

PAPER • OPEN ACCESS

## Identification and Detection *Odontoglossum ringspot virus* on Native Orchids Collection of Nurserys in Java, Indonesia

To cite this article: Mahfut 2020 *J. Phys.: Conf. Ser.* **1641** 012075

View the [article online](#) for updates and enhancements.



**IOP | ebooks™**

Bringing together innovative digital publishing with leading authors from the global scientific community.

Start exploring the collection—download the first chapter of every title for free.

# Identification and Detection *Odontoglossum ringspot virus* on Native Orchids Collection of Nurserys in Java, Indonesia

**Mahfut**

Faculty of Mathematics and Natural Sciences, University of Lampung,  
Lampung, 35145, Indonesia.

Corresponding author: mahfut.mipa@fmipa.unila.ac.id

**Abstract.** Nature orchid are one of original floral in Indonesia. Virus infection is one of the limiting factor in the cultivation of orchid. Infection *Odontoglossum ringspot virus* (ORSV) was reported infests native orchids collection in Indonesia. The purpose of this study was to identification and ORSV that infects native orchid nurserys collection in Java, Indonesia. Symptomatic samples were collected from 5 nurserys collections, i.e. Rumah Bunga Rizal (Bandung), Bali Tanaman Hias (Cianjur), Borobudur Orchids Center (Magelang), Kebun Anggrek Bungarinte (Yogyakarta), and Titi Orchids (Yogyakarta). Detection and identification was conducted by serological test using ORSV specific antisera, RT-PCR and DNA sequencing. The serological test using ORSV antisera showed that 3 of 11 sampels reacted positively against ORSV antiserum i.e *Phalaenopsis amabilis* (Cianjur.1, Cianjur.2, and Magelang). RT-PCR of the 3 samples using specific primer of ORSV coat protein (CP) gene amplified a DNA with size  $\pm 474$  bp. Homology analysis of those 3 Indonesian isolates showed highest index similiarity (IS) was 99.8% with corresponding sequences from 10 other ORSV isolates. Phylogenetic analysis showed that ORSV Cianjur.1 and Cianjur.2 isolates clustered in separated group far from ORSV isolates in other countries.

## 1. Introduction

Native orchids have an essential function as a parent crossing in plant breeding [6]. Disease infection becomes an obstacle in cultivating and developing the potential of this plant [5]. The type of virus that is reported to infect orchids and has a wide spread in the world, including in Indonesia is *Odontoglossum ringspot virus* (ORSV). This viral infection causes a decrease in plant vigor, flower quality [4], and the ability of plant photosynthesis [5;9;10].

Areas of spread of ORSV infections include natural forests [5;6;7;8], botanical gardens [7;8;9], dan nurserys [7;8]. Based on survey of five orchid nurseries, i.e Rumah Bunga Rizal (Bandung), Balai Tanaman Hias (Cianjur), Borobudur Orchids Center (Magelang), Kebun Anggrek Bungarinte (Yogyakarta), and Titi Orchids (Yogyakarta), some native orchids was found with symptoms of virus infection that are thought to be caused by ORSV. Therefore, the ORSV identification and detection research needs to be done to update the health status of the natural orchid nursey collection in Indonesia. The application of the results of this study is one of the potential efforts to support the concept of conservation of natural orchids in Indonesia through efforts to protect plants.



## 2. Materials and Methods

*2.1. Detection of Protein Using Serology.* Serological detection was carried out using the DAS-ELISA method on 11 of the most representative orchid leaf sample samples based on infection symptoms from each location. ELISA uses ORSV specific antiserum (Agdia Inc.). Samples were read using an ELISA-reader (BioTek) at a wavelength of 405 nm. If the absorbance value approaches the positive control value or 2-3 times the control buffer then the sample is said to be positive [2].

*2.2. Detection of Nucleic Acid Using RT-PCR.* RNA isolation was performed on positive samples infected with ORSV by ELISA using a Total RNA isolation kit (SBS Genetech Co., Ltd., China). Amplification of RNA by RT-PCR was done by separate methods using specific primers, namely ORSV CP-F1 (5'-ATGTCTTACACTATTACAGACCCG-3') and ORSV CP-R1 (5'-GGAAGAGGTCCAAGTAAGTCC-3') [6]. The RT reaction was carried out at 37°C for 60 minutes, followed by incubation at 96°C for 5 minutes and ended at 4°C. CDNA amplification begins with the stage of predenaturation at 95°C for 5 minutes, followed by 34 cycles, including denaturation at 95°C for 30 seconds, annealing at 50°C for 45 seconds, and extension at 70°C for 1 minute [7]. PCR products were analyzed using electrophoresis on 2% agarose gel. The DNA bands were then visualized on a UV transilluminator (Bio-Rad Transilluminator 2000) and documented using a digital camera.

*2.3. DNA Sequencing and Phylogenetic Analysis.* The amplified DNA is traced by its nucleotide sequence by sending DNA to the 1<sup>st</sup> Base, Malaysia. Nucleotide sequences are analyzed and combined with Suite for Sequence Analysis DNA STAR Lasergene DM Version 3.0.25 software. Alignment analysis of nucleotide sequence ORSV isolates from Indonesia was performed on sequences registered at Genbank using Basic Local Alignment Search Tool (BLAST) ([www.ncbi.nlm.nih.gov](http://www.ncbi.nlm.nih.gov)). Selection based on the distribution of selected regions obtained 4 registered ORSV isolates from Indonesia and 10 ORSV isolates from other countries (Singapore, China, India, Germany, South Korea, Argentina, and Brazil). TMV-Yunnan isolates are used as a comparison outside the group (outgroup). Phylogenetic analysis was performed using Molecular Evolutionary Genetics Analysis (MEGA) software with Neighbor Joining (NJ) method dan Kimura-2 parameter model for distance estimation. The bootstrap value used is 1000 repetitions.

## 3. Results and Discussion

### 3.1. Virus Detection

Based on results of serology detection, the incidence of virus infection was 27.3%. A total of 3 out of 11 total samples reacted positively to the ORSV antiserum with an average absorbance value of 1,125-1,152. The 3 positive samples are 2 samples from the Balai Taman Hias (Cianjur.1, Cianjur.2) and 1 sample from Borobudur Orchids Center (Magelang). The overall positive orchid sample is *Phalaenopsis amabilis* L. (Blume). RT-PCR results on 3 positive ORSV samples showed DNA fragments measuring  $\pm 474$  bp amplified using the CP-specific ORSV gene primer (Figure 1).

### 3.2. Analysis of Nucleotide Sequence

The results of nucleotide sequencing obtained total nucleotide genes CP isolates ORSV-Cianjur.1, Cianjur.2, and Magelang measuring 475-480 nucleotides. BLAST analysis showed that 3 ORSV isolates had a homology of 99% with ORSV isolates from other countries in Asia, Africa, America and Europe. The results of the analysis of 10 other ORSV isolates showed up to 99.8% homology with ORSV isolates from Indonesian nurseries.

### 3.3. Phylogenetic Tree of CP Gene ORSV

The results alignment of the nucleotide sequence show a point mutation in the form of substitution and insertion in ORSV isolates in Indonesia. Magelang isolates experienced the most mutations, namely insertion 4 times, while substitution in 3 isolates occurred 17 times each. This causes these isolates to separate from other Indonesian isolates. The effect of mutations that occur is able to cause changes in the triplet codon coding for amine acids. Magelang isolates show differences in the frequency of the amino acids Cys and Leu which have increased respectively 0.5% and 1.2% and a decrease in Ser and Val of 1.4 % and 0.3%. In contrast to isolates Cianjur.1 and Cianjur.2 which experienced a decrease in the amino acids Cys and Leu by 0.2% and 0.1%, as well as an increase in Ser 0.8% and Asn 0.6% (Table 1). The nucleotide sequence of ORSV CP gene isolates in this study not found deletion.

Phylogenetic analysis showed that 3 ORSV isolates from Indonesian nurseries had a very close kinship. The results of the phylogenetic tree analysis divided ORSV isolates into two main groups, namely the Bogor and German isolate groups separated from the second group of 10 other isolates. This second group is divided into 2 subgroups, namely the first group consisting of Magelang isolates, 3 Indonesian isolates that have been registered at Genbank, and 5 isolates from other countries, while the second subgroup consists of 2 nursery isolates (Cianjur.1 and Cianjur.2). When compared between branches, isolates Cianjur.1 and Cianjur.2 were separate from ORSV isolates from other countries (Figure 2). Although all isolates formed several groups, kinship between isolates was still very close. This can be seen in the phylogenetic tree which only forms sub groups.

The ORSV findings that infect orchids in Indonesian nurseries show a lack of effort to maintain collection plants by the botanical garden. Some of the nucleotide mutations that occur cause two ORSV isolates from nursery in Indonesia, namely Cianjur.1 and Cianjur.2 apart from other isolates. The process of nucleotide mutations in each isolate causes changes in amino acids that are formed in the composition of the viral genome. Changes in amino acids will change the function of genes arranged so that the infectivity also changes [7;8].

The CP gene is conserved so it has the capability of the proffreading mechanism like most other nuclear genes. But with a relatively small size of the viral genome, a slight error will significantly influence the mutation rate. The rate of mutation will produce a genetic variation of the virus thereby increasing the probability of faster evolution [12]. Long branches in isolates Cianjur.1 and Cianjur.2 also indicate that the virus has evolved and can even lead to speciation.

Indonesian ORSV allegedly originated from Germany. [1;3] reported Germany was ranked 14<sup>th</sup> as an importer of seeds and orchid plants to Indonesia from 1997-2001, in addition to the United States, Brazil, India, Singapore, South Korea, China, Japan, Taiwan, and several West Asian countries. Based on this, another effective way to protect and maintain the health status of natural orchids in Indonesia is to limit and control the importation of orchids from other countries.

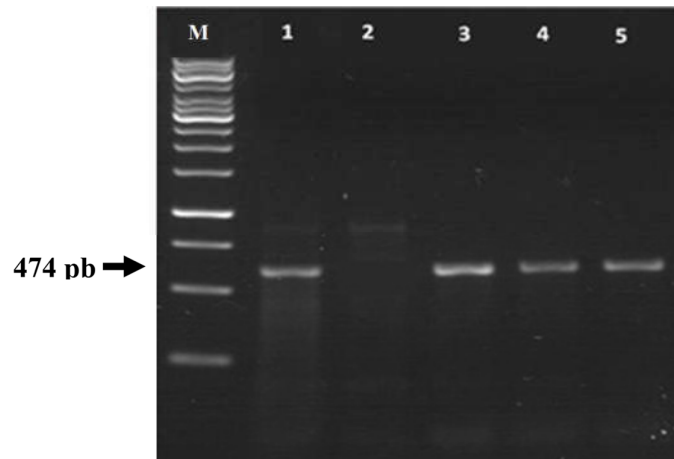
### References

- [1] [BPPP] Balai Penelitian dan Pengembangan Pertanian 2005 *Prospek dan Arah Pengembangan Agribisnis Anggrek* (Jakarta: Departemen Pertanian RI) pp 2-15
- [2] Daryono BS and Natsuaki KT 2009 Survei virus yang menyerang labu-labuan di Yogyakarta dan Jawa Tengah *Jurnal Perlindungan Tanaman Indonesia* **15** 83–89.
- [3] [DPBS] Deputi Bidang Kesejahteraan Rakyat 2011 *Peraturan Presiden Republik Indonesia Nomor 93 Tahun 2011 Tentang Kebun Raya* (Jakarta: Lembaran Negara Republik Indonesia) pp 1-12
- [4] Koh KW, Lu HC and Chan MT 2014 Virus resistance in orchids *Plant Sci.* **228** 26-38
- [5] Kumalawati AD, Abdullah S, Setiadi BS and Mahfut 2011 *Proc. International Conference on Biological Science* (Yogyakarta/Fakultas Biologi Universitas Gadjah Mada) vol 2 p 54
- [6] Mahfut and Daryono BS 2014 Deteksi Odontoglossum ringspot virus (ORSV) Terhadap Anggrek Alam di Hutan Wonosadi, Gunung Kidul *Biogenesis* **2** 101-108
- [7] Mahfut, Joko T and Daryono BS 2016<sup>a</sup> Molecular Characterization Molecular of

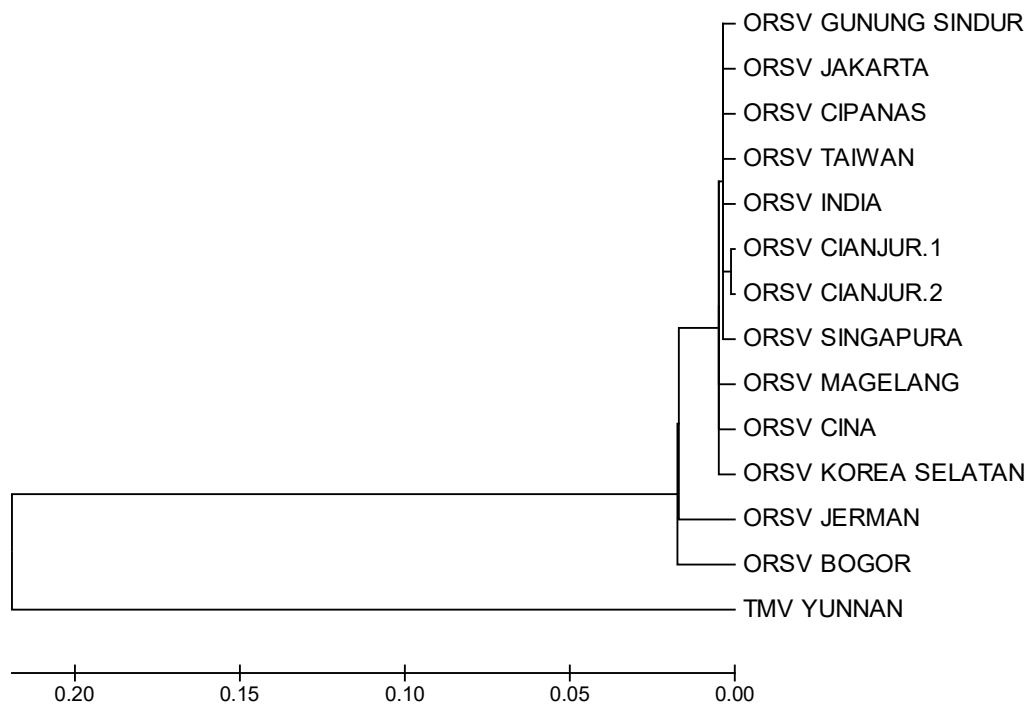
- Odontoglossum ringspot virus (ORSV) in Jawa and Bali, Indonesia *Asian Journal of Plant Pathology* **10** 9-14
- [8] Mahfut, Daryono BS, Joko T and Somowiyarjo S 2016<sup>b</sup> Survei Odontoglossum ringspot virus (ORSV) yang Menginfeksi Anggrek Alam Tropis di Indonesia *Jurnal Perlindungan Tanaman Indonesia* **20** 1-6
- [9] Mahfut Daryono BS and Somowiyarjo S 2017<sup>a</sup> Deteksi Odontoglossum ringspot virus (ORSV) yang Menginfeksi Anggrek Asli Koleksi Kebun Raya di Indonesia *Jurnal Fitopatologi Indonesia* **13** 1-8
- [10] Mahfut, Daryono BS and Somowiyarjo S 2017<sup>b</sup> *Prosiding Seminar Nasional Pengendalian Penyakit Pada Tanaman Pertanian Ramah Lingkungan II Perhimpunan Fitopatologi Indonesia Komisariat Daerah Yogyakarta, Solo, dan Semarang* (Yogyakarta/Fakultas Pertanian Universitas Gadjah Mada) pp 354-360
- [11] Mahfut, Daryono BS, Indrianto A and Somowiyarjo S 2019 *Proc. International Conference on Science and Technology* (Makassar/UIN Alauddin) vol 1
- [12] Matthews REF 1992 *Plant Fundamental of Plant Virology* (California/Academy Press Inc.) p 1

**Table 1.** The frequency of amino acid CP ORSV gene from nursery in Indonesia

Origin of Isolates	Frequency Of Amino Acid (%)																				
	Ala	Cys	Asp	Glu	Phe	Gly	His	Ile	Lys	Leu	Met	Asn	Pro	Gln	Arg	Ser	Thr	Val	Trp	Tyr	Total
Indonesia-Cianjur.1	1,42	0,00	3,55	4,26	4,96	3,55	2,13	8,51	3,55	19,86	4,26	3,55	2,84	5,67	5,67	9,22	4,26	7,09	2,84	2,84	141,00
Indonesia- Cianjur.2	1,42	0,00	3,55	4,26	4,96	3,55	2,13	8,51	3,55	20,57	4,26	2,84	2,84	5,67	4,96	9,22	4,96	7,09	2,84	2,84	141,00
Indonesia-Magelang	2,05	0,68	3,42	3,42	5,48	3,42	2,74	7,53	3,42	21,23	4,11	3,42	2,74	5,48	5,48	8,22	4,79	6,85	2,74	2,74	146,00



**Figure 1.** Results of RT-PCR visualization of several ORSV isolates in agarose gel 2%. (M) Marker 1 kb (Rainbow invitrogen), (1) positive control, (2) negative control of healthy plants, (3-4) ORSV from Balai Tanaman Hias (Cianjur.1 and Cianjur.2), dan (5) ORSV from Borobudur Orchid Center (Magelang)



**Figure 2.** The phylogenetic tree of ORSV isolates based on the nucleotide sequence of CP 3 gene isolates from Indonesian nurseries compared to isolates from other countries. TMV-Yunnan is used as a comparison outside the group