

KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
BADAN LAYANAN UMUM UNIVERSITAS LAMPUNG

SP2D

SURAT PERINTAH PENCAIRAN DANA (SP2D)

Dari Bendahara Pengeluaran
Tanggal 02 Agustus 2019
Nomor Q1877/UN26/SP2D/2019
Tahun Anggaran 2019

SIGNATURE

PM 00354/LS/LP2M/2019
31 Juli 2019
Lembaga Penelitian dan Pengabdian kepada Masyarakat UNILA
Belanja 5742002001057 A 5251 10

Belanja Penyediaan Barang dan Jasa BLU Lainnya

Bank BNI 46 Cabang Pembantu UNILA
h memindahbukukan dari rekening nomor

019.3922.784

Uang sebesar	Rp.	52.500.000,00
PPH	Rp.	0,00
PPN	Rp.	0,00
Jumlah diterima	Rp.	52.500.000,00

Lima Puluh Dua Juta Lima Ratus Ribu Rupiah

Drs Suratman Umar, M Sc., Griya GM Indah Blok C2/12A RT 004 RW 002 Gedung Meneng Ra
Bandar Lampung

14.590.651.7-323.000

0443717626 (Bpk Suratman)

BNI Cabang Tanjung Karang

Pembayaran Tahap I 70% untuk Kegiatan Penelitian Kerjasama Internasional Unila sesuai dengan SPK nomor 3093/UN26.21/PN/2019 tgl 10 Juli 2019, BAST nomor 3123/UN26.21/PN/2019 tgl 11 Juli 2019 BAP nomor 3183/UN26.21/PN/2019 tgl 12 Juli 2019

LOGI DAN...
A.S. Peng...
sa...
Rektor...
Umum dan Keuangan,



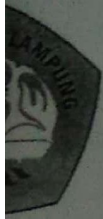
Bandar Lampung tanggal seperti di
Bendahara Pengeluaran UNILA

Muhammad Ismail
NIP. 198907062010121011

imad Kamil
6101011985031003

Bendahara Pengeluaran 2. Biru untuk BPP 3. Hijau untuk Bank

UNIVERSITAS LAMPUNG
REKTORAT
JANSEK DARI 18022784
JANSEK H 443 26 Bpk SURATMAN IDR 52.500.000,00-
UN26/SP2D/19 02082019
REKTOR UNIVERSITAS LAMPUNG IDR 52.500.000,00-



KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
UNIVERSITAS LAMPUNG
LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT
GedungRektoratLantai 5, Jalan Prof. Dr. SumantriBrojonegoro No. 1 Bandar Lampung 3514
Telepon (0721) 705173, Fax. (0721) 773798, e-mail : lppm@kpa.unila.ac.id
www.lppm.unila.ac.id

**SURAT PERJANJIAN (KONTRAK) PEKERJAAN
PELAKSANAAN KEGIATAN PENELITIAN KERJASAMA INTERNASIONAL**

NOMOR : 3093/UN26.21/PN/2019
TANGGAL : 10 Juli 2019

ANTARA

PEJABAT PEMBUAT KOMITMEN
LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT
UNIVERSITAS LAMPUNG

DAN

Drs. Suratman Umar, M.Sc (Ketua)
PENANGGUNGJAWAB KEGIATAN PENELITIAN DENGAN JUDUL
Investigation Of Antibacterial Bioactive Compounds From Jengkol Plant (
Pithecellobium lobatum) Through Bioassay Guided Fractination Approach

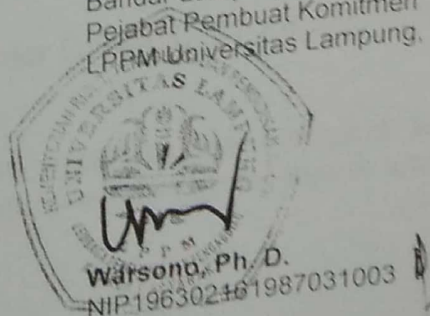
FAKULTAS MIPA
UNIVERSITAS LAMPUNG
BANDAR LAMPUNG
2019

RINGKASAN KONTRAK

Kegiatan yang dananya berasal dari DIPA BLU Universitas Lampung

1. No./Tgl.DIPA : DIPA-042.01.2.400954/2019, 05 Desember 2018
2. Kode Keg./Sub.Keg/MAK : 5742.002.001.057.A.525119, Tahun Anggaran 2019 (Penelitian)
3. No. dan Tanggal SPK : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019
4. Nama Penanggungjawab : Drs. Suratman Umar, M.Sc / Penanggung Jawab Kegiatan Penelitian KERJASAMA INTERNASIONAL Unila
5. Alamat Penanggungjawab : Jl.Prof. Sumantri Brojonegoro No.1 Bandar Lampung
6. Nomor Pokok Wajib Pajak : 14.590.651.7-323.000
7. Nilai SPK/Surat Perjanjian : Rp 75.000.000,-
8. Uraian dan volume Pekerjaan : Penelitian dengan Judul "Investigation Of Antibacterial Bioactive Compounds From Jengkol Plant (Pithecellobium lobatum) Through Bioassay Guided Fractination Approach"
9. Cara Pembayaran :
 1. Kegiatan penelitian pembayaran angsuran I (satu) sebesar 70% (dari nilai pekerjaan) atau 70% x Rp 75.000.000,- yakni sebesar Rp 52.500.000,- (Lima puluh dua juta lima ratus ribu), setelah surat perjanjian pelaksanaan pekerjaan ini ditandatangani oleh kedua belah pihak dan menyerahkan proposal-proposal kegiatan tersebut dari Pihak Kedua kepada Pihak Pertama
 2. Kegiatan penelitian pembayaran angsuran II (dua) sebesar 30% (dari nilai pekerjaan) atau 30% x Rp 75.000.000,- yakni sebesar Rp 22.500.000,- (Dua puluh dua juta lima ratus ribu), setelah pekerjaan selesai 100% dinyatakan dengan Berita Acara Serah Terima pekerjaan dan menyerahkan laporan hasil kegiatan dari Pihak Kedua kepada Pihak Pertama.
 3. Pembayaran tersebut di atas dilakukan melalui kas Badan Layanan Umum (BLU) ke Rekening Pihak Kedua pada Bank : **BNI Tanjung Karang** dengan nomor rekening 0443717626, a.n. **Drs. Suratman Umar, M.Sc** sebagai penanggung jawab kegiatan penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung.
10. Jangka waktu pelaksanaan : 119 (Seratus Sembilan Belas Hari) kalender terhitung tanggal 10 Juli – 5 November 2019
11. Tanggal Penyelesaian Pekerjaan : 5 November 2019
12. Jangka waktu Pemeliharaan :
Ketentuan Sanksi :
 1. Apabila terjadi ketelambatan pekerjaan tanpa adanya alasan yang diterima oleh pemberi pekerjaan dikenakan sanksi/denda sebesar 1/1000 (satu permil) untuk setiap hari keterlambatan denga denda maksimal sebesar 5%, (lima persen) dari jumlah harga borongan.
 2. Segala resiko yang timbul akibat keterlambatan pekerjaan tersebut ini sepenuhnya menjadi beban dan tanggung jawab pihak II. Maka kami sebagai pihak I dapat membatalkan SPK secara sepihak dan pihak II tidak berhak menuntut kerugian apapun dari instansi kami.

Bandar Lampung, 10 Juli 2019
Pejabat Pembuat Komitmen
LPPM Universitas Lampung,


Warsono, Ph. D.
NIP196302461987031003

SURAT PERJANJIAN (KONTRAK) PEKERJAAN
ELAKSANAAN KEGIATAN PENELITIAN KERJASAMA INTERNASIONAL
UNIVERSITAS LAMPUNG

NOMOR : 3093/UN26.21/PN/2019
TANGGAL : 10 Juli 2019

da hari ini **Rabu** tanggal **Sepuluh** bulan **Juli** tahun **Dua Ribu Sembilan Belas**, kami
ng bertanda tangan di bawah ini :

Nama : Warsono, Ph. D.
Jabatan : Pejabat Pembuat Komitmen LPPM Universitas Lampung
Alamat : Jl. Prof. Soemantri Brojonegoro No.1 Bandar Lampung

anjutnya dalam perjanjian ini disebut **PIHAK PERTAMA**

Nama : Drs. Suratman Umar, M.Sc
Jabatan : Penanggungjawab Pelaksanaan Kegiatan Penelitian KERJASAMA
INTERNASIONAL
dengan Judul "*Investigation Of Antibacterial Bioactive
Compounds From Jengkol Plant (Pithecellobium lobatum)
Through Bioassay Guided Fractination Approach*".
Alamat : Jl. Prof. Soemantri Brojonegoro No.1 Bandar Lampung

anjutnya dalam perjanjian ini disebut **PIHAK KEDUA**

IAK PERTAMA DAN KEDUA berdasarkan :

Peraturan Presiden nomor 54 tahun 2010; tentang pengadaan barang/jasa pemerintah
Undang-undang RI nomor 17 tahun 2003 tentang Keuangan Negara;
Undang-undang nomor 20 tahun 2003 tentang Sistem Pendidikan Nasional;
Undang-undang nomor 15 tahun 2004 tentang Pemeriksaan Pengelolaan dan Tanggung
Jawab Keuangan Negara;
Keppres Nomor 42 tahun 2002 nomor 72 tahun 2004 tentang Pelaksanaan Anggaran
Pendapatan dan Belanja Negara;
Peraturan Menteri Keuangan Nomor 606/KMK.66/2004 tentang Pedoman Pembayaran
Pelaksanaan Anggaran;
DIPA Universitas Lampung Nomor DIPA-042.01.2.400954/2019, tanggal 05 Desember
2018

ngan ini menyatakan setuju dan sepakat untuk mengikat diri dalam suatu perjanjian
aksanaan pekerjaan, dengan ketentuan dan syarat-syarat tercantum dalam pasal-pasal

PASAL 4 JANGKA WAKTU PELAKSANAAN

Jangka waktu pelaksanaan kegiatan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung tersebut dalam pasal 1 adalah 119 (Seratus Sembilan Belas Hari) terhitung sejak ditandatanganinya perjanjian ini. Laporan ini harus diserahkan **PIHAK KEDUA** selambat-lambatnya tanggal 5 November 2019 sebanyak (3) Tiga Eksemplar.

Apabila laporan Penelitian tidak diselesaikan tepat pada waktunya, **PIHAK KEDUA** dapat mengajukan Adendum sebanyak 1 kali saja, dan apabila **PIHAK KEDUA** berhenti/diberhentikan dari jabatan atau dipindahkan ke instansi lain, **PIHAK KEDUA** wajib mempertanggungjawabkan penggunaan dana penelitian yang telah diterima dari **PIHAK PERTAMA**, selanjutnya **PIHAK PERTAMA** berhak menunjuk orang lain untuk melaksanakan pekerjaan tersebut.

PASAL 5 SANKSI

Jika **PIHAK KEDUA** tidak dapat melaksanakan pekerjaan sesuai dengan batas Waktu pelaksanaan yang tercantum dalam pasal 4 dalam perjanjian ini maka untuk tiap hari keterlambatan **PIHAK KEDUA** wajib membayar denda keterlambatan sebesar 1/1000 (satu permil) dari nilai kontrak.

PIHAK KEDUA bertanggung jawab penuh apabila dalam pelaksanaan pekerjaan ini tidak sesuai dengan ketentuan yang berlaku, atau terdapat hal – hal atau temuan pemeriksaan yang mengakibatkan kerugian negara.

PASAL 6 PENYELESAIAN PERSELISIHAN

Jika terjadi perselisihan antara kedua belah pihak, pada dasarnya akan diselesaikan secara musyawarah.

Jika perselisihan itu tidak dapat diselesaikan secara musyawarah, maka akan diselesaikan oleh "panitia pendamai" yang berfungsi sebagai juri/wasit yang dibentuk dan diangkat oleh kedua belah pihak yang terdiri dari:

- Seorang wakil dari **PIHAK PERTAMA** sebagai anggota
- Seorang wakil dari **PIHAK KEDUA** sebagai anggota
- Seorang pihak ketiga yang ahli sebagai Ketua, yang telah disetujui oleh **PIHAK KEDUA**

Keputusan panitia pendamai ini mengikat kedua belah pihak, dan biaya penyelesaian perselisihan yang dikeluarkan akan ditanggung secara bersama.

Jika keputusan ini sebagaimana dimaksud ayat 3 pasal ini tidak dapat diterima oleh salah satu pihak, maka penyelesaian perselisihan akan diteruskan melalui pengadilan Negeri.

**PASAL 7
LAIN-LAIN**

Segala sesuatu yang belum diatur dalam surat perjanjian ini yang dipandang perlu oleh kedua belah pihak akan diatur lebih lanjut dalam surat perjanjian tambahan (*Addendum*) dan merupakan perjanjian yang tidak dapat terpisahkan dari perjanjian ini.

Surat perjanjian ini dibuat rangkap 4 (empat) untuk Pihak Pertama dan Pihak Kedua, selebihnya diberikan kepada pihak-pihak yang berkepentingan dan ada hubungannya dengan pekerjaan.

**PASAL 8
PENUTUP**

Surat perjanjian ini dibuat dan ditandatangani oleh kedua belah pihak di atas materai Rp.6.000.- (enam ribu rupiah) pada lembar ke satu dan lembar kedua yang mempunyai kekuatan hukum sama.

Perjanjian ini berlaku mulai tanggal ditandatangani oleh kedua belah pihak.

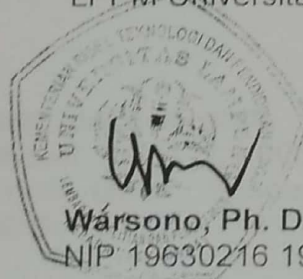
PIHAK KEDUA
nanggungjawab Kegiatan,



1192898

3. Suratman Umar, M.Sc
P 196406041990031002

PIHAK PERTAMA
Pejabat Pembuat Komitmen
LPPM Universitas Lampung,



Warsono, Ph. D.
NIP 19630216 198703 1 003



KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
UNIVERSITAS LAMPUNG

LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT

GedungRektoratLantai 5, Jalan Prof. Dr. SumantriBrojonegoro No. 1 Bandar Lampung 35145

Telepon (0721) 705173, Fax. (0721) 773798, e-mail : lppm@kpa.unila.ac.id

www.lppm.unila.ac.id

=====

SURAT PERNYATAAN TANGGUNGJAWAB MUTLAK

ng bertanda tangan di bawah ini :

a m a : Drs. Suratman Umar, M.Sc
P : 196406041990031002
abatan : Penanggungjawab Kegiatan Penelitian **KERJASAMA
INTERNASIONAL**

nyatakan dengan sesungguhnya :

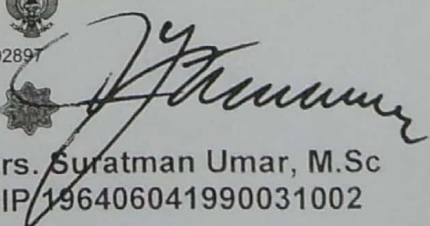
1. Perhitungan tahap I sebesar 70% yang terdapat pada kegiatan Penelitian **KERJASAMA INTERNASIONAL** Unila sebesar Rp 52.500.000,- (Lima puluh dua juta lima ratus ribu) telah dihitung dengan benar. Sesuai kontrak Nomor : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019.
2. Apabila dikemudian hari terdapat kelebihan atas pembayaran kegiatan Penelitian **KERJASAMA INTERNASIONAL** Unila, kami bersedia menyetorkan kelebihan tersebut ke Kas Negara.
3. Segala akibat yang mungkin timbul dari perubahan di atas menjadi tanggungjawab kami sepenuhnya.

Demikian pernyataan ini kami buat dengan sebenar-benarnya.

Bandar Lampung, 10 Juli 2019

:gungjawab Kegiatan,




Drs. Suratman Umar, M.Sc
NIP/196406041990031002

KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
UNIVERSITAS LAMPUNG

LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT

GedungRektoratLantai 5, Jalan Prof. Dr. SumantriBrojonegoro No. 1 Bandar Lampung 35145

Telepon (0721) 705173, Fax. (0721) 773798, e-mail : lppm@kpa.unila.ac.id

www.lppm.unila.ac.id

SURAT PERNYATAAN TANGGUNGJAWAB MUTLAK

ng bertanda tangan di bawah ini :

m a : Warsono, Ph. D.

P : 196302161987031003

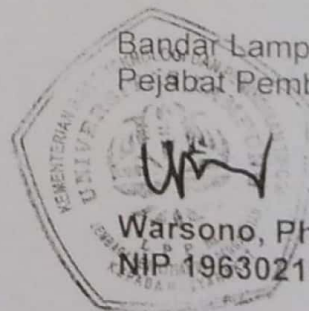
atan : Pejabat Pembuat Komitmen LPPM Universitas Lampung

nyatakan dengan sesungguhnya :

1. Perhitungan tahap I sebesar 70% yang terdapat pada kegiatan Penelitian **KERJASAMA INTERNASIONAL** Unila sebesar Rp 52.500.000,- (Lima puluh dua juta lima ratus ribu) telah dihitung dengan benar. Sesuai kontrak Nomor : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019.
2. Apabila dikemudian hari terdapat kelebihan atas pembayaran kegiatan Penelitian **KERJASAMA INTERNASIONAL** Unila, kami bersedia menyetorkan kelebihan tersebut ke Kas Negara.
3. Segala akibat yang mungkin timbul dari perubahan di atas menjadi tanggungjawab kami sepenuhnya.

Demikian pernyataan ini kami buat dengan sebenar-benarnya.

Bandar Lampung, 10 Juli 2019
Pejabat Pembuat Komitmen,



Warsono, Ph. D.

NIP 196302161987031003



KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
UNIVERSITAS LAMPUNG

LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT

Gedung Rektorat Lantai 5, Jalan Prof. Dr. Sumantri Brojonegoro No. 1 Bandar Lampung 3514

Telepon (0721) 705173, Fax. (0721) 773798, e-mail : lppm@kpa.unila.ac.id

www.lppm.unila.ac.id

Nomor : 3093/UN26.21/PN/2019
Lampiran : 1 (satu) berkas
Perihal : Pengajuan SPP dan SPM

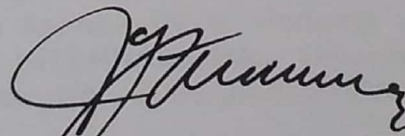
10 Juli 2019

Kepada Yth.
Wakil Rektor II
Universitas Lampung
Di Bandar Lampung

Dengan ini kami sampaikan permohonan penerbitan SPP dan SPM untuk keperluan pembayaran tahap I sebesar 70% Kegiatan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung sebesar Rp 52.500.000,- (Lima puluh dua juta lima ratus ribu) yang dilaksanakan sesuai dengan Surat Perjanjian nomor : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019.

Atas perhatian dan kerjasamanya diucapkan terima kasih.

Penanggungjawab Kegiatan,



Drs. Suratman Umar, M.Sc
NIP 196406041990031002

BERITA ACARA SERAH TERIMA PEKERJAAN
Nomor : 3123/UN26.21/PN/2019

Pada hari **Kamis** tanggal **Sebelas** bulan **Juli** tahun **Dua Ribu Sembilan Belas**, kami sampaikan yang bertanda tangan di bawah ini :

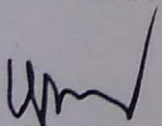
1. Nama : Warsono, Ph. D.
Jabatan : Pejabat Pembuat Komitmen Lembaga Penelitian Dan Pengabdian Kepada Masyarakat Universitas Lampung
Alamat : Jl. Sumantri Brojonegoro No.1 Bandar Lampung 35145
Selanjutnya disebut sebagai Pihak Pertama
2. Nama : Drs. Suratman Umar, M.Sc
Jabatan : Penanggung Jawab Kegiatan Penelitian KERJASAMA INTERNASIONAL
Alamat : Jl. Sumantri Brojonegoro No.1 Bandar Lampung 35145
Selanjutnya disebut sebagai Pihak Kedua

Dengan ini menyatakan telah dilaksanakan serah terima pertama pekerjaan pelaksanaan kegiatan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung setelah Surat Perjanjian Pelaksanaan Pekerjaan di tandatangi oleh kedua belah pihak dengan pembayaran dilaksanakan dalam dua angsuran yaitu pertama sebesar 70% dan kedua 30% dari nilai pekerjaan, sebagai berikut :

1. **Pihak Pertama** telah menerima dari **Pihak Kedua** proposal pelaksanaan kegiatan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung sesuai dengan Surat Perjanjian Pelaksanaan Pekerjaan No : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019
2. **Pihak Kedua** telah menyerahkan kepada Pihak Pertama proposal pelaksanaan kegiatan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung sesuai dengan Surat Perjanjian Pelaksanaan No : 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019.

Demikian berita acara serah terima pekerjaan ini dibuat untuk dapat dipergunakan sebagaimana mestinya.

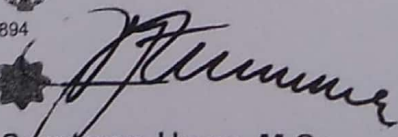
PIHAK PERTAMA
PEJABAT PEMBUAT KOMITMEN
LPPM UNIVERSITAS LAMPUNG,



Warsono, Ph. D.
NIP 196302161987031003

PIHAK KEDUA
PENANGGUNG JAWAB KEGIATAN,




Drs. Suratman Umar, M.Sc.
NIP 196406041990031002

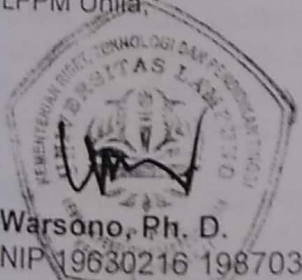
TAHUN ANGGARAN : 2019
DIPA NOMOR/TANGGAL : 042.01.2.400954/2019
MAK : Tanggal 05 Desember 2018.
: 5742.002.001.057.A 525119


KWITANSI

SUDAH DITERIMA DARI : Pejabat Pembuat Komitmen LPPM Universitas Lampung
BANYAKNYA UANG : *Lima puluh dua juta lima ratus ribu*
UNTUK PEMBAYARAN : Tahap I Pekerjaan Penelitian **KERJASAMA INTERNASIONAL** Universitas Lampung Tahun 2019 sesuai dengan Surat Perjanjian No. 3093/UN26.21/PN/2019, Tanggal 10 Juli 2019, dan BAP No. 3183/UN26.21/PN/2019, Tanggal 12 Juli 2019
JUMLAH : Rp 52.500.000 -

Setuju dibayar
Pejabat Pembuat Komitmen
LPPM Unila,

Bandar Lampung, 12 Juli 2019
Yang Menerima
Penanggungjawab
Kegiatan Penelitian,


Warsono, Ph. D.
NIP 19630216 198703 1 003


6000
Drs. Suratman Umar, M.Sc
NIP 198406041990031002

KEMENTERIAN RISET, TEKNOLOGI DAN PENDIDIKAN TINGGI
UNIVERSITAS LAMPUNG

LEMBAGA PENELITIAN DAN PENGABDIAN KEPADA MASYARAKAT

Gedung Rektorat Lantai 5, Jalan Prof. Dr. Sumantri Brojonegoro No. 1 Bandar Lampung 35145

Telepon (0721) 705173, Fax. (0721) 773798, e-mail : lppm@kpa.unila.ac.id

www.lppm.unila.ac.id

BERITA ACARA PEMBAYARAN

Nomor : 3183/UN26.21/PN/2019

Pada hari **Jum'at** tanggal **Dua belas** bulan **Juli** tahun **Dua Ribu Sembilan Belas**, kami yang bertanda tangan di bawah ini :

1. Nama : Warsono, Ph. D.
Jabatan : Pejabat Pembuat Komitmen LPPM Unila
Alamat : Jl. Sumantri Brojonegoro No.1 Bandar Lampung
DISEBUT SEBAGAI PIHAK PERTAMA
2. Nama : Drs. Suratman Umar, M.Sc
Jabatan : Penanggungjawab Kegiatan Penelitian **KERJASAMA INTERNASIONAL**
Alamat : Jl. Sumantri Brojonegoro No.1 Bandar Lampung
DISEBUT SEBAGAI PIHAK KEDUA

Dengan ini telah melaksanakan Kegiatan Penelitian **KERJASAMA INTERNASIONAL** Unila, sesuai dengan Surat Perjanjian Nomor : 3184/UN26.21/PN/2019, Tanggal 12 Juli 2019

PIHAK KEDUA berhak menerima pembayaran dari **PIHAK PERTAMA** sebesar 70% dari nilai kontrak atau $70\% \times \text{Rp } 75.000.000,- = \text{Rp } 52.500.000,-$ (Lima puluh dua juta lima ratus ribu) yang digunakan untuk 1 kegiatan penelitian, melalui kas Badan Layanan Umum Universitas Lampung.

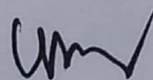
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Bandar Lampung, 12 Juli 2019

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Berdasarkan Surat Keputusan Nomor : 1688 /UN26.21/PP/2019 tanggal 21 Juni 2019 dan perjanjian kontrak Nomor : 3093/UN26.21/PN/2019, tanggal 10 Juli 2019 mendapatkan Anggaran Penelitian dengan judul "*Investigation Of Antibacterial Bioactive Compounds From Jengkol Plant (Pithecellobium lobatum) Through Bioassay Guided Fractination Approach*"

Dengan ini menyatakan bahwa :

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No	Uraian	Jumlah
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Drs. Suratman Umar, M.Sc
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RT 004 RW 002 GEDONG M. NENGI

RAJABASA

PANITAR LAMPUNG

TERDAFTAR

19-01-2021



Contoh Tanda Tangan

Bangetuhan Matigas Bank

TANJUNG KARANG

No. D 5883641

Kantor Cabang
No. Rekening
Nama

0443717626 - IDR
BPK : SURATHAN

142 - 28691

NO. D 5883641

1. Penarikan BNI Taplus dapat dilakukan di semua Teller BNI di dalam negeri, di BNI ATM, ATM LINK, ATM Bersama dan ATM Prima di seluruh Indonesia serta ATM berlogo Cirrus atau Maestro di seluruh dunia.
2. BNI Taplus dilengkapi dengan fasilitas Kartu Debit BNI dan BNI e-Banking yang memudahkan Anda dalam melakukan transaksi perbankan.

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2. Jika buku BNI Taplus/Kartu Debit BNI hilang, agar segera memberitahu Bank.
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5. Penarikan di Teller harus menyertakan buku BNI Taplus dan asli identitas diri yang berlaku. Penarikan dengan nominal tertentu di Teller yang rekeningnya memiliki fasilitas Kartu Debit BNI harus menyertakan Kartu Debit BNI.
6. Perubahan data nasabah agar dilaporkan kepada Bank.
7. Rekening yang tidak bertransaksi selama 6 bulan berturut-turut akan dimatikan pada idomandi. Rekening dormant dapat aktif kembali dengan transaksi predebitan/peng kreditan melalui fasilitas BNI e-Banking, penarikan tunai, penyetoran tunai, pemindahbukan melalui Kantor Cabang BNI atas pembelaan di cabang.
8. Rekening dormant yang tidak aktifkan dan bersaldo nihil dalam jangka waktu tertentu akan otomatis ditutup oleh sistem.
9. Penggantian buku BNI Taplus yang habis karena mutasi harus dapat dilakukan di seluruh Kantor Cabang BNI di dalam negeri.
10. Penggantian buku BNI Taplus karena hilang yang memiliki fasilitas Kartu Debit dapat dilakukan di seluruh Kantor Cabang BNI di dalam negeri.
11. Penggantian Kartu Debit BNI karena hilang/muda berlaku jatuh tempo harus dapat dilakukan di seluruh Kantor Cabang BNI di dalam negeri.

Keterangan Kode Transaksi:

1. Penyetoran
2. Penarikan
3. Kliring
4. Pemindahbukan
5. Pajak
6. Bunga
7. Pembayaran Kasalahan
8. Pemindahan Saldo
9. Biaya Administrasi
10. Akun Transaksi Di

11. Akun Transaksi Cr
13. Irigasi Debit Phone Banking
22. Penarikan ATM
23. Transaksi Kredit Phone Banking
24. Transaksi Maestro/POS
25. Transaksi Tarik Cirrus/Link/Bersama/Prima
26. Biaya Inquiry Cirrus/Link/Bersama/Prima
27. Pembayaran-Pembayaran
28. Biaya Tarik Cirrus/Link/Bersama/Prima
22. Pemindahan melalui ATM

88. Biaya Penolakan Transaksi Cirrus karena dana tidak cukup
81. Biaya Penolakan Transaksi Cirrus akibat hal hal lain
91. Reversal/Pembatalan ATM
92. Reversal/Pembatalan Maestro/POS
94. Reversal/Pembatalan Tarik Cirrus/Link/Bersama/Prima
95. Reversal/Pembatalan Biaya Inquiry Cirrus/Link/Bersama/Prima
98. Reversal/Pembatalan Pembayaran
97. Reversal/Pembatalan Biaya Tarik Cirrus/Link/Bersama/Prima
90. Reversal/Pembatalan Biaya Tarik Cirrus/Link/Bersama/Prima

**FINAL REPORT
PENELITIAN KERJASAMA INTERNASIONAL (PKSI)
UNIVERSITAS LAMPUNG**



**INVESTIGATION OF ANTIBACTERIAL BIOACTIVE
COMPOUNDS FROM JENGKOL PLANT (*Archidendron jiringa*
(Jack) I. C. Nielsen) THROUGH *BIOASSAY GUIDED
FRACTIONATION* APPROACH**

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COLLABORATOR:

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Funded by:

**DIPA BLU Universitas Lampung
No. 3093/UN26.21/PN/2019, Date: 10 July 2019**

**FAKULTAS MATEMATIKA DAN ILMU PENGETAHUAN ALAM
UNIVERSITAS LAMPUNG
NOVEMBER 2019**

HALAMAN PENGESAHAN
PENELITIAN KERJASAMA INTERNASIONAL UNIVERSITAS LAMPUNG

Judul Penelitian : Investigation of Antibacterial Bioactive Compounds from Jengkol Plant (*Archidendron jiringa* (Jack) I. C. Nielsen) Through *Bioassay Guided Fractination* Approach

Manfaat Sosial Ekonomi : Membantu Meningkatkan Kesehatan Masyarakat

Jenis Penelitian : Penelitian Terapan

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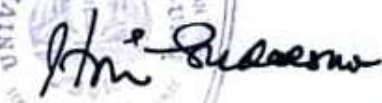
Lokasi Kegiatan : Laboratorium Kimia Organik dan LTSIT Unila

Lama Kegiatan : 8 bulan

Biaya penelitian : Rp. 75.000.000,-

Sumber Dana : DIPA BLU Universitas Lampung

Bandar Lampung, 24 Oktober 2019


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SUMMARY

Infectious diseases caused by bacteria has become the global health issues especially antibacterial drug resistance. The most serious concern with antibacterial resistance is that some bacteria have become resistant to almost all antibacterial drugs and making them less effective. Overusing or misusing of these drugs can make resistance develop even faster. Various efforts have been made by many researchers to find antibacterial drugs including through chemical constituent studies from natural plants. Plants are the one of the most important natural resources because they are relatively safe from the side effects and cheap. One of the potential plants that has antibacterial properties is the jengkol plant (*Archidendron jiringa* (Jack) I. C. Nielsen). Jengkol plant is the one of Lampung's natural resources that has not been optimally used, and its utilization is in line with Unila's superior research focus in the field of technology development and drug health. In this research, a chemical constituents study of roots and stem barks of jengkol plants have been carried out through several stages including isolation, purification, identification, and biological activity evaluation. The stages of isolation of secondary metabolites were carried out by extraction technique followed by fractionation using chromatographic methods such as vacuum liquid chromatography (VLC), and column chromatography (CC). Meanwhile, characterization and identification of the active fraction was carried out by proton Nuclear Magnetic Resonance (¹H-NMR). The pure compounds will be tested for their biological activity against grams positive *Bacillus subtilis* and negative bacteria *Escherichia coli* using the agar disc diffusion and dilution method. The research outputs obtained has been prepared as a draft manuscript that ready to submit to international reputable scientific journals such as the Brazillian Journal of Pharmacognosy (International, Elsevier, IF: 1.754, Q2) or Pharmaceutical Biology (International, Q2 Scimago, Scopus indexed).

Keywords: Jengkol plant; *Archidendron jiringa*; antibacterial; bioassay guided fractionation; grams positive and negative bacteria

PRAKATA

Bismillah,

Alhamdulillah puji dan syukur penulis ucapkan ke hadirat Alloh Subhanahu wata'ala atas segala rahmat dan karunia-Nya laporan kemajuan penelitian ini dapat diselesaikan dengan baik sesuai dengan doa dan harapan. Laporan kemajuan dengan tema penelitian:” Investigation of Antibacterial Bioactive Compounds From Jengkol Plant (*Archidendron jiringa* (Jack) I. C. Nielsen) Through *Bioassay Guided Fractination* Approach” ini merupakan salah satu persyaratan yang harus dibuat dan menjadi bagian dari rangkaian kegiatan penelitian secara keseluruhan.

Dalam pelaksanaan kegiatan penelitian dan penulisan laporan kemajuan ini tidak lepas dari berbagai kesulitan dan rintangan, namun itu semua dapat penulis lalui berkat rahmat dan pertolongan-Nya serta bantuan dan dorongan semangat dari berbagai pihak yang membantu baik secara langsung maupun tidak langsung.

Pada kesempatan ini, penulis menyampaikan terimakasih sedalam-dalamnya kepada:

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Penulis sangat menyadari bahwa laporan ini masih jauh dari kesempurnaan, namun semoga hasil penelitian ini dapat memberikan manfaat yang banyak kepada para pembaca khususnya baik penulis pribadi. Aamiin.

Bandar Lampung, 28 Oktober 2019

Peneliti

Dr. Noviany, M.Si

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I. INTRODUCTION

1.1 Research Background

Infection disease is a serious global health problem and its caused the deaths of 13 million people worldwide every year, especially in developing countries like Indonesia. WHO data shows that viral, bacterial, fungal and parasitic infections are the biggest cause of death for the world's population. The use of antibiotics in the prevention of infectious diseases is the only solution¹. While the continuous use of antibiotics raises the new problems for health especially the resistance of microorganisms that cause infection.

Bacteria are the most abundant organisms and widespread in living things. In terms of distribution and number, most bacteria may cause disease in humans and other living things². *Bacillus subtilis* and *Escherichia coli* are the common pathogenic bacteria in humans, *E. coli* is a gram-negative bacterium found in the large intestine of humans, and it is a major cause of diarrheal diseases, especially in infants and children^{3,4}. The mortality rate from diarrhea in Indonesia is still around 7.4%, while the mortality rate due to persistent diarrhea is higher at 45%. In 1000 inhabitants, 200-374 subjects experience diarrhea with 60-70% of whom are children under the age of 5 years⁵.

Bacterial resistance to antibiotics is a problem that has not been resolved until now. Refdanita et al.⁶ found that *E. coli* was resistant to chloramphenicol group antibiotics by 83.9% and amoxicillin by 86.2%. The greater the percentage of bacterial resistance to an antibiotic state that bacteria are no longer susceptible to these antibiotics. Various studies to overcome the bacterial resistance have been carried out, but there have been no effective reports. Thus, it is necessary to look for new antibacterial substances that are still active and selective. The search for antibiotic sources from natural ingredients is still the main trend for researchers. The ability of bioactive compounds of natural materials as a healing medium is estimated because of the content of secondary metabolites, including terpenoids, steroids, coumarin, flavonoids, and alkaloids.

One of the plants that has not been studied intensively in Indonesia is the family of Fabaceae. The Fabaceae family has quite interesting bioactivity such as antioxidants, antimalarial, anticancer, and antibacterial. The jengkol plant

(*Archidendron jiringa* (Jack) I. C. Nielsen) which belongs to the family Fabaceae is commonly used by the Indonesian people as traditional medicine. Jengkol leaves are efficacious as medicine for eczema, scabies, sores, and ulcers, the skin of the fruit is used as ulcer medicine¹. Several studies have been carried out on jengkol plants, both the leaves, fruit peel, and seeds. Nurussakinah (2010)⁷ has carried out the phytochemical screening on jengkol fruit peel, seeds, bark and leaves extracts. Based on these screening its contain alkaloids, steroids, triterpenoids, glycosides, saponins, flavonoids, and tannins. While research on the roots and bark of the jengkol plant has never been done.

Based on above explanation, it is necessary to do chemical research on the stem bark of the jengkol plant (*A. jiringa*) by isolating and identifying secondary metabolites, followed by testing the antibacterial activity against *Staphylococcus aureus*, *Escherichia coli*, and *Pseudomonas aurogenosa*. Bioassay-guided approach through their antibacterial activity will be used to isolate the bioactive compounds.

1.2 Objectives of Research

This study will be conducted through 3 stages: extraction, isolation / fractionation / purification based on antibacterial bioassays-guided approach. Antibacterial testing shall be carried out using a dilution technique using gram positive and negative bacteria. The specific objective of the study is to obtain antibacterial active fractions/compounds from the roots and bark of *A. jiringa* which are described in detail in the research method.

1.3 Importance of Research

The utilization of Lampung's local natural resources is one of the leading research focuses by the University of Lampung (Unila) in the field of technology development and medicinal health. The antibiotic resistance is the major issues and its encourages researchers to study the antibacterial potential of available Lampung natural products which is not been studied and utilized optimally. For example, jengkol plants was chose to be studied because the chemical composition from this plant have potential to explored as a natural antibacterial substance. This finding can be used for preclinical screening *in vitro* / *in vivo* for the discovery of new antibiotic materials. It also reflexes the Unila's strategic plan in the prevention of infectious and tropical diseases caused by bacterial.

The theme proposed in this project is relevant to the 2016-2020 Unila research strategic plan, and in line with the superior field road map outlined in the theme research roadmap: Superior Material Based on Natural Resource Conservation, output/expected product is a superior biological-based material product and local natural resource-based organic active product⁸.

Based on this strategic plan and road map of researcher (Fig.1), the jengkol plant which grows in Lampung will be used as a source of natural ingredients that is very potential to be developed. The results of the research obtained are expected to increase the economic value of jengkol plants and help improve the welfare and health of the community through research that is synergized with other fields such as medicine, pharmacy, and management in a sustainable manner. Thus, this research is very urgent to be carried out to support the strategic plan achievements and the 2016-2020 Unila research road map.

II. LITERATURE REVIEW

Indonesia's biodiversity, especially on high and low levels of plants, places Indonesia as the third 'megadiversity' country after Brazil and China. This wealth makes research and exploration of new sources of bioactive compounds from plants will be more practical and beneficial especially for researchers in Indonesia. Because until now, plants still play an important role in the discovery of new drugs to overcome various human diseases⁹. According to Tulp et al.¹⁰, it is estimated that nearly 50% of the circulating drugs are now sourced from plants. Therefore, interest in the source of bioactive compounds can help make the new drug industry more efficient.

Exploration of new bioactive compounds from Indonesian plants, has become one of the prospective and profitable research focuses, especially for the pharmaceutical industries. Because until now, plants still play an important role in the discovery of new drugs to overcome various diseases. The Leguminosae or Fabaceae plant family, especially the species in the Papilionoideae subfamily has long attracted the attention of researchers not only due to variations in the structure of the compounds produced, but also because of their interesting biological activity. Several types of secondary metabolite compounds such as alkaloids, flavonoids, coumarins, phenylpropanoids,

anthraquinones, terpenoids and cyanogenate glycosides have been successfully isolated from this plant family¹¹. Among the various groups of compounds, the type of isoflavonoids is the main component found in the subfamily Papilionoideae.

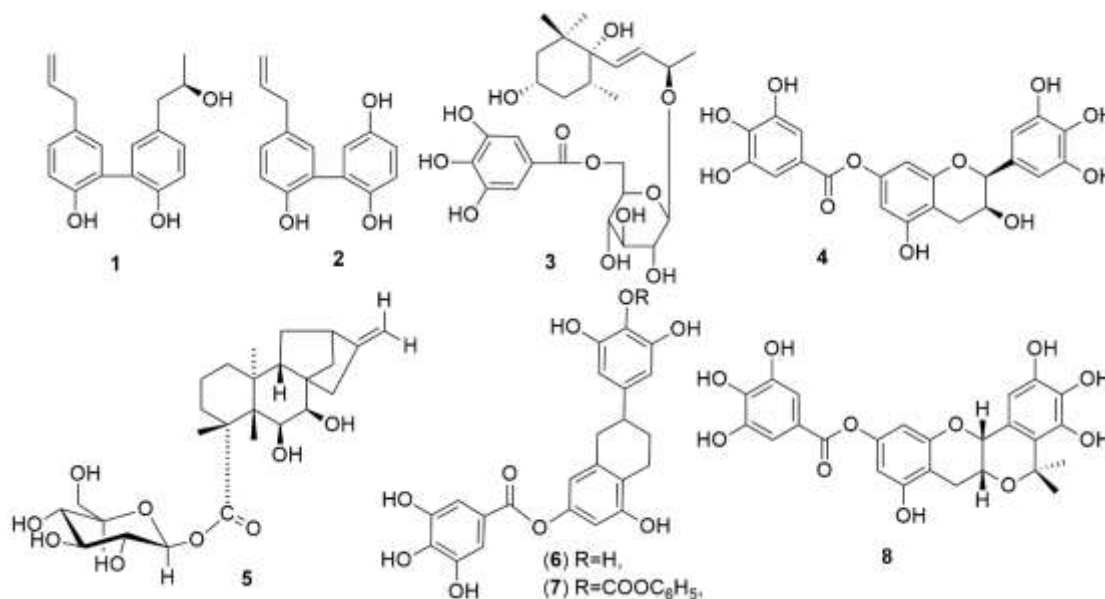
In 2007, Veitch¹² reported that more than 420 new compounds of isoflavonoid were successfully isolated from this plant. In line with these findings, it turns out that in nature, this group of compounds has a broad distribution of biological functions such as antimicrobials, insect repellents, allelopathic substances¹³, inhibitors of pathogenic/disease attacks¹⁴. In addition, the results of clinical studies of isoflavonoid compounds showed a positive effect on human health and nutrition including in the prevention of heart disease, menopausal symptoms and osteoporosis¹⁵⁻¹⁸.

Among 13,000 species in subfamily Papilionoideae, *A. jiringa* is one of the most potent medicinal plants to be developed as a new source of bioactive compounds. Several chemical studies have been reported from different species, including Lou et al.¹⁹, who have isolated two new 3,3'-neolignan compounds, namely clypearianine A (**1**) dan randaiol (**2**) from the twigs and leaves of *Pithecellobium clypearia* which exhibited antioxidant activity. Other researchers reported that a new clypearioside (**3**) along with a stereoisomer, (-)-(2S,3S)-epigallocatecin-7-gallate (**4**) has been obtained from the water leaves and stem bark extracts of the same plant²⁰.

In 2008, a new compound was identified as (-) 19- β -D-glucopyranosyl-6,7-dihydroxikaurenoate (**5**) from *P. albicans* seeds²¹. In addition, Li et al²² have successfully isolated two antiviral flavan derivatives that were identified as 7-O-galloyltricetifavan (**6**) and 7,4'-di-O-galloyltricetifavan (**7**) from the methanol extract of *P. clypearia* leaves. Furthermore, compound (**8**) has been obtained from the twigs and leaves of the same plant as well which showed to inhibit the expression of proinflammatory cytokines IL-6 or MCP-1 induced by influenza H1N1 virus in human A549 lung carcinoma cells²³. According to the literature search, the most previous study that has been done were carried out on the seeds as well as the leaves of plants, while the root and stem bark had not been done intensively. Therefore, in this study, the chemical investigation will be conducted on the root and stem bark of *A. jiringa* extract and the antibacterial activity will be evaluated.

Based on the research roadmap of the researcher (Fig.1) which is in line with Unila's strategic plan, exploration of the secondary metabolites content from the

Leguminosae family plants through isolation and characterization techniques has been carried out since the last decade. From previous studies that have been done on the roots of turi plants (*Sesbania grandiflora*), obtained one terpenoid, betulin acid, seven compounds isoflavonoids, xenognosin B, liquiritigenin, 7,2',4'-trihydroxyisoflavone, demethylvestitol, vestitol, medicarpin, sativan, together with one new natural phenolic compound, 1,1'-bi-2-naphthol^{24,25}. All isolated compounds were tested for their activity in vitro against strains of *M. tuberculosis* bacteria. Another recent study on the part of *S. grandiflora* bark has been isolated from two new phenolic compounds, sesbagrandiflorain A and B which also show anti-TB bioactivity in the medium category²⁶. Based on the results of these studies, it can be stated that *S. grandiflora* is a valuable traditional medicinal plant especially in dealing with diseases caused by bacterial infections.



Jengkol plant (*A. jiringa*) have close kinship with turi plant, both of which are included in the Leguminosae family. Based on the chemotaxonomy of the plant, jengkol plant is predicted containing the same bioactive components as turi plant. This fact is also supported by the traditional knowledge of Indonesian in particular Lampungese people who have used some parts of the jengkol plant as traditional medicine. Therefore, the study of jengkol plant is urgent to be done to find alternative sources of other bioactive compounds and clarify other potential of these plants as a

useful source for the discovery of new types of drugs, especially those that have the potential as antibiotics.

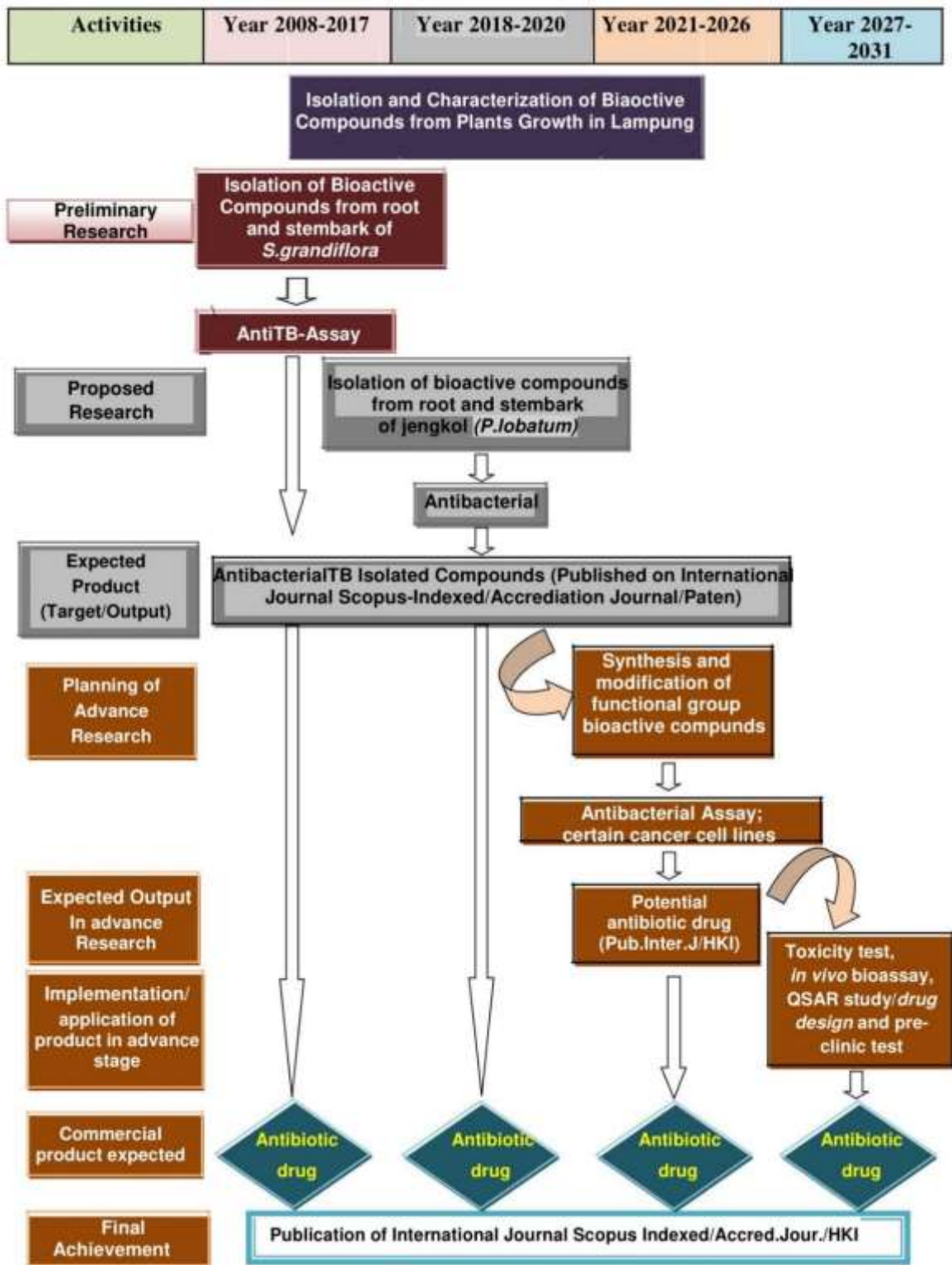


Figure 1. The Road Map of Research

In this study, isolation and identification of bioactive compounds from the root bark and stem bark of *A. jiringa* were carried out. Bioactive compounds were obtained through the bioassay guided fractionation approach which relies on antibacterial tests in vitro against gram positive and negative bacteria. Pure compounds that show strong antibacterial activity will be used for the next stage of research such as in vivo and pre-clinical trials with collaborators from various other disciplines.

III. RESEARCH METHODOLOGY

This research will be carried out for approximately 7 months, starts from March to September 2019 in Organic and LTSIT Laboratories University of Lampung and NMR instrument facility at Universiti Sains Malaysia. The stem roots of *A. jiringa* were collected in March 2019 in Gedongmeneng, Bandar Lampung, Indonesia. The identity of the plant specimen was authenticated and deposited at the Bogoriense Herbarium, Bogor, Indonesia.

3.1 Instruments, Chemicals, and Solvents

The laboratory equipments and instruments will be used in this study, including laboratory glasswares, rotary evaporator, thin layer chromatography (TLC), vacuum liquid chromatography (VLC), analytical balance, Eppendorf tubes, melting point apparatus, UV lamp, NMR, and UV-Visible spectrophotometers.

The commercial chemicals and reagents used in the isolation and characterization of all the isolated compounds were as follows: acetone, acetonitrile, benzene, cerium (IV) sulphate tetrahydrat, chloroform, dichloromethane, diethyl ether, dimethyl sulfoxide, ethanol, ethyl acetate, *n*-hexane, hydrochloric acid, isopropylalcohol, methanol, sephadex LH-20 silica gel G 60, silica gel 60 GF₂₅₄, silica gel 60 PF₂₅₄ containing gypsum, TLC aluminium sheets, and toluene.

3.2 Antibiotics, Bacterial Strains, and Consumables

Chloramphenicol, amoxicillin; gram positive bacteria: *Bacillus subtilis* and gram negative bacteria: *Escherichia coli*; nutrient agar broth, Resazurin; disposable sterile petri dishes, disposable syringe, micropipette tips, tissue culture plates 96-well.

3.3 Experimental Procedures

A. Preparation of extracts

Fresh chopped roots/stembarks (2.5 kg/each part of plant) were cleaned by rinsing under running tap water to remove soil and dirt. The roots was dried in an open space for three weeks and the air-dried roots are finally ground into powder form. The powdered air-dried roots (1.5 kg) were extracted with polar gradient solvent maceration technique. The solvent used at this stage starts from the solvent which has the lowest polarity, *n*-hexane, followed by ethylacetate and ends with a high polarity organic solvent, methanol. Each extraction was going carried out three times in each type of solvent. Before changing the type of solvent, the extract residue was first dried air for at least three days or until the residue is dry / free of solvent. The filtrates obtained from maceration obtained from polar gradient solvents were then separated from the residue by ordinary filtration. The filtrates were then concentrated under reduced pressure using a vacuum rotary evaporator. The process of isolation and purification have been carried out to concentrated methanol extract obtained.

B. Isolation and purification

Isolation and purification of methanol extract from roots/stembarks of jengkol plant was done in the usual way, through several stages of chromatographic separation which includes vacuum liquid chromatography and column chromatography using various gradient organic solvents polarity such as *n*-hexane, chloroform, dichloromethane, ethylacetate, acetone, isopropyl alcohol and methanol, either in the form of a mixture with a certain ratio or without a solvent mixture. The active extract was then purified by column chromatography using a suitable solvent. The biactive fraction was determined by determination of NMR spectroscopy and by thin layer chromatography (TLC) using eluent variations.

C. Structure Elucidation

Determination of the predictions of bioactive fraction has been determined spectroscopically using NMR spectroscopic analysis.

3.4 Screening of the Antibacterial activity

For the testing antibacterial activity, the microbial strains employed in the biological assay are Gram positive bacteria: *Bacillus subtilis* and gram negative bacteria: *Escherichia coli* (obtained from stock culture Hospital Abdul Muluk, Bandar Lampung, Indonesia). Original culture were further stored at low temperature in the

refrigerator to maintain stock culture. Fresh cultures were used for testing antibacterial activity using disc diffusion assay method.

A. Disc diffusion assay

The antibacterial activity of the stem roots extract (*n*-hexane, ethyl acetate and methanol) have been tested by disc diffusion method²⁷ against pathogenic bacteria gram negative (*Escherichia coli*) and gram positive (*Bacillus subtilis*). In this method, freshly prepared agar media was dispensed into the sterilized petri-dish. The agar was allowed to solidify and 100µl of bacterial suspension poured over the agar media and spread by a spreader or a rod. Ampicillin and chloramfenicol (30µg/dish) were used as positive control, while methanol was used as a negative control. In the each culture medium petri-dish four dish were used, one disk of antibiotics, two discs separately for (*n*-hexane, ethyl acetate and methanol) extracts, one disk used as a control (methanol). The plates were sealed and incubated overnight at 37°C in the incubator. Antibacterial activity was assigned by measuring the inhibition zone formed around the discs. The diameter of zone of inhibition (mean of three replicates SD) as indicated by clear area was measured to determine the antibacterial activity. The experiment was replicated three times to confirm the reproducible.

B. Determination of MIC via Resazurin assay

Resazurin assay has been performed in 96 well titration plates with some modification²⁸. In complete nutrient broth two fold dilutions of plant extracts and antibiotics were prepared in the test wells. The final concentration were 20µ of each bacterial suspension was added to 180 µl of antibiotics and plant extracts (30-0.02mg/ml in sequence) contained in culture medium as well as the antibiotics concentration will be 0.06 mg/l ampicillin and chloramfenicol 0.12mg/l. For comparative study control plates were prepared only with culture medium and bacterial suspension. The plates were sealed and incubated for 12 hours at 37 C for additional 5 hour. At the intervals of 1 hour plates were observed for colour change blue to pink and pink to colorless in live bacterial strains containing wells. Preliminary micro titre plate assay revealed that the fast decolonization of resazurin extract didn't possessed antibacterial potential. The bioactivity of extracts were screened which showed that the extracts inhibit the dye reduction. The flow chart of the research can be seen in Fig.2.

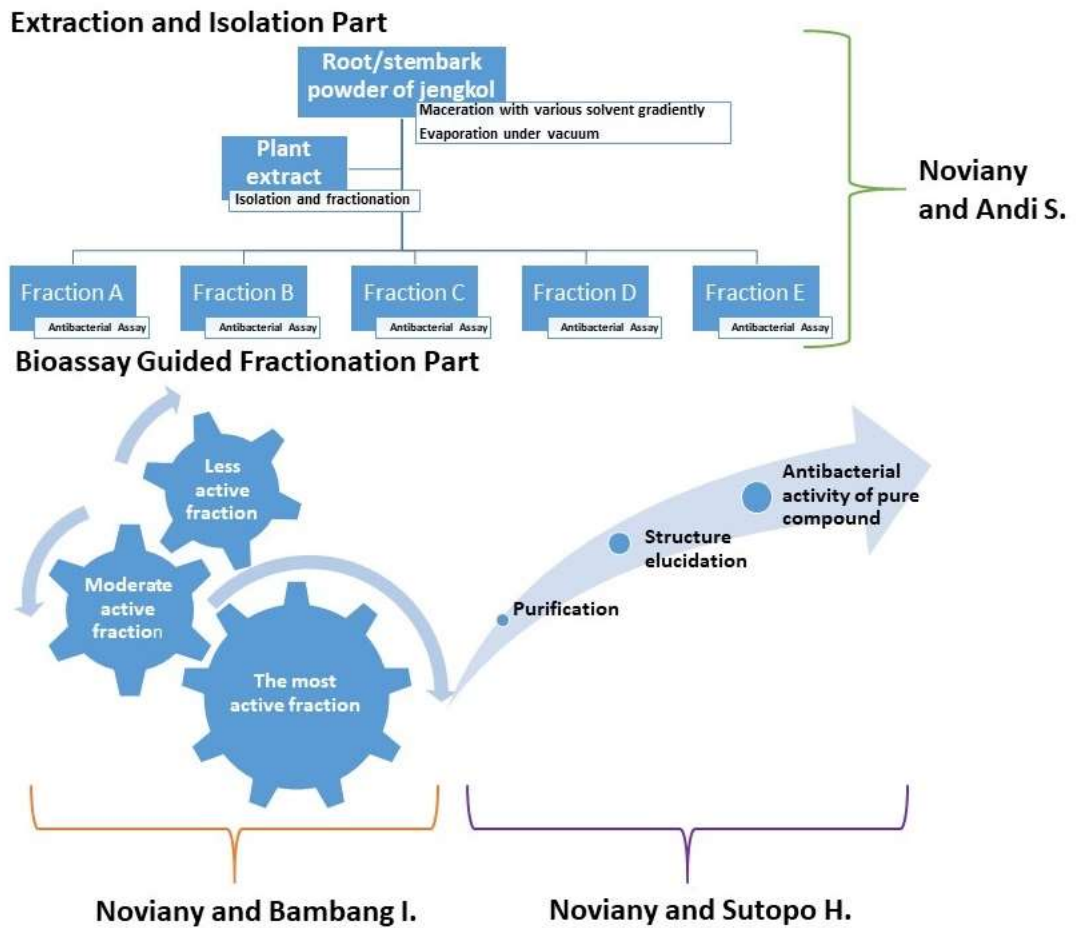


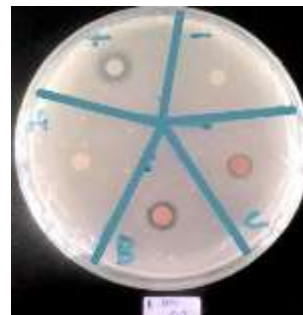
Figure 2. The Flow Chart of Research

IV. RESULTS AND DISCUSSIONS

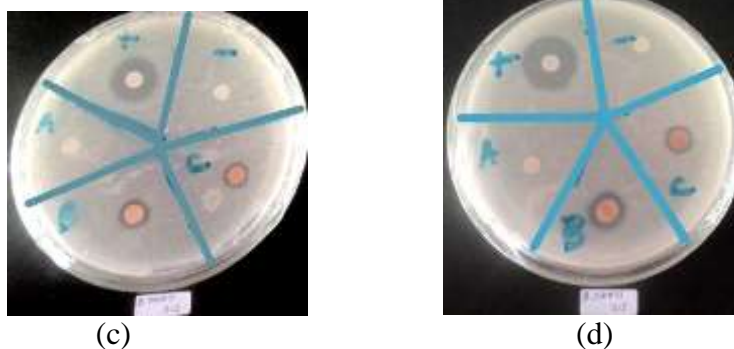
The plant material was macerated using gradient polarity of solvent and afforded *n*-hexane (3.6 kg), ethyl acetate (55.8 kg), and methanol (67.2 kg) extracts. EtOAc extract performed the most bioactive extract against two bacterial strains, *Escherichia coli* and *Bacillus subtilis* using agar disc diffusion (Fig. 3; Table 1) and microdilution methods (Fig. 4; Table 2).



(a)



(b)



(c) (d)
A: *n*-hexane; B: EtOAc; C: methanol; (+): chloramphenicol (*B. subtilis*) and amoxicillin (*E. coli*); (-): MeOH 12,5%

Figure 3. Antibacterial assay result by disc diffusion method (a) *E. coli* (0,5 g/disc); (b) *E. coli* (0,3 g/disc); (c) *B subtilis* (0,5 g/disc); (d) *B. subtilis* (0,3 g/disc)

Table 1. Zone diameter of inhibition of stem roots extracts (in mm) against *E. coli* and *B. subtilis*

Concentration	Zone diameter of inhibition against <i>E. coli</i>			Zone diameter of inhibition against <i>B. subtilis</i>		
	<i>n</i> -hexane	EtOAc	MeOH	<i>n</i> -hexane	EtOAc	MeOH
0,3 mg / disc	-	10	8	-	8	7
0,5 mg / disc	-	11	9	-	9	8

Among three extracts tested as tabulated on Table 1, EtOAc extract was found as the most active with the inhibition zone ranging of 8-11 mm against both bacterial strains.

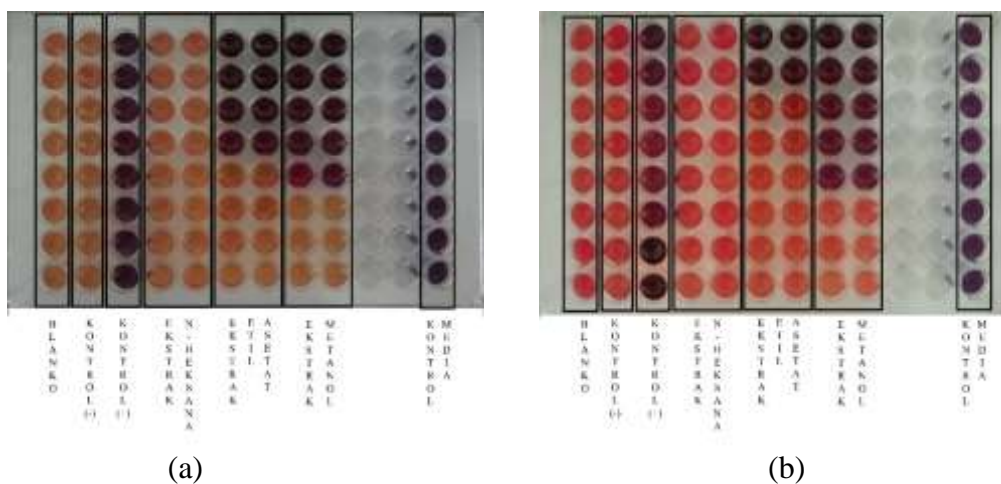


Figure 4. Antibacterial assay result by dilution method (a) *E. coli*; (b) *E. coli*

Table 2. MIC's values of *n*-hexane, EtOAc, and methanol against *E. coli* and *B. subtilis*

No	Extract types	OD mean of <i>E. coli</i>	OD mean of <i>B. subtilis</i>	MIC's value ($\mu\text{g/mL}$) of <i>E. coli</i>	MIC's value ($\mu\text{g/mL}$) of <i>B. subtilis</i>
1	<i>n</i> -hexane	0.76838	0.54231	-	-
2	EtOAc	1.88706	1.48556	12,5	50
3	methanol	1.68644	1.66288	6,25	6.25
4	positive	1.773	1.48025	0,78	0,78
5	negative	0.74013	0.57575	-	-

Antibacterial bioactivity testing using dilution method against *E. coli* and *B. subtilis* bacteria was carried out to find out extracts that had better antibacterial bioactivity. Each extract was dissolved in 12.5% methanol and made in a concentration of 2 mg/ mL, then the ability of the three extracts was tested as an antibacterial agent. Methanol is a good solvent in the extraction of bioactive compounds, because methanol is able to dissolve all types of compounds ranging from those that are polar, semi-polar and even non-polar in small concentrations²⁹. The test results based on Table 5 showed that semi-polar ethyl acetate extract and polar methanol have a minimum inhibitory concentration better than *n*-hexane extract. In the dilution antibacterial test it is not enough to use visual observation only to observe the presence or absence of bacterial growth. But this tends to be subjective from each person's eyesight so the risk of error is relatively greater. This is because the color test makes it difficult to observe, so absorbance values before and after incubation are used to help determine the presence or absence of bacteria. The wavelength used to measure the number of microbes is 600 nm, because cells in the mitochondria and cytoplasm absorb at that wavelength³⁰.

After incubating for 18 hours and measuring Optical Density, the reaction color was added to rezazurin and re-incubated for 4 hours until the color changes. This color change occurs because rezazurin has a blue color that does not fluorescent and can be reduced to a fluorescent pink color. The change in color from blue to pink is an indicator of cell reduction. Color change in rezazurin is carried out by enzymes in cells in the mitochondria and cytoplasm³¹. Based on the results of the antibacterial tests that have been carried out, ethyl acetate extract (EA) was chosen for the fractionation and purification process because it has the best antibacterial activity in

inhibiting bacterial growth to a minimum inhibitory concentration of 12.5 and 50 $\mu\text{g/mL}$ compared to extract *n*-hexane which has no inhibition against all bacterial strains. Furthermore, the EA extract was monitored using thin layer chromatography (TLC) to determine the separation pattern using a combination of eluents in order to obtain the separation pattern of the components of the compound. The eluent combination used is ethyl acetate/acetone 1: 1 as can be seen in Figure 5.

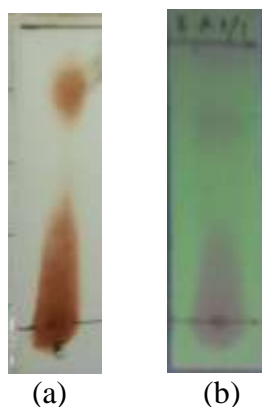


Figure 5. TLC chromatogram of EtOAc with eluent EtOAc / acetone 1:1 (a) sprayed by staining reagent of cerium(IV)sulphate; (b) observed under UV lamp λ 254 nm

Based on the antibacterial activity results of extracts, the ethyl acetate extract (55.8 g) was selected to be isolated and fractionated further using VLC method on silica-gel (35-70 Mesh). The column was eluted with a stepwise gradient polarity of solvent systems including *n*-hexane–EtOAc (100%-0% of *n*-hexane); EtOAc-acetone (100%-0% of EtOAc); and acetone-MeOH (100%-0% of acetone), affording 23 fractions (200 mL each). Due to the similarity of TLC's profile of some fractions using EtOAc-acetone (1:1) as eluent (Fig.2), these fractions were analysed also by $^1\text{H-NMR}$ spectroscopy. According to their chemicals profile analysed by TLC and their proton NMR spectrum, these fractions were grouped and combined into seven primary fractions, E₂1 (fr.1-10), E₂2 (fr.11), E₂3 (fr.12), E₂4 (fr.13), E₂5 (fr.14-15), E₂6 (fr.16-19), E₂7 (fr.20-23). All fractions E₂1- E₂7 were tested for their antibacterial property separately using disc diffusion and microdilution methods. The MIC's values and optical density (OD) means of bioactive fractions tested were tabulated on Table 3.

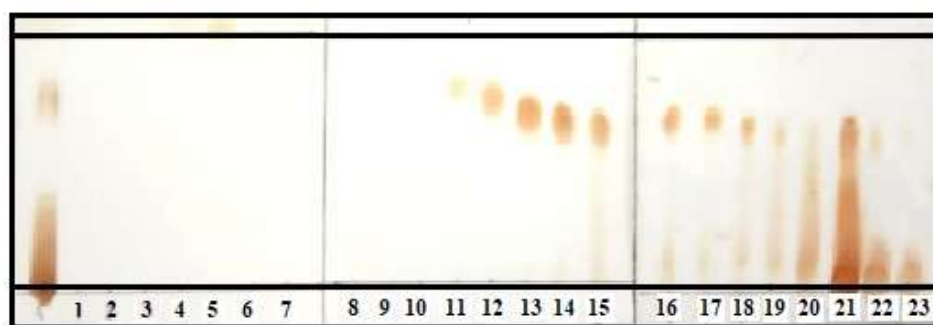


Figure 6. TLC chromatogram of EtOAc extract fractionation by VLC

Table 3. The MIC's values and optical density (OD) means of all fractions tested against *E. coli* and *B. subtilis*

No	Fraction code	OD mean against <i>E. coli</i>	OD mean against <i>B. subtilis</i>	MIC ($\mu\text{g/mL}$) against <i>E. coli</i>	MIC ($\mu\text{g/mL}$) against <i>B. subtilis</i>
1	E ₂ 1	1.16363	1.02819	50	-
2	E ₂ 2	1.49338	1.54488	25	25
3	E ₂ 3	1.25269	1.74956	25	25
4	E ₂ 7	1.39525	2.34931	25	6,25
5	Positive control	2.16563	2.28425	0,78	0,78
6	Negative control	0.86413	0.60838	-	-

Noted: (+) control: chloramphenicol (*B. subtilis*) and amoxicillin (*E. coli*); (-) control: MeOH 12.5%

Three fractions exhibited antibacterial activity against both bacterial strains with quite similar MIC's values, therefore fraction E₂2 (439.5 mg) was subjected to further fractionate due to the simplest chemical profile on its TLC (Fig.7). Fraction E₂2 was redissolved in acetone and then purified with CC on silica gel G-60 (35-70 Mesh) using *n* – hexane / isopropyle alcohol with the ratio volumes of 70/30, 60/40, 50/50, and 40/60, generating 6 major subfractions, E₂2a (132.0 mg); E₂2b (8.4 mg); E₂2c (9.0 mg); E₂2d (47.0 mg); E₂2e (2.0 mg); and E₂2f (50.3 mg). All subfractions were screened against both bacterial strains tested using microdilution methods. The MIC's values and optical density (OD) means of bioactive subfractions were described on Table 4. Among them, two subfractions (E₂2e and E₂2f) performed the most antibacterial activity against *B. subtilis* and *E. coli* with the MIC's values ranging of

12.5-25 $\mu\text{g/mL}$. Only subfraction E₂2f was selected to be analyzed further by ¹H-NMR spectroscopy as having sufficient quantity.

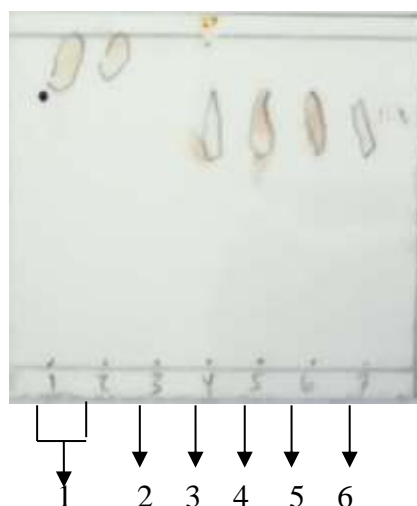


Figure 7. TLC chromatogram of CC of fraction E₂8₁ with eluent isopropyl alcohol / *n*-hexane (6/4)

Table 1. The MIC's values and optical density (OD) means of sub fractions tested against *E. coli* and *B. subtilis*

No	Subfractions code	OD mean against <i>E. coli</i>	OD mean against <i>B. subtilis</i>	MIC ($\mu\text{g/mL}$) against <i>E. coli</i>	MIC ($\mu\text{g/mL}$) against <i>B. subtilis</i>
1	E ₂ 8 ₁ 1	1.49731	1.27825	50	100
2	E ₂ 8 ₁ 2	1.44425	1.22788	50	50
3	E ₂ 8 ₁ 3	1.29094	0.99919	25	50
4	E ₂ 8 ₁ 4	1.259	0.84613	25	50
5	E ₂ 8 ₁ 5	1.39463	1.18294	12,5	25
6	E ₂ 8 ₁ 6	1.34638	1.141	25	25
7	Positive control	1.54525	1.45938	0,78	0,78
8	Negative control	0.85238	0.5695	0	0

Based on the results obtained it can be seen a decrease in the minimum inhibitory concentration compared to the antibacterial activity test on VLC results as observed on the purification results of fraction E₁₃. This fact occurs due to the possible

composition of active compounds contained in the sample. According to Priya et al.³², when an increase or decrease in a component of an active compound there is a possibility of influencing the activeness of the compound itself. From the test results shown in Table 4, it was found that the E₂₈₁₆ subfraction gave a smaller value of the minimum inhibitory concentration compared to other subfractions both against *E. coli* and *B. subtilis* bacteria. Based on the activeness of the two subfractions, E₂₈₁₆ was then selected for ¹H-NMR analysis to predicted the structure of the E₂₈₁₆ subfraction.

The spectrum shown in Figure 8 indicates that the E₂₈₁₆ subfraction is not pure. However, interpretation of the ¹H-NMR data can still be done by referring to the results of the phytochemical test of the fraction which shows the content of phenolic compounds or flavonoids in the subfraction, therefore a comparison is sought in journals reporting the presence of phenolic or flavonoid compounds in the same plant type to estimate the types of bioactive compounds which is obtained. From the NMR spectrum of proton subfraction E₂₈₁₆ shows indications of aromatic protons in the chemical shift region δH 6 - δH 7 ppm. The signal for the methoxy group (-OCH₃) at δH 3.7 ppm, the proton signal at 8.01 ppm with the peak of the chemical shift of the hydroxyl group (-OH) attached to an aromatic ring. As well as the typical peaks for alkanes at 0.89 - δH 2.2 ppm.

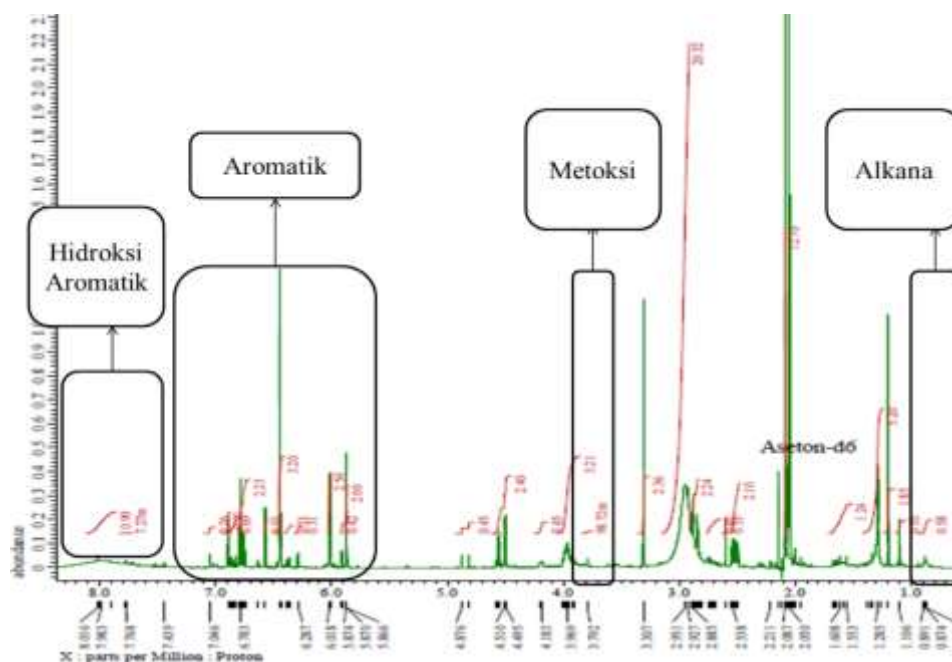


Figure 8. ¹H-NMR spectrum of E₂₈₁₆ subfraction

Sopian et al.³³ (2019) have succeeded in isolating phenolic compounds from the stem of the jengkol plant and were identified as 1-(2,6-dihydroxy-4-methoxyphenyl)-decan-1-one (Figure 19). Based on the comparison of the proton chemical shift values between the isolated subfraction and 1-(2,6-dihydroxy-4-methoxyphenyl)-decan-1-one, it is estimated that the main active compound contained in the E₂₈₁₆ subfraction is the phenolic compounds namely 1-(2,6-dihydroxy-4-methoxyphenyl) decan-1-one (Fig. 9). However, further purification in the E₂₈₁₆ subfraction still needs to be done to ensure the active compounds which are responsible for inhibiting the test bacteria.

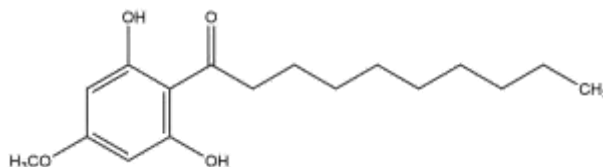


Figure 9. Proposed structure of E₂₈₁₆ subfraction: 1-(2,6-dihydroxy-4-methoxyphenyl)-decan-1-one

V. OUTPUTS

The targeted outputs of the research (both mandatory and additional) are mentioned as below:

1. Publishing in International reputable scientific journals (Targeted: submitted to Asian Pacific Journal of Tropical Medicine, Q2 Scimago, Scopus indexed or if possible the Journal of Asian Natural Products Research, Q2 Scimago, Taylor and Francis, IF: 1091). These journal targeted were proposed and mentioned in the proposal, but due to the insufficient data obtained from the research, the targeted journal was changed to Brazillian Journal of Pharmacognosy (International, Elsevier, IF: 1.754, Q2) or Pharmaceutical Biology (International, Q2 Scimago, Scopus indexed). Additionally, the status of paper in this research is still as a draft of manuscript (Appendix 1). The preparation of manuscript is still going in progress. However, we will update the status of the paper once it has been submitted and under review.
2. Attending to International Conference/Seminar as oral presenter (Targeted: registered). This output has been achieved and approved as can be seen in Appendix 2.

- The Technology Readiness Level (TRL) has been achieved from this study is TRL 6.
- Obtained the prototipe/lead compounds: some bioactive fractions exhibited good antibacterial activity with the MIC's values ranging of 12.5-25 $\mu\text{g/mL}$ (see Appendix 3).

The final output of the research is described on fishbone chart which can be seen on Fig.10.

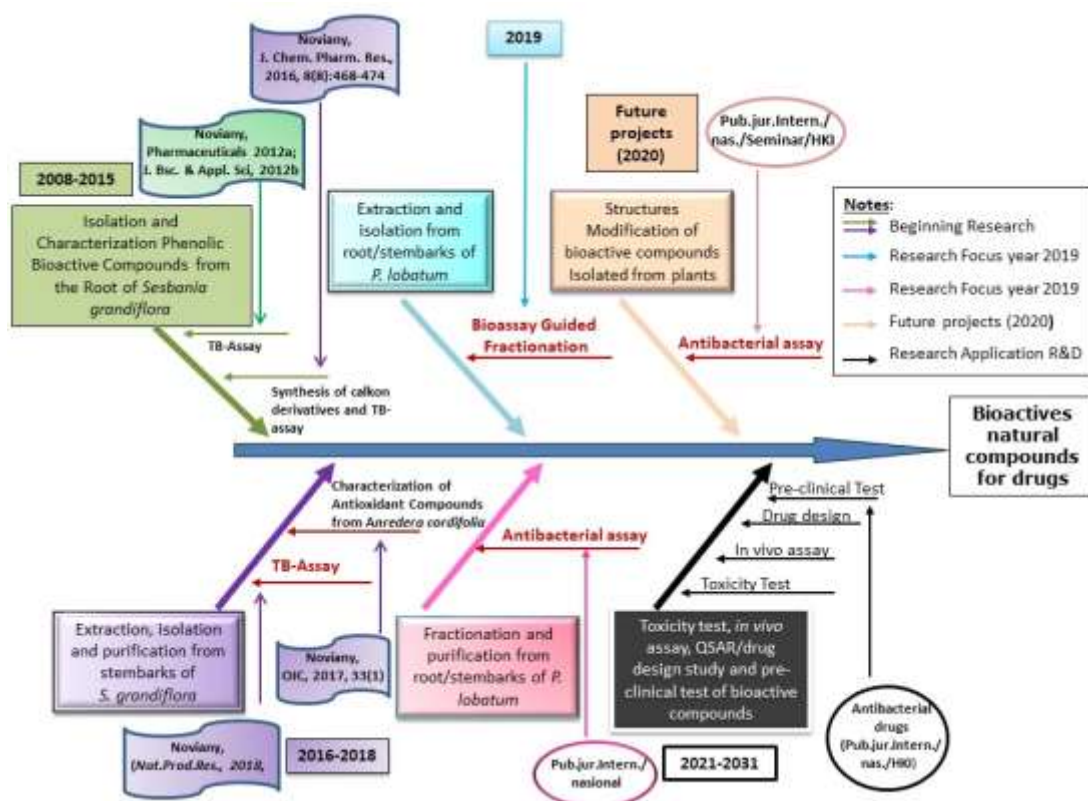


Figure 10. The Fishbone Chart of Research

VI. CONCLUSIONS

Based on the results of the research conducted it can be concluded that:

- Ethyl acetate extract of jengkol (*Archidendron jiringa* (Jack) I. C. Nielsen) root bark has antibacterial activity against *E. coli* and *B. subtilis* bacteria which is better than *n*-hexane extract.
- E13 fraction and E28 fraction have better antibacterial activity against *E. coli* and *B. subtilis* bacteria compared to other fractions from the same column.

3. E13 fraction and E28 fraction have better antibacterial activity against *E. coli* and *B. subtilis* compared to E13 subfraction and E28 fraction as a result of its purification.
4. Based on the ¹H-NMR result data the compounds detected in the E₂₈₁₆ subfraction is phenolic compound type.

VII. SCHEDULE OF RESEARCH

No	Aktivities	Month, year 2019							
		2	3	4	5	6	7	8	9
1	Sampling for roots and stembark of jengkol								
2	Maceration								
3	Extraction and fractionation								
4	Bioassay guided fractionation								
5	Isolation using chromatography techniques								
6	Purification								
7	Structure elucidation								
8	Antibacterial assay of pure compounds								
9	Final report and write up the manuscript								

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APPENDIX 1. DRAFT OF MANUSCRIPT

Bioassay-Guided Separation Approach for Characterization of New Antibacterial Fractions from the Stem Roots Extracts of *Archidendron jiringa*

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ABSTRACT

Infectious diseases caused by bacteria has become the global health issues especially antibacterial drug resistance. The most serious concern with antibacterial resistance is that some bacteria have become resistant to almost all antibacterial drugs and making them less effective. *Archidendron jiringa* is one of the most potent medicinal plants to be developed as a new source of antibacterial components. In current study, the separation of bioactive fractions of *A. jiringa* stem roots was carried out through several stages including isolation, fractionation, and characterization based on the antibacterial assay-guided approach. The stages of isolation of secondary metabolites were conducted by gradually extraction followed by fractionation using chromatographic methods. The antibacterial potential of extracts has been evaluated by the disc diffusion and microdilution methods employing the resazurin assay against one gram negative resistant bacteria, *Escherichia coli* and one positive bacteria, *Bacillus subtilis*. Among three extracts obtained, the ethyl acetate and methanol extracts performed the significant antibacterial effect, while no antibacterial activity was showed on the hexane extract. The fractionation of ethyl acetate extract led to the isolation of the most bioactive fractions (E₂₈₁₅ and E₂₈₁₆) with the MIC's values ranging of 12.5 - 25 µg/mL for both bacteria strains. Due to the less quantity, only the fraction E₂₈₁₆ was subjected to analyse by ¹H-NMR spectroscopy. The results exhibited that the bioactive fraction was obtained as a mixture of at least three major constituents. However, the purification of the bioactive fraction is required to further clarify the antibacterial compound that can be utilized as a new promising antibacterial agent. The bioassay-guided separation approach and the dye resazurin as an indicator of the growth of bacteria are applied for the first time for the phytopharmacological investigation from this plant. The present study represented the effective method for subsequent finding and isolation of potential novel antibacterial constituents from *A. jiringa* stem roots, in particular against the multi-drug resistant strains.

Keywords

Antibacterial assay, *Archidendron jiringa*, bioassay-guided separation, medicinal plant; resistant bacteria

ABBREVIATIONS

<i>B. subtilis</i>	<i>Bacillus subtilis</i>
CC	column chromatography
<i>E. coli</i>	<i>Escherichia coli</i>
EtOAc	ethyl acetate
IPA	isopropyl alcohol
IR	infrared
MeOH	methanol
<i>M. tuberculosis</i>	<i>Mycobacterium tuberculosis</i>

NMR	nuclear magnetic resonance
TLC	thin layer chromatography
UV	ultraviolet
VLC	vacuum liquid chromatography

Introduction

Infection disease is a serious global health problem and its caused the deaths of 13 million people worldwide every year, especially in developing countries like Indonesia. WHO data shows that viral, bacterial, fungal and parasitic infections are the biggest cause of death for the world's population. The use of antibiotics in the prevention of infectious diseases is the only solution¹. While the continuous use of antibiotics raises the new problems for health especially the resistance of microorganisms that cause infection. Bacteria are the most abundant organisms and widespread in living things. In terms of distribution and number, most bacteria may cause disease in humans and other living things². *Bacillus subtilis* and *Escherichia coli* are the common pathogenic bacteria in humans, *E. coli* is a gram-negative bacterium found in the large intestine of humans, and it is a major cause of diarrheal diseases, especially in infants and children^{3,4}. The mortality rate from diarrhea in Indonesia is still around 7.4%, while the mortality rate due to persistent diarrhea is higher at 45%. In 1000 inhabitants, 200-374 subjects experience diarrhea with 60-70% of whom are children under the age of 5 years⁵.

Bacterial resistance to antibiotics is a problem that has not been resolved until now. Refdanita et al.⁶ found that *E. coli* was resistant to chloramphenicol group antibiotics by 83.9% and amoxicillin by 86.2%. The greater the percentage of bacterial resistance to an antibiotic state that bacteria are no longer susceptible to these antibiotics. Various studies to overcome the bacterial resistance have been carried out, but there have been no effective reports. Thus, it is necessary to look for new antibacterial substances that are still active and selective. The search for antibiotic sources from natural ingredients is still the main trend for researchers. The ability of bioactive compounds of natural materials as a healing medium is estimated because of the content of secondary metabolites, including terpenoids, steroids, coumarin, flavonoids, and alkaloids.

One of the plants that has not been studied intensively in Indonesia is the family of Fabaceae. The Fabaceae family has quite interesting bioactivity such as antioxidants, antimalarial, anticancer, and antibacterial. The jengkol plant (*Archidendron jiringa* (Jack) I. C. Nielsen) which belongs to the family Fabaceae is commonly used by the Indonesian people as traditional medicine. Jengkol leaves are efficacious as medicine for eczema, scabies, sores, and ulcers, the skin of the fruit is used as ulcer medicine¹. Several studies have been carried out on jengkol plants, both the leaves, fruit peel, and seeds. Nurussakinah (2010)⁷ has carried out the phytochemical screening on jengkol fruit peel, seeds, bark and leaves extracts. Based on these screening its contain alkaloids, steroids, triterpenoids, glycosides, saponins, flavonoids, and tannins. While research on the stem roots of the jengkol plant has never been done.

In our ongoing investigation for new lead constituents from medicinal plants, we elaborated the bioactive secondary metabolites of *A. jiringa* stem roots and assayed their antibacterial activity. The aims of the study is to obtain antibacterial active fractions from the stem roots of *A. jiringa* based on bioassay-guided separation approach through their antibacterial property.

Materials and methods

Plant material

Samples of the stem roots of *A. jiringa* were assembled on 25 January 2018 from Unila's Housing area at Gedongmeneng District, Bandar Lampung, Lampung Province, Indonesia. The plant specimens (NV6/NRGD/2018) was identified at the Herbarium Bogoriense, LIPI Bogor, Indonesia.

General experimental procedures

TLC was done on silica gel 60 GF₂₅₄ plates (Merck; 0.25 mm) and sprayed with staining reagen Ce(SO₄)₂. Column chromatography (CC) was made on silica gel (Kieselgel 60, 70-230 mesh ASTM; Merck) and sephadex LH-20. ¹H NMR spectrum was measured in acetone-d₆ (TMS as an internal standard), on an NMR Agilent 500 MHz spectrophotometer (Agilent Technologies, JNM-ECZ500R/S1) or Bruker 500 MHz spectrometer. Finally, *Microplate Reader* Hospitex-Italy was used

to measure the absorbance resulted on the Resazurin assay, while UV spectra were performed using an Eppendorf BioSpectrometer® kinetic instrument.

Bacteria strains and biochemicals

Ampicillin and chloramphenicol were purchased from Sigma Aldrich, resazurin sodium salt was purchased from Sigma-Aldrich, gram positive bacteria: *Bacillus subtilis* ITBCCB148 was obtained from the Microbiology and Fermentation Technology Laboratory, Bandung Institute of Technology, gram negative bacteria: *Escherichia coli* UNIATCC25922, nutrient agar broth, disposable sterile petri dishes (Idealcare), disposable syringe, micropipette tips, sterile tissue culture plates 96-well.

Preparation of extracts

Fresh chopped the stem roots of *A. jiringa* (2.5 kg) was cleaned by rinsing under running tap water to remove soil and dirt. The samples was dried in an open space for three weeks and the air-dried roots are finally ground into powder form. The powdered air-dried stem roots (1.5 kg) were extracted with polar gradient polarity of solvent using maceration technique. The solvent used at this stage starts from the solvent which has the lowest polarity, *n*-hexane, followed by ethyl acetate and finally with a high polarity organic solvent, methanol. Each extraction was conducted three times in each type of solvent. Before changing the type of solvent, the extract residue is first dried air for at least three days or until the residue is dry/free of solvent. The filtrates obtained from maceration obtained from polar gradient solvents are then separated from the residue by ordinary filtration. The filtrates are then concentrated under reduced pressure using a vacuum rotary evaporator to yield *n*-hexane (3.6 kg), ethyl acetate (55.8 kg), and methanol (67.2 kg) extracts, respectively. Each extract obtained was subjected to assay of its antibacterial activity using agar disc diffusion and microdilution methods. The general research flowchart can be seen in Fig.1.

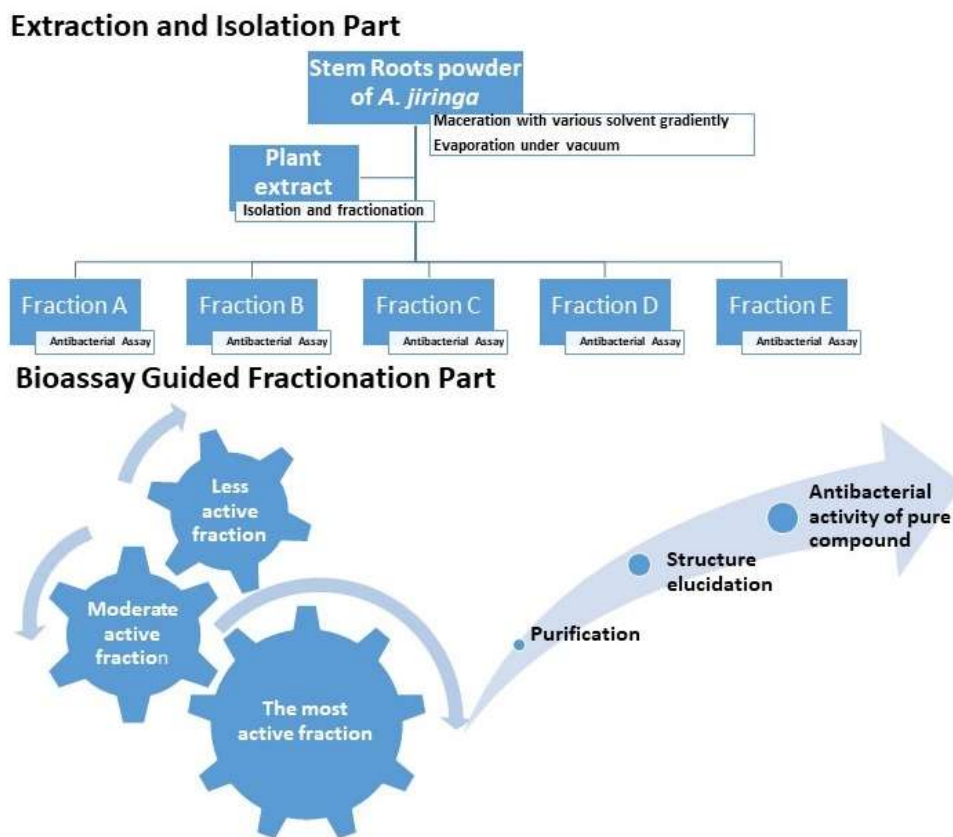


Figure 1. Research flow chart of bioassay guided separation

Bioassay guided separation of ethyl acetate extract

Based on the antibacterial activity results of extracts, the ethyl acetate extract (55.8 g) was selected to be isolated and fractionated further using VLC method on silica-gel (35-70 Mesh). The column was eluted with a stepwise gradient polarity of solvent systems including *n*-hexane–EtOAc (100%-0% of *n*-hexane); EtOAc-acetone (100%-0% of EtOAc); and acetone-MeOH (100%-0% of acetone), affording 23 fractions (200 mL each). Due to the similarity of TLC's profile of some fractions using EtOAc-acetone (1:1) as eluent (Fig.2), these fractions were analysed also by ¹H-NMR spectroscopy. According to their chemicals profile analysed by TLC and their proton NMR spectrum, these fractions were grouped and combined into seven primary fractions, E₂1 (fr.1-10), E₂2 (fr.11), E₂3 (fr.12), E₂4 (fr.13), E₂5 (fr.14-15), E₂6 (fr.16-19), E₂7 (fr.20-23). All fractions E₂1- E₂7 were tested for their antibacterial property separately using disc diffusion and microdilution methods.

The MIC's values and optical density (OD) means of bioactive fractions tested were tabulated on Table 1.

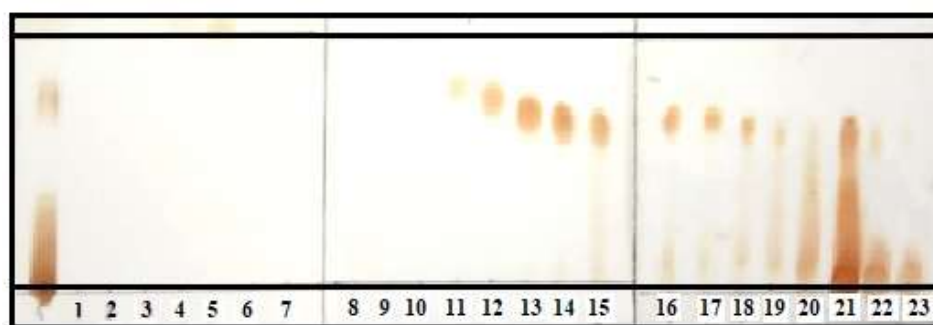


Figure 2. TLC chromatogram of EtOAc extract fractionation by VLC

Table 1. The MIC's values and optical density (OD) means of all fractions tested against *E. coli* and *B. subtilis*

No	Fraction code	OD mean against <i>E. coli</i>	OD mean against <i>B. subtilis</i>	MIC ($\mu\text{g/mL}$) against <i>E. coli</i>	MIC ($\mu\text{g/mL}$) against <i>B. subtilis</i>
1	E ₂₁	1.16363	1.02819	50	-
2	E ₂₂	1.49338	1.54488	25	25
3	E ₂₃	1.25269	1.74956	25	25
4	E ₂₇	1.39525	2.34931	25	6,25
5	Positive control	2.16563	2.28425	0,78	0,78
6	Negative control	0.86413	0.60838	-	-

Noted: (+) control: chloramphenicol (*B. subtilis*) and amoxicillin (*E. coli*); (-) control: MeOH 12.5%

Three fractions exhibited antibacterial activity against both bacterial strains with quite similar MIC's values, therefore fraction E₂₂ (439.5 mg) was subjected to further fractionate due to the simplest chemical profile on its TLC. Fraction E₂₂ was redissolved in acetone and then purified with CC on silica gel G-60 (35-70 Mesh) using *n* – hexane / isopropyle alcohol with the ratio volumes of 70/30, 60/40, 50/50,

and 40/60, generating 6 major subfractions, E₂2a (132.0 mg); E₂2b (8.4 mg); E₂2c (9.0 mg); E₂2d (47.0 mg); E₂2e (2.0 mg); and E₂2f (50.3 mg). All subfractions were screened against both bacterial strains tested using microdilution methods. The MIC's values and optical density (OD) means of bioactive subfractions were described on Table 2. Among them, two subfractions (E₂2e and E₂2f) performed the most antibacterial activity against *B. subtilis* and *E. coli* with the MIC's values ranging of 12.5-25 µg/mL. Only subfraction E₂2f was selected to be analyzed further by ¹H-NMR spectroscopy as having sufficient quantity.

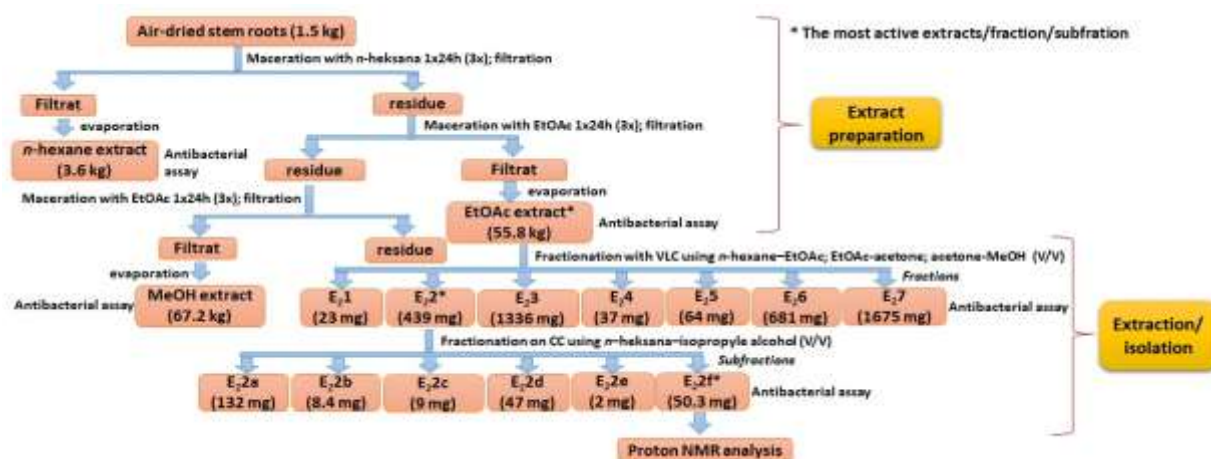


Figure 3. Scheme of bioassay guided separation of EtOAc extract

Phytochemical screening

The phytochemical screening was performed for triterpenes/steroids, alkaloids, flavonoids and saponins by using the standard procedures⁸. The formation of precipitate or the colour intensity was applied for analytical response of screening results.

Evaluation of antibacterial activity

For the testing antibacterial activity, the microbial strains employed in the biological assay are Gram positive bacteria: *Bacillus subtilis*; Gram negative bacteria: *Escherichia coli* has been obtained from stock culture Hospital Abdul Muluk, Bandar Lampung, Indonesia. Original culture are further stored at low temperature in the

refrigerator to maintain stock culture. Fresh cultures are used for testing antibacterial activity using disc diffusion assay and dilution methods.

Disc diffusion assay

The antibacterial activity of the stem roots extract (*n*-hexane, ethyl acetate and methanol) were tested by disc diffusion method⁹ against pathogenic bacteria gram negative (*Escherichia coli*) and gram positive (*Bacillus subtilis*). In this method, freshly prepared agar media is dispensed into the sterilized Petri-dish. The agar is allowed to solidify and 100µl of bacterial suspension poured over the agar media and spread by a spreader or a rod. Ampicillin and chloramfenicol (30µg/dish) is used as positive control, while methanol is used as a negative control. In the each culture medium petri-dish four dish were used, one disk of antibiotics, two discs separately for (*n*-hexane, ethyl acetate and methanol) extracts, one disk used as a control (methanol). The plates are sealed and incubated overnight at 37°C in the incubator. Antibacterial activity is assigned by measuring the inhibition zone formed around the discs. The diameter of zone of inhibition (mean of three replicates SD) as indicated by clear area was measured to determine the antibacterial activity. The experiment is replicated three times to confirm the reproducible.

Determination of MIC via Resazurin assay

Resazurin assay was carried out in 96 well titration plates with some minor modification¹⁰. In complete nutrient broth two fold dilutions of plant extracts and antibiotics are prepared in the test wells. The final concentration (20µL of each bacterial suspension) was added to 180 µl of antibiotics and plant extracts (30-0.02 mg/mL in sequence) contained in culture medium as well as the antibiotics concentrations of 0.06 mg/mL and 0.12 mg/mL for ampicillin and chloramphenicol, respectively. For comparative study control plates were prepared only with culture medium and bacterial suspension. The plates were sealed and incubated for 12 hours at 37 C for additional 5 hour. At the intervals of 1 hour plates were observed for colour change blue to pink and pink to colorless in live bacterial strains containing wells. Preliminary micro titre plate assay reveal that the fast decolonization of resazurin extract doesn't possessed antibacterial potential. The bioactivity of extracts were screened which shows that the extracts inhibit the dye reduction.

Results and discussion

The plant material was macerated using gradient polarity of solvent and afforded *n*-hexane (3.6 kg), ethyl acetate (55.8 kg), and methanol (67.2 kg) extracts. EtOAc extract performed the most bioactive extract against two bacterial strains, *Escherichia coli* and *Bacillus subtilis* using agar disc diffusion and microdilution methods.

2. Conclusions

The ethyl acetate extract of jengkol (*Archidendron jiringa* (Jack) I. C. Nielsen) root bark has antibacterial activity against *E. coli* and *B. subtilis* bacteria which is better than *n*-hexane extract. E13 fraction and E28 fraction have better antibacterial activity against *E. coli* and *B. subtilis* bacteria compared to other fractions from the same column. E13 fraction and E28 fraction have better antibacterial activity against *E. coli* and *B. subtilis* compared to E13 subfraction and E28 fraction as a result of its purification. Based on the ¹H-NMR result data the compounds detected in the E₂₈,6 subfraction is phenolic compound type. This study is the first reported from this plant and could be potential to further investigated for the invention of new promising antibacterial agent from plants.

Conflict of Interest

The authors declare that they have no conflict of interest.

Acknowledgments

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APPENDIX 2. ATTENDING INTERNATIONAL CONFERENCE

1. ABSTRACT

**Isolation and Separation Bioactive Secondary Metabolites from Jengkol Plant
(*Archidendron jiringa* (Jack) I. C. Nielsen) Through
*Bioassay Guided Fractination Approach***

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Infectious diseases caused by bacteria has become the global health issues especially antibacterial drug resistance. The most serious concern with antibacterial resistance is that some bacteria have become resistant to almost all antibacterial drugs and making them less effective. Plants are the one of the most important natural resources because they are relatively less side effects and cheap. One of the potential plants that has antibacterial properties is the jengkol plant (*Archidendron jiringa* (Jack) I. C. Nielsen). Jengkol plant is the one of Lampung's natural resources that has not been optimally used, particularly in the field of technology development and drug health. In current research, isolation and separation secondary metabolites of the roots bark of *A. jiringa* have been conducted through *Bioassay Guided Fractination Approach*. Bioactive secondary metabolites from jengkol plant roots has been carried out using general isolation procedures including extraction with maceration method, fractionation and purification by various chromatographic techniques such as vacuum chromatography (VLC), column chromatography (CC) and medium performance liquid chromatography (MPLC). The preliminary identification of bioactive metabolites were recognized by TLC monitoring and ¹H-NMR spectroscopy analysis, and its displayed a good antibacterial activity against resistant bacteria, *S. aureus* and

E. coli with the same MIC value of 25 μ g / mL Based on the $^1\text{H-NMR}$ spectroscopy analysis, the major compound identified on the bioactive metabolites was predicted as a phenolic compound. However, the purification of these bioactive metabolites is still in progress and will be reported in the future.

Keywords: *Archidendron jiringa*; jengkol plant; bioassay guided fractionation; antibacterial activity; *Staphylococcus aureus*; *Escherichia coli*;

2. ACCEPTANCE LETTER

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No: 004/08/2019
2019

18th August

LETTER OF ACCEPTANCE

Dear Mrs. Dr Noviany Noviany S.Si., M.Si,

On behalf of the organizing committee, I would like to inform you that the **The 14th Joint Conference on Chemistry (14th JCC)** will be held on 10-11 September 2019 at Solo Paragon Hotel and Residence, Surakarta, Indonesia.

We have accepted your abstract entitled: "Isolation and Separation Bioactive Secondary Metabolites from Jengkol Plant (*Archidendron jiringa* (Jack) I. C. Nielsen) Through Bioassay Guided Fractination Approach"

for Oral_Presentation.

For presenter(s) who interesting to submit their full article, please submit your

full-paper manuscript on (<https://jcc.uns.ac.id>) and sign in to your dashboard, by August 14th, 2019. All submitted paper will be peer reviewed. Selected paper will be published in AIP Conference proceeding, which thus author should prepare their paper according the AIP template (<https://aip.scitation.org/apc/authors/preppapers>). In case the Scientific Committee suggested paper to be publish in others publisher/journal and the author(s) agree to do so, the author(s) should reformat their paper according to its respective template (<https://jcc.uns.ac.id/index.php/abstract-and-paper-submission/>). The author may have to pay additional cost for submission to and/or publication in journal.

If you have any questions feel free to contact the conference committee at (<https://jcc.uns.ac.id/index.php/contact-us/>) or jcc14@mail.uns.ac.id.

Regards,
Organizing Committee



Dr. rer. nat. Atmanto Heru Wibowo, M.Si

