

Analysis of the reducing sugar of cassava (*Manihot esculenta* Crantz.) mutant plantlets resistant to Fusarium wilt

Cite as: AIP Conference Proceedings **2331**, 050010 (2021); <https://doi.org/10.1063/5.0041846>
Published Online: 02 April 2021

Endang Nurcahyani, Hardoko Insan Qudus, and Ferina Evlin



View Online



Export Citation

ARTICLES YOU MAY BE INTERESTED IN

[Community structure and potential of mangrove ecotourism on Harapan Island and Bira Island of Kepulauan Seribu](#)

AIP Conference Proceedings **2331**, 050012 (2021); <https://doi.org/10.1063/5.0041796>

[The conceptualization of marine environmental awareness in early childhood](#)

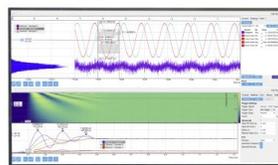
AIP Conference Proceedings **2331**, 050013 (2021); <https://doi.org/10.1063/5.0042008>

[Relationship between environmental pollution knowledge and green purchase intention of students](#)

AIP Conference Proceedings **2331**, 050018 (2021); <https://doi.org/10.1063/5.0041661>

Challenge us.

What are your needs for periodic signal detection?



Zurich
Instruments

Analysis of The Reducing Sugar of Cassava (*Manihot esculenta* Crantz.) Mutant Plantlets Resistant to Fusarium Wilt.

Endang Nurcahyani^{1, 2, a)}, Hardoko Insan Qudus³, and Ferina Evlin²

¹Applied Biology Study Program, Faculty of Mathematics and Natural of Sciences, University of Lampung, Bandar Lampung, Lampung, Indonesia. Jl. Prof. Dr. Soemantri Brojonegoro No. 1, Bandar Lampung, Lampung, Indonesia 35145

²Biology Masters Study Program, Faculty of Mathematics and Natural of Sciences, University of Lampung, Bandar Lampung, Lampung, Indonesia. Jl. Prof. Dr. Soemantri Brojonegoro No. 1, Bandar Lampung, Lampung, Indonesia 35145

³Chemistry Masters Study Program, Faculty of Mathematics and Natural of Sciences, University of Lampung, Bandar Lampung, Lampung, Indonesia. Jl. Prof. Dr. Soemantri Brojonegoro No. 1, Bandar Lampung, Lampung, Indonesia 35145

^{a)}Corresponding author: endang.nurcahyani@fmipa.unila.ac.id

Abstract. Indonesia is the second country producing Cassava (*Manihotesculenta* Crantz) after Nigeria. One of the Indonesian Province, Lampung, is a Cassava central land in Indonesia, with a total production of 7,39 million tons in 2015. However, there are still many Cassava cultivation production constraints, such as Fusarium wilt disease. The disease is caused by the fungus *Fusariumoxysporum*, which until now still cannot be effectively addressed. So far, the farmers have eradicated the fungus by using fungicides, which have adverse impacts on the environment. Therefore, it's necessary to control the disease biologically by using a superior variety resistant to *Fusariumoxysporum*. Fusaric acid is a toxin produced by *Fusariumoxysporum* fungus which is mostly found in vitro selection in plants. Disease resistance can be obtained by induced resistance. The purpose of this study is to analyze the content of the reducing sugar of the Cassava plantlet after induced fusaric acid in vitro. The research using Completely Randomized Design, of one factor, with fusaric acid concentrations, consists of 5 levels 0 ppm, 60 ppm, 80 ppm, 100 ppm, dan 120 ppm in Murashige and Skoog medium. The data analysis used Analysis of Variance with a 5 percent significance level. The results of this study indicate that the increased concentration of fusaric acid, then also increases the reducing sugar content in the *Fusariumoxysporum* resistant Cassava plantlet.

INTRODUCTION

Cassava (*Manihotesculenta* Crantz.), is the third most important crop in the world and a staple food source and also become an income throughout the tropics. Cassava cultivation can be a livelihood for more than 500 million farmers [1]; [2]. Cassava is an important food commodity in Indonesia. In the future, this commodity will have a more strategic role in the lives of the people and the country's economy. Based on the area of the harvest of food commodities, cassava is on the third rank after rice and corn, which are the three main sources of carbohydrates in the community [3].

One of the problems encountered in cultivating Cassava is Fusarium wilt caused by the fungus *F.oxysporum* (*Fo*) and is medium-transmitted.[11,12] It has been reported that 50% of Cassava tubers produced and harvested in Nigeria were lost due to disease. The main causes of the decay of Cassava include *Aspergillusflavus*, *Aspergillusniger*, *Botryodiplodiatheobromae*, *Collectotrichumspp*, *Geotrichumcandidum*, *Penicilliumchrysogenum*, *Pennicilliumdigatum*, *Fusariumoxysporum*. [13,14,15] This organism reduces the quantity and quality of the plant tubers [16]

Indonesia is the second-largest producer of Cassava after Nigeria, with an average total supply of five years of 9.67 million tons or 10.61% of the total world cassava supply, followed by Brazil, India, and the United Republic of Tanzania each ranging from 8.67 - 4.96 million tons or 9.52% - 5.44%, with the rest contributing below 5.30% [4].

The Central Statistics Agency noted that the Cassava land center in Indonesia was controlled by Lampung province with a harvest area of 324,100 ha in 2012. In 2013, cassava production in Lampung reached 8.33 million tons and in 2015 it was 7.39 million tons. This situation makes Lampung as a supplier of one-third national Cassava production from national production of 23.92 million tons [5]. However, there are still many production constraints in Cassava cultivation, including Fusarium wilt disease. This disease is caused by the fungus *Fusariumoxysporum* (*Fo*) which until now still can't effectively be treated.

One of the alternatives methods to control the disease that is efficient, effective, and safe to the environment is the resistant varieties. The use of high yielding varieties that are resistant to high yielding fo is one of the important alternative disease control that doesn't cause negative impacts. Development of *Fo* resistant cassava varieties can be carried out by in vitro selection methods, namely culturing explants in the form of tissue or organs in a medium containing selective fusaric acid concentration [6];[7]; [8].

Research on Cassava Induced Resistance with fusaric acid (FA) has been carried out previously. They found out indications of tolerant FA concentration for the selection of in vitro resistant plantlets. Inoculation of *Fusariumoxysporum* (*Fo*) fungi isolates in resistant Cassava was carried out in vitro, followed by the analysis of DNA patterns compared with controls. The outcome of the study, in the form of a Cassava mutant with a new (specific) DNA band measuring 550 bp (OPA_1) and 300 bp (OPA_10), was predicted as an RAPD marker candidate for Cassava's resistance to *Fo* [17].

The outcome of the study, in the form of a Cassava mutant with a new (specific) DNA band measuring 550 bp (OPA_1) and 300 bp (OPA_10), was predicted as a RAPD marker candidate for Cassava's resistance to *Fo* [17]. Based on the results of this study, it is necessary to study more deeply to ascertain whether the new DNA band is a protein peroxidase that causes Cassava mutants resistant to *Fo*, namely by sequencing analysis of ITS r-DNA and Protein Profiles as well as other specific characters namely Total Phenol content and reducing sugar. The purpose of this study was to analyze the reducing sugar content of Cassava fusarium wilt resistant mutants. Cassava mutant plantlets treated with various concentrations of fusaric acid: (A) 0 ppm; (B) 60 ppm; (C) 80 ppm; (D) 100 ppm; and (E) 120 ppm is presented in **Figure 1**.

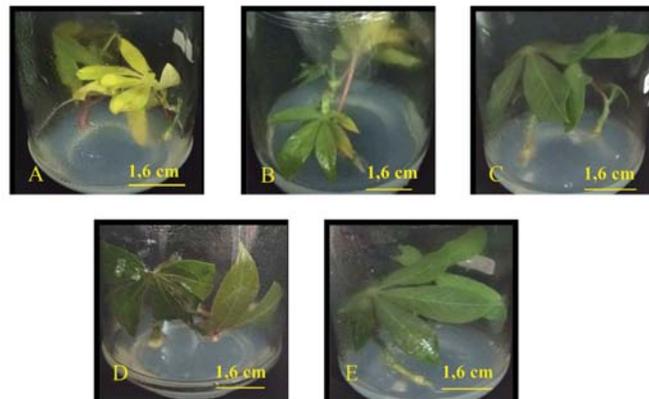


FIGURE 1. Control Cassava Planlets (A) and Cassava mutant plantlets treated with various concentrations of fusaric acid: B.60 ppm;C.80 ppm; D. ppm; and E.120 ppm.

METHOD

This research was conducted from April 2020 to July 2020 at the In Vitro Culture Laboratory, Biology Department, Faculty of Mathematics and Natural Sciences, University of Lampung.

Analysis of changes in sugar reduction level on the substrate is using the Luff Schoorl method. Five grams of the fermented substrate are taken from the bottle every 24 hours, then added with 50 mL of distilled water and stirred evenly. The suspension was centrifuged at 4000 rpm for 20 minutes, and the supernatant is used to test the reducing sugar levels. Put pipette 10 mL supernatant into a boiling flask, then pour 10 mL of Luff Schoorl reagent. The sample was boiled on reflux for 10 minutes, then carefully added 6 mL KI 20% and 10 mL H₂SO₄ through the pumpkin wall. Titrate the sample with 0.1 N Na₂S₂O₃ until it turns into yellow, then

adds 1% starch titration continuously until the blue color disappears. Create titration blank using water **instead of a sample**. Reducing sugar levels are calculated by the formula:

$$\text{Reducing sugar (\%)} = (\text{AT} \times \text{Fp}) / (\text{sample weight} \times 1000) \times 100\%$$

AT = weight of reducing sugar; Fp = dilution factor

Data analysis in this study used a Completely Randomized Design, while quantitative data from each parameter were analyzed using Variance Analysis with a confidence level of 95%. If there is a real difference, then continue with the Duncan Multiple Range Test at a 95% confidence level.

RESULT AND DISCUSSION

Fusarium wilt is a very important and economically harmful disease because until now, there has been no effective chemical control [9]. This disease is caused by the fungus *Fusariumoxysporum* f.sp. *lycopersici* (Sacc.). This fungus is a soil-borne pathogen that can survive for a long time in the form of chlamidiospores, even though there are no host plants. Therefore, Fusarium wilt disease is difficult to control. That is why biological control is very efficient in controlling this disease. Fuction using fusaric acid is one of the biological control methods used to control Fusarium wilt.

Characterization of the cassava planlet has a relationship with the resistance of *Fusariumoxysporum*. It can be seen from the content of the total reducing sugar content of cassava plantlets. The average reducing sugar levels in Cassava mutants that are resistant to fusarium wilt are shown in Table 1.

TABLE 1. Average Content of Cassava Planlet Reduction Sugar (%)

Treatment (ppm)	Average content of Reducing Sugar (%)
0 (Kontrol)	3,52 ± 0,05 ^c
60	4,05 ± 0,03 ^{bc}
80	4,71 ± 0,29 ^b
100	6,14 ± 0,09 ^a
120	6,82 ± 0,05 ^a

Note: Numbers followed by letters are not the same, not significantly different at the 95% confidence level

Reducing sugar levels are improved by increasing the concentration of fusaric acid. Reducing sugar level is an increase in cassava plantlets that given fusaric acid treatment and compared with planets without treatment (control). The highest reduction sugar level occurred in the cassava plantlets that were given with fusaric acid scavenging treatment with a concentration of 120 ppm. This treatment shows that the higher concentration used in fusaric acid, the higher content of the sugar reduction produced by the cassava plantlet. This can occur because of the influence of fusaric acid on cassava plantlets. The fusaric acid that is induced on cassava plantlets will be absorbed by plants and one of the responses that will arise is the increase in reducing sugar content in cassava plantlets.

According to the study of [10], the amylase and cellulose activity changes because of the lower level of inoculum concentration that impact enough biomass to reduce the formation of products, while the level highest inoculum concentration allows deep produce more biomass, so, the product formation is getting lower. Enzyme hydrolysis is the hydrolysis of starch and cellulose into glucose. Glucose is then used as energy by mold *Aspergillusniger* for metabolism and growth.

CONCLUSION

Cassava plantlets that are resistant to Fusarium wilt disease show different expression characteristics with cassava plantlets that are not resistant to *Fusariumoxysporum*, namely an increase in total dissolved reducing sugar content in cassava planets that are resistant to *Fusariumoxysporum*, along with the increased concentration of fusaric acid.

ACKNOWLEDGMENT

Thanks to the authors to the Institute for Research and Community Service through the BLU fund of the University of Lampung, based on the Letter of Assignment of “**Penelitian PASCASARJANA**” 2020 Number of Contracts: 1510/UN26.21/PN/2020 tanggal 24 Maret 2020.

REFERENCES

1. O. C. Eleazu, K. C. Eleazu, and S. Kolawole, *Acta Sci. Pol., Technol. Aliment.* **13**, pp. 249-256 (2014).
2. S. K. Amponsah, E. Y. H. Bobobee, W. A. Agyare, J. B. Okyere, J. Aveyire, S. R. King, and J. Sarkodie-Addo, *American Society of Agricultural and Biological Engineers* **30**(3), pp. 391-403 (2014).
3. M. Fauzi, E. H. Kardhinata, dan L. A. Putri, *Jurnal Online Agroteknologi* **3**(3), pp. 1082– 1088 (2015).
4. Anonymous, *Statistik Harga Komoditas Pertanian Tahun* (Pusat Data dan Sistem Informasi Pertanian (Pusdatin), Jakarta, 2013).
5. Anonymous, ‘Produksi Ubi Kayu’, (2015). [Online]. Available from: <https://www.bps.go.id/> [Accessed Februari 6, 2020).
6. E. Nurcahyani, Sumardi, B. Irawan, E. Y. Sari, and T. L. Sari, *WJPLS* (2019a) **5**(2), pp. 195-198 (2019).
7. E. Nurcahyani, Sumardi, B. Irawan, E. Y. Sari, and T. L. Sari, *Journal of Tropical Upland Resources* **1**(1), pp. 93-102 (2019).
8. E. Nurcahyani, Sumardi, H. I. Qudus, S. Wahyuningsih, Sholekhah, and A. Palupi, *World Journal of Pharmaceutical and Life Sciences. WJPLS* (2020) **6**(2), 25-28 (2020).
9. C. M. I. Borrero, Trillas, J. Ordovás, J. C. Tello, and M. Aviles, *Phytopathology* **94**(10), pp. 1094- 1101 (2004).
10. Imandi, S. Babu, S. K. Karanam, and H. R. Garupati, *Journal Microbial Biochemistry Thecnology* **2**, pp. 29-35 (2010).
11. A. E. Arinze, *An Inaugural Lecture Series* **43**, pp. 29-7 (2005).
12. R. N. Okigbo, R. Putheti, and C. T. Achusi, *E-J Chem* **6**(4), pp. 1274-1280 (2009).
13. A. O. Ogunleye and O. T. Ayansola, *American Journal of Microbiology and Biotechnology* **1**(1), pp. 9-20 (2014).
14. N. R. Okigbo, C. E. Enweremadu, C. K. Agu, R. C. Irondi, and B. C. Okeke, *Advances in Applied Science Research* **6**(10), pp. 7-1 (2015).
15. I. V. Gwa, A. A. Bem, and J. K. Okoro. *Journal of Phytopathology and Plant Health* **3**, pp. 38-43 (2015).
16. N. A. Amusa, A. A. Adegbite, S. Muhammed, and R. A. Baiyewu, *African Journal of Biotechnology* **2**(12), pp. 497-502 (2003).
17. E. Nurcahyani, B. Irawan, Sumardi, E. Y. Sari, and T. L. Sari, *Journal of Tropical Upland Resources* **1**(1), pp. 93-102 (2019).