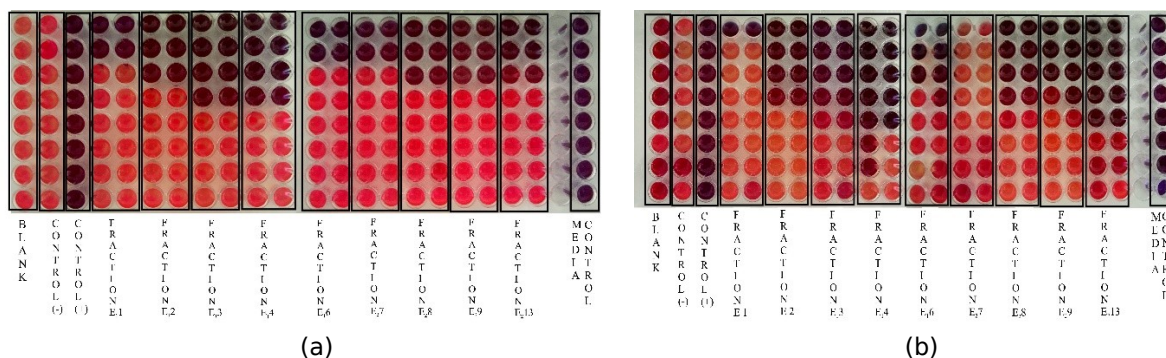


Figure 4. The MIC test on VLC fractions against: (a) *E. coli*; (b) *B. subtilis*.

Table 4. The result of MIC test of VLC fraction against *E. coli* and *B. subtilis* using dilution method.

No	Fraction code	OD average on <i>E. coli</i>	OD average on <i>B. subtilis</i>	MIC (µg/mL) on <i>E. coli</i>	MIC (µg/mL) on <i>B. subtilis</i>
1	E ₂ 1	1.16363	1.02819	50	-
2	E ₂ 2	1.49338	1.54488	25	25
3	E ₂ 3	1.25269	1.74956	25	25
4	E ₂ 7	1.39525	2.34931	25	6,25
5	Positive control	2.16563	2.28425	0,78	0,78
6	Negative control	0.86413	0.60838	-	-

From the test results shown in Table 4, it was found that the fractions of E₂, E₂3, and E₂7 gave a smaller value of the minimum inhibitory concentration compared to other fractions both against *E. coli* and *B. subtilis* bacteria. The E₂2 fraction (439.5 mg) was then selected for further purification by mass consideration. Phytochemical screening, especially flavonoid and phenolic tests, was carried out on the E₂8 fraction, which showed positive tests for the presence of flavonoids and phenolics. Purification of the E₂2 fraction (439.5 mg) produced six main subfractions, namely E₂2a (132.0 mg), E₂2b (8.4 mg), E₂2c (9.0 mg), E₂2d (47.0 mg), E₂2e (2.0 mg),

and E₂2f (50.3 mg). Then, the subfractions were carried out using MIC test 96-well plates, to determine the minimum inhibitory concentration of the subfraction. The results of MIC on *E. coli* bacteria are that the E₂2c, E₂2d, E₂2e, and E₂2f fractions exhibit a better minimum inhibitory concentration compared to other fractions. However, the MIC results in *B. subtilis* bacteria all fractions showed lower inhibitory ability than in *E. coli* bacteria. The results of the E₂2 subfraction MIC can be seen in Figure 5 and for the average OD of each fraction can be seen in Table 5.

Table 5. The result of MIC test on E₂8₁ subfraction using 96-well plates against *E. coli* and *B. subtilis*

No	Subfraction code	Average of OD on <i>E. coli</i>	Average of OD on <i>B. subtilis</i>	MIC (µg/mL) in <i>E. coli</i>	MIC (µg/mL) in <i>B. subtilis</i>
1	E ₂ 2a	1.49731	1.27825	50	100
2	E ₂ 2b	1.44425	1.22788	50	50
3	E ₂ 2c	1.29094	0.99919	25	50
4	E ₂ 2d	1.25900	0.84613	25	50
5	E ₂ 2e	1.39463	1.18294	12,5	25
6	E ₂ 2f	1.34638	1.14100	25	25
7	Positive control	1.54525	1.45938	0,78	0,78
8	Negative control	0.85238	0.56950	0	0

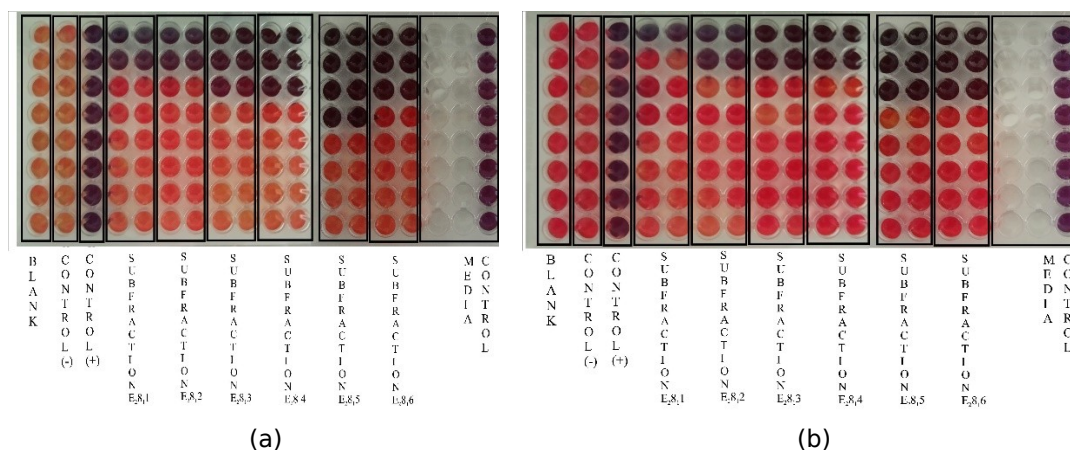


Figure 5. MIC test on E₂2 subfractions using 96-well plates against: (a) *E. coli*; (b) *B. subtilis*.

Based on the results obtained, a decrease in the minimum inhibitory concentration can be seen compared to the antibacterial activity test on the VLC results as observed on the purification results of the E₂2 fraction. This fact occurs due to the possible composition of active compounds contained in the sample. According to Priya *et al.* (29), when an increase or decrease in a component of an active compound occurs, a possibility of influencing the activeness of the compound itself exists. From the test results shown in Table 5, researchers found that

the E₂2e and E₂2f subfractions gave a smaller minimum inhibitory concentration value compared to other subfractions, both against *E. coli* and *B. subtilis* bacteria. Among them, two subfractions (E₂2e and E₂2f) performed the most antibacterial activity against *B. subtilis* and *E. coli*, with the MIC's values ranging from 12.5–25 µg/mL.

However, E₂2f was chosen to be further characterized because in terms of quantity more than E₂2e. As can be seen in Figure 6, subsequent

characterization of the E₂f subfraction was carried out using proton nuclear magnetic spectroscopy (¹H-NMR).

The spectrum shown in Figure 6 indicates that the E₂f subfraction is not pure. However, interpretation of the ¹H-NMR data can still be done, by referring to the results of the phytochemical test of the fraction which shows the content of phenolic compounds or flavonoids in the subfraction. Therefore, a comparison reporting the presence of phenolic or flavonoid compounds in the same plant type is sought in journals to estimate the types of bioactive compounds which are obtained. From the NMR spectrum of proton E₂f subfraction, the results show indications of aromatic protons in the chemical shift region δ_H 6–7. The signal for the methoxy group (-OCH₃) at δ_H 3.7, the proton signal at δ_H 8.01 with the peak of the chemical shift of the hydroxyl group (-OH) attached to an aromatic ring. As well as the typical peaks for alkanes at δ_H 0.89-2.2.

The isolation of phenolic compounds from the fruit skin of jengkol plants, and the researchers identified

them as 1-(2,6-dihydroxy-4-methoxyphenyl) decan-1-one (Figure 7) has previously been reported (30). Based on the comparison of the proton chemical shift values between the isolated subfraction and 1-(2,6-dihydroxy-4-methoxyphenyl) decan-1-one, researchers estimated that the main active compound contained in the E₂f subfraction is a phenolic compound, namely 1-(2, 6-dihydroxy-4-methoxyphenyl) decan-1-one. However, further purification in the E₂f subfraction still needs to be performed, to ensure the active compounds which are responsible for inhibition of the test bacteria. The results obtained in the spectrum exhibit similarities with the spectrum that was reported by others (30), as can be seen in Table 6. However, antibacterial studies have not yet been found against compound 1-(2,6-dihydroxy-4-methoxyphenyl) decan-1-one, so it is thought to be synergistic: between compounds 1-(2,6-dihydroxy-4-methoxyphenyl) decan-1-one with other compounds that cause antibacterial activity, as seen in the antibacterial test results in fractions of VLC results.

Table 6. Comparison of chemical shifts in ¹H-NMR subfraction E₂f with compounds 1-(2,6-dihydroxy-4-methoxyphenyl)decan-1-one.

No	Proton type	δ_H proton in Subfraction E ₂ f (ppm)	δ_H Proton in reference compound (30)
1	Aromatic	6 - 7	7.04
2	Methoxy	3.7	3.79
3	Hydroxy Aromatic	8.01	8.01
4	Alkane	0.89 -2.2	0.89-2.21

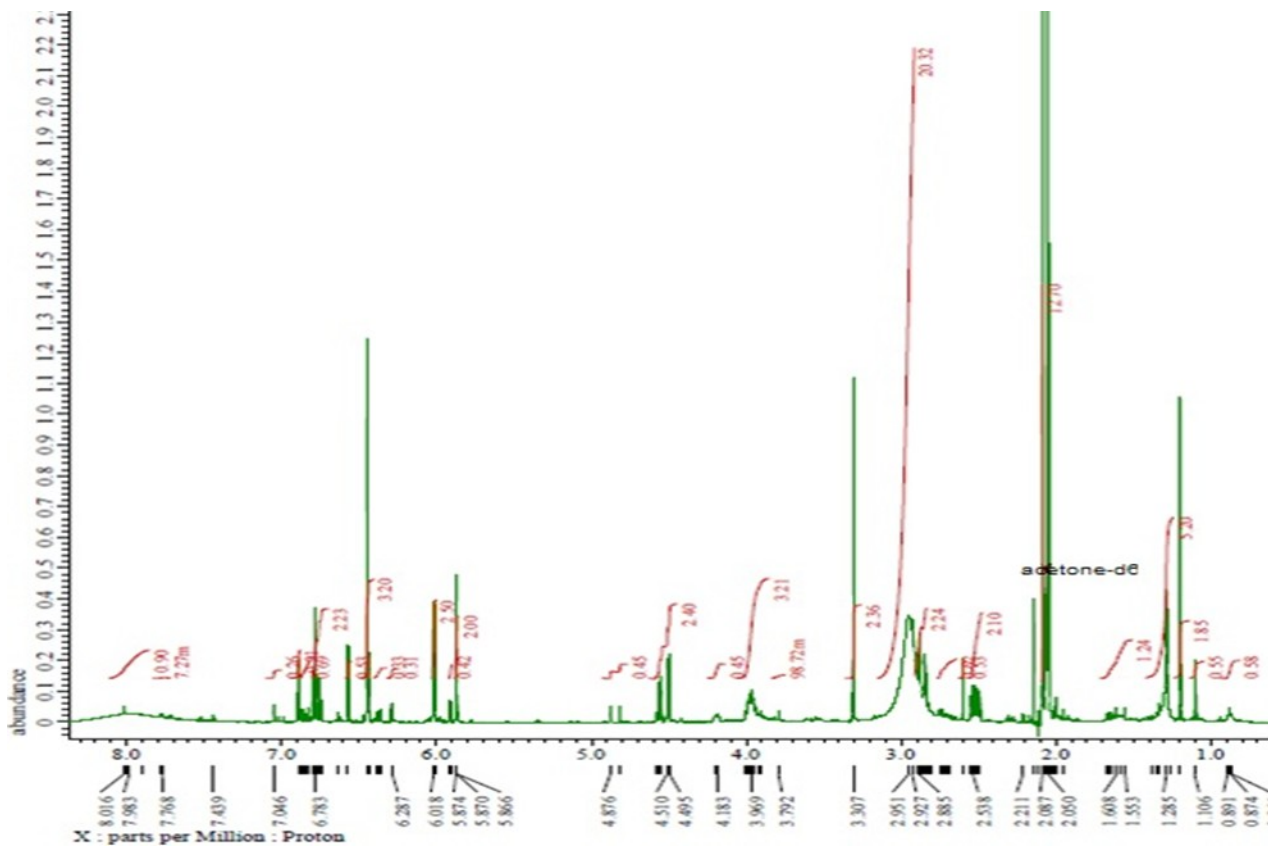
Based on the results of interpretation of ¹H-NMR, data seen in Table 7 indicate that the E₂f subfraction is not pure, but it is seen in the integration that indicates the possibility of the presence of three mixed compounds distributed in the chemical shift region δ_H 0.89–8.01 ppm. The first group exhibits chemical shifts that are distributed in

the area of δ_H 8.01, 6.8, 6.79, 6.76, 6.74, 6.20, 5.87, 4.58, 3.99, 2.53, and 1.67 ppm. The second group is distributed in the area of chemical shift δ_H 7.05, 6.82, 5.91, and 4.20 ppm. Whereas, the third group was distributed in the area of chemical shift δ_H 6.36, 6.29, and 0.89 ppm. The complete shift of proton chemistry and its integration can be seen in Table 7.

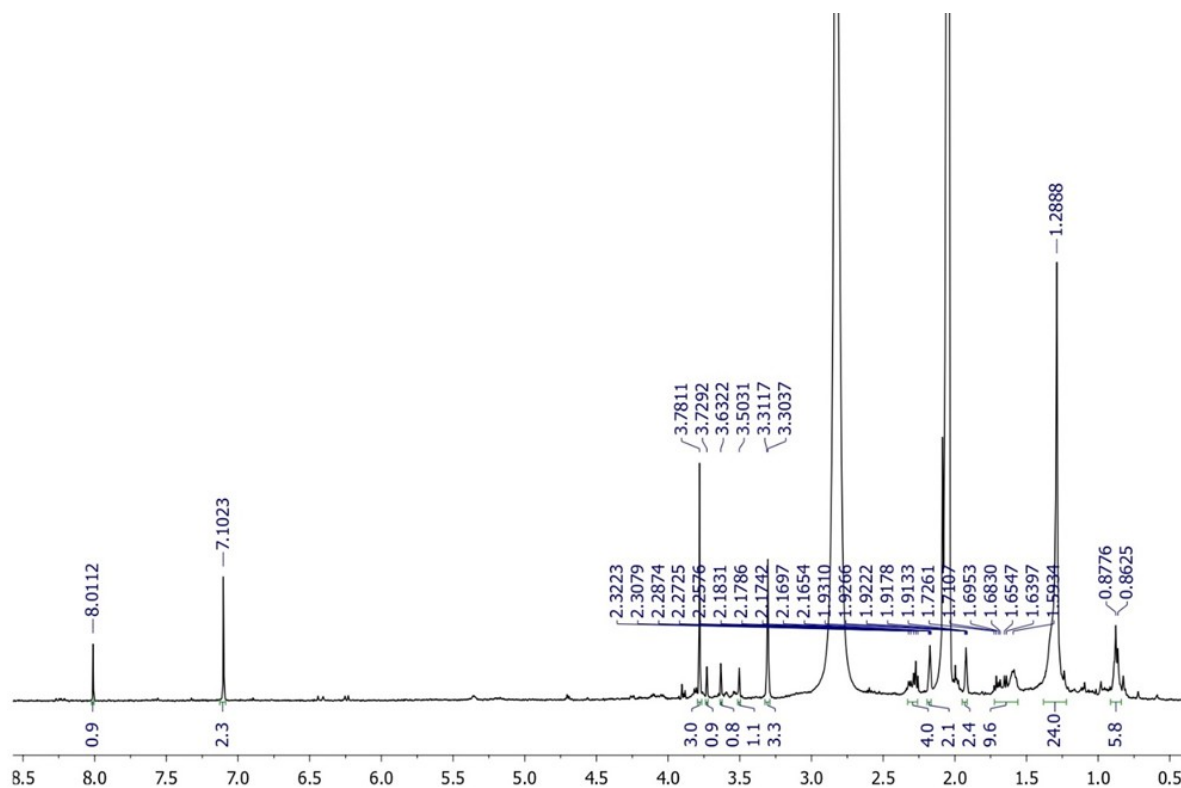
Table 7. Coupling constant and ¹H-NMR Integration of E₂f subfraction.

No	δ_H (ppm)	Multiplicity	Integration	J (Hz)*
1	8.01	<i>d</i>	0.90 ¹	16.5
2	7.05	<i>d</i>	0.26 ²	2.5
3	6.8	<i>d</i>	0.91 ¹	2.5
4	6.82	<i>d</i>	0.69 ²	1.5
5	6.79	<i>m</i>	1.23 ¹	4
6	6.76	<i>d</i>	2.23 ¹	1.5
7	6.74	<i>d</i>	2.23 ¹	1.5
8	6.36	<i>d</i>	0.33 ³	2.5
9	6.29	<i>d</i>	0.31 ³	2.5
10	6.20	<i>m</i>	2.50 ¹	2.5
9	5.91	<i>m</i>	0.42 ²	2.5
10	5.87	<i>m</i>	2.00 ¹	2
13	4.58	<i>m</i>	2.40 ¹	7.5
14	4.20	<i>t</i>	0.45 ²	4
15	3.70	<i>m</i>	3.21 ¹	3
16	2.53	<i>m</i>	2.10 ¹	3.5
17	1.67	<i>m</i>	1.24 ¹	5.5
18	0.89	<i>m</i>	0.58 ³	8.5

Superscripted 1, 2, and 3 are codes for different types of groups of compounds.



(a)



(b)

Figure 6. ¹H-NMR spectra of (a) E₂2f subfraction; (b) Reference compound (30).

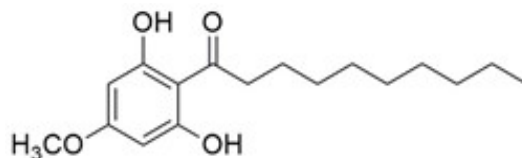


Figure 7. Structure of compound 1- (2,6-dihydroxy-4-methoxyphenyl) decan-1-one.

CONCLUSIONS

The EtOAc extract of jengkol (*A. jiringa* (Jack) I. C. Nielsen) root bark exhibits antibacterial activity against *E. coli* and *B. subtilis* bacteria, which is better than the *n*-hexane extract. Compared to other fractions from the same column, the E13 fraction and E28 fraction exhibit better antibacterial activity against *E. coli* and *B. subtilis* bacteria. The E13 fraction and E28 fraction exhibit better antibacterial activity against *E. coli* and *B. subtilis*, compared to the E13 subfraction and E28 fraction as a result of its purification. Based on the $^1\text{H-NMR}$ result data, the compounds detected in the E₂₈ subfraction are a phenolic compound type. Lastly, this study is the first reported from this plant and the potential exists to further investigate the invention of new promising antibacterial agent from plants.

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CONFLICTS OF INTEREST

The authors declare no conflicts of interest.

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