

ITS-rDNA Sequence Analysis of Mycorrhizal Endophytes from Native Tropical Orchids in Indonesia

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Abstract

Tricodherma sp. mycorrhiza is an important role as a biocontrol agent. Its association with Phalaenopsis amabilis was molecularly identified through ITS- rDNA sequence analysis. The aims of the study were to identify isolated molecular of orchids mycorrhiza from native tropical orchids in Indonesia, conducted as one of native orchid conservation efforts in Indonesia. One isolate of Tricodherma sp. were isolated from the root of orchid plant in Yogyakarta based on morphological and microscopical analysis. Verification analysis molecular of these isolates resulted in 600-750 bp DNA products located on the ITS1-5.8S-ITS4 region. The sequenced products showed insertion and substitution occurrences, which may result in strain diversity and possible variation in severity. Reconstruction of phylogenetic trees using Maximum Parsimony and Bootstrap-1000 approach showed that Indonesian isolates have undergone speciation and have been positioned in the cluster, which are already far apart from the other isolates. Isolate Tricodherma sp. from Yogyakarta, Indonesia successfully isolated based on identification of ITS- rDNA sequences. Results of this study were expected to become the basic information in an effort of native orchid cultivation and protection against infectious diseases in Indonesia. The study was the first to report regarding Tricodherma sp. isolated from native tropical orchids in Indonesia.

Keywords: *Tricodherma; ITS- rDNA; Phalaenopsis; Indonesia*

1. Introduction

Phalaenopsis is a genus of orchids, which some of its members have important role as parent crosses. The presence in its native habitat (forest) has been reported to have greatly diminished. Thus, Infections by bacteria, fungi, and viruses are still major obstacles in conducting cultivation and development of native orchids in Indonesia [1;7;8;9;10;11;12;13;14;17]. Cultivating and protecting *Phalaenopsis sp.* against diseases in Indonesia could be done through the induction of endophytic microorganisms [2;15;16;18], which includes the Orchid Mycorrhizal Fungi (OMF). One type of OMF which have been isolated and identified is *Tricodherma*.

Tricodherma could playing a role as biological control agents in crop protection [5], in general OMF has a role in stimulating the germination of orchid seeds [22], in supporting the provision of nutrients for growth and development of plantlets. In addition for induce the resistance of *Phalaenopsis sp.* against infections by bacteria, fungi, and viruses [3]. In this research, the identification of *Tricodherma* was molecularly conducted through analysis of ITS-rDNA sequence isolated from *Phalaenopsis amabilis* grown in Indonesia. This research was expected to become the basic information on the development of cultivation and protection of nature orchids and where possible, on the prevention of the occurrence of the mentioned diseases in Indonesia.

2. Materials and Methods

2.1. Plants Materials

The sampling of healthy roots of *Phalaenopsis amabilis* was conducted at four different locations: orchid garden in Condong Catur and Parakan (Yogyakarta), Balikpapan Botanical Garden (East Kalimantan), and Sultan Adam Forest (South Kalimantan). On the separated study, isolation results showed positive samples of mycorrhizal endophyte *Tricoderma* based on morphological and microscopical analysis.

2.2. Molecular Analysis

Genomic DNA isolation was performed using techniques modified from *cetyltrimethylammonium bromide* (CTAB) method [4] on samples of pure cultures of isolated mycorrhizal endophyte *Tricoderma*. Genomic DNA was PCR amplified according to the manual instructions of *GoTaq® Green PCR mix* (Promega). Predenaturation reaction and amplification was carried out using methods by [6], with a pair of universal primers, rDNA-ITS1 (5'-TCCGTAGGTGAACCTGCGG-3') and rDNA-ITS4 (5'-TCCTCCGCTTATTGATATGC-3'). PCR products were visualized using electrophoresis on 2% agarose gel stained with ethidium bromide and 100 bp *Vivantis DNA ladder* was used as marker. The visualized DNA bands indicated the length of the targeted base pairs of ITS- rDNA, which were subjected to sequencing.

2.3. Phylogenetic Analysis

Sequencing results were analyzed using *Sequence Scanner* software; the nucleotide sequences were combined using *EditSeq* and *SeqMan* of the *Software Suite for Sequence Analysis DNASTAR Lasergene DM Version 3.0.25*. *BLAST* software was used to determine and to compare the sequence homology with the data contained in the DDBJ database. Comparison between sequences of isolates was carried out using *Algorithm Multiple Alignment Parameters DNA with Kimura-2 Parameters*, relationship and phylogenetic analysis using the *Neighbour Joining* of *MEGA 5 Beta* program. Statistical analysis on internal branch was done using *the bootstrap* value with 1000 replication.

3. Results and Discussion

3.1. Sample Collection

A total of 12 samples of healthy roots of *Phalaenopsis amabilis* were isolated from 4 different locations, such as orchid garden in Condong Catur and Parakan (Yogyakarta), Balikpapan Botanical Garden (East Kalimantan), and Sultan Adam Forest (South Kalimantan). Positive result of *Tricoderma* was found from orchid garden in Parakan, which had colony characteristic traits i.e. yellowish to white color, colony appearance like a cotton, 90° branching hyphae shape, binucleate, with colony growth rate of 0.72 mm/hour, referring to [21;23].

3.2. Molecular Analysis

ITS- rDNA amplification results showed a specific band with a size between 600 and 750 bp similar to that reported by [6]. Internal transcribed spacer (ITS) is an area of the nuclear ribosomal DNA (nrDNA), which has the role of providing important information on the reconstruction of phylogenetic trees at different taxonomic levels [20], as well as similarity at the level of intrageneric[9].

Sequencing results were combined and analyzed using *DNASTAR Lasergene DM Version 3.0.25*. Total number of nucleotide of MP isolates which successfully

scanned was 661 with 41.4% GC content. Analysis results were obtained by incorporating sequences to <http://blast.ncbi.nlm.nih.gov/> site and confirmed that MP isolates were *Tricodherma*. Furthermore, search analysis of homologous sequences carried out using BLAST on NCBI site resulted in 47 sequences indicating a close relationship with high level of similarity of 99%. This result showed that the MP sequences are representative of *Tricodherma* isolates, a non-pathogenic (endophyte) associating with the root of orchid (OMF).

The whole isolates originated from various countries in Asia (Israel, China, Taiwan, Japan, Iran, Singapore, and India), America (USA, Cuba, and Argentina), and Europe (Italy, Netherlands and Spain). Some isolates of *Tricodherma*, was collected from the roots of *Vanda miss Joaquim* (AJ318420) and *Brite Ng* (AJ318429) orchids, and also from the roots of other plants such as *Rosa hybrida cv. Linda* (KC825348), strawberry *Fragaria x ananassa* (AY927319) and apples (EU002945.1). Total bases amplified from each isolate ranged from 560 to 725 bp, except from DQ097889.1 isolate with a total amplified base of 1500 bp. Maximum BLAST score between isolates ranged from 1005 to 1186 with homology (ident.) 99% and query cover ranged from 84 to 99%.

Analysis on 14 other *Tricodherma* isolates selected based on the distribution area of each different country, showed that isolates from Indonesia were highly different from those of other countries with similarity index (IS) ranging from 45.2 to 49.9%. It was clear that isolates from Indonesia has been undergoing speciation and are different from isolates from other countries.

Alignment result of nucleotide sequences of 15 *Tricodherma* isolates showed high occurrence of point mutations in isolates from Indonesian, which were largely insertions and substitutions. Of the 606 total bases aligned, there were 44 bases having substitution consisting of 21 transition and 23 transversion bases.

Each of these mutations had an influence on changes in amino acids formed. Of the total 194 amino acids belonging to isolates from Indonesia, some of which had very different percentage from those of other isolates. Some amino acids of isolates from Indonesia had increased their number dramatically compared to those of other isolates, i.e. Ala, Glu, Val, Gln, Lys, and Tyr by 0.7%, 1.76%, 1.8%, 2.3%, 3, 1%, and 4.73%, respectively, of total average. Several other amino acids had also decreased in number, i.e. Asp, Arg, Trp, Thr, and Leu by 0.11%, 1.53%, 2.07%, 2.25%, and 4.32%, respectively, of total average.

It appeared that the resulted amino acid changes had enormous influence in the process of adaptation to the environment in Indonesia. Results of the relationship analysis between isolates through the reconstruction of phylogenetic tree showed that isolates from Indonesia were on separated branches and far apart from 14 other isolates (**Figure 1**).

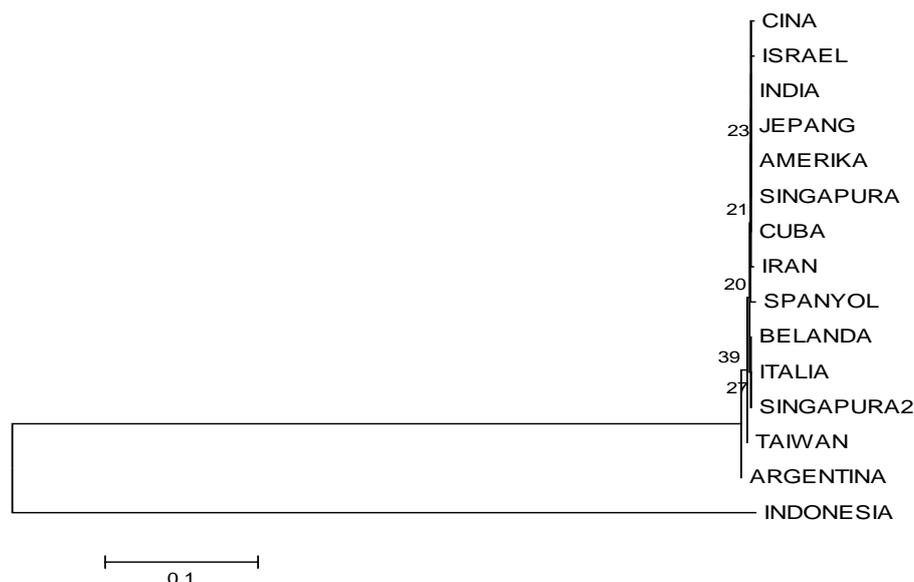


Figure 1. Reconstruction of phylogenetic trees of *Tricoderma* isolates based on nucleotide sequences with Maximum Parsimony method with Bootstrap-1000 approach

Maximum Parsimony method with *Bootstrap-1000* approach classify the data based on specific characters (discrete character states) on which nucleotide sequences are used to obtain information about evolution through evolutionary data changes. The results of analysis were classified based on distribution on the geographical areas. A lengthy branch, which well separating between the two groups, indicated that isolates from Indonesia had considerably evolved and it led to speciation events. Such changes in sequence through mutation was supported by the ability of the mycorrhizal itself to adapt to environmental changes as well as to its host range. The adaptability of mycorrhiza is strongly influenced by the presence of amino acid changes due to mutations that alter the function of genes which they arrange.

The reconstruction of phylogenetic tree, in addition to explaining relationship based on geographic regions, also determine the origin and history of distribution of the isolates. Trading activities of plants, fruits, and seedlings from several countries which are reported to have been infected by *Tricoderma* is most likely the cause of its spreading. Indonesia is known as an importer country of orchid plant seedlings from Asian countries such as Thailand, Singapore and Taiwan [8], and possibly have imported seedling and fruit of apple and strawberry plants from other countries such as India and Italy that had been reported infected by *Tricoderma* isolates. Proximity with Singapore in terms of geographical location is also very possible to allow distribution activities of orchids between the two countries.

This study results proved that nucleotide sequences and amino acid of *Tricoderma* isolates from Indonesia were different with other isolates from another country (Asia, Europe, and America) due to adaptation to various environmental conditions. Furthermore, the mutations are capable of causing the *Tricoderma* to lose its ability to infect and to associate with host plants. Another possible result of mutations is that *Tricoderma* could have the ability to infect plants while showing pathogenic characters or otherwise become mycorrhizal endophytic.

One way to control pathogens of orchids is by using biological control, such as the mechanisms of mycorrhizal resistance induction. *Tricoderma* as OMF has a role as a biological agent in disease control [5] including infection of ORSV. This

study results clarified the MP sequence as OMF *Trichoderma* isolates from Indonesia based on identification of its ITS- rDNA sequences. Furthermore, this study was expected to be the basic information beneficial for the improvement of cultivation effort as well as for the development of biocontrol agents through natural orchid protection against pathogens and diseases.

References

- [1] A.D. Kumalawati, S. Abdullah, B.S. Setiadi, and Mahfut, “Study on Genetic Diversity and Conservation of Orchids in Wonosadi Forest, Gunung Kidul Based on Molecular Analysis”, Proceedings of International Conference on Biological Science, Yogyakarta, Indonesia. (2011). pp. 54.
- [2] E.A. Tanawy, “Acquainting With Salt Tolerant Endophytic Bacteria Isolated From Rice”, Plant Growth, vol. 1, no. 2, (2009). pp. 72-79.
- [3] J.A.E. Moreno, E.A.G. Acuña, A.E.B. Román, D.J. Contreras, and T.J. López, “Chemical and Biological Fertilization of Phalaenopsis (Orchidaceae) Under Greenhouse Conditions”, Terra Latinoamericana, vol. 18, no. 2, (2000). pp. 125-131.
- [4] J.J. Doyle and J.L. Doyle, “A Rapid DNA Isolation Procedure For Small Quantities of Fresh Leaf Tissue”, Phytochem. Bull., vol. 19, (1987). pp. 11–15.
- [5] J.T. Otero, A.T. Mosquera, and N.S. Flanagan, “Tropical Orchid Mycorrhizae: Potential Applications in Orchid Conservation, Commercialization, And Beyond”, Lankesteriana, vol. 13, no. 1–2, (2013). pp. 57-63.
- [6] K. Nadarajah, N.S. Omar, M.M. Rosli, and O.S. Tze, “Molecular Characterization and Screening For Sheath Blight Resistance Using Malaysian Isolates of *Rhizoctonia solani*”, BioMed Research International, (2014). pp. 1-18.
- [7] Mahfut and B.S. Daryono, “Deteksi Odontoglossum ringspot virus (ORSV) Terhadap Anggrek Alam di Hutan Wonosadi, Gunung Kidul”, Biogenesis, vol. 2, no. 2, (2014), pp. 101-108.
- [8] Mahfut, T. Joko, and B.S. Daryono, “Molecular Characterization Molecular of Odontoglossum ringspot virus (ORSV) in Jawa and Bali, Indonesia”, Asian Journal of Plant Pathology, vol. 10, no. 1-2, (2016^a). pp. 9-14.
- [9] Mahfut, B.S. Daryono, T. Joko, and S. Somowiyarjo, “Survei Odontoglossum ringspot virus (ORSV) yang Menginfeksi Anggrek Alam Tropis di Indonesia”, Jurnal Perlindungan Tanaman Indonesia, vol. 20, no. 1, (2016^b). pp. 1-6.
- [10] Mahfut, B.S. Daryono, and S. Somowiyarjo, “Deteksi Odontoglossum ringspot virus (ORSV) yang Menginfeksi Anggrek Asli Koleksi Kebun Raya di Indonesia”, Jurnal Fitopatologi Indonesia, vol. 13, no. 1, (2017^a). pp. 1-8.
- [11] Mahfut, B.S. Daryono, and S. Somowiyarjo, “Identifikasi Molekuler DNA Kloroplas Pada Anggrek Terinfeksi Odontoglossum ringspot virus (ORSV) di Magelang, Jawa Tengah”, Prosiding Seminar Nasional Pengendalian Penyakit Pada Tanaman Pertanian Ramah Lingkungan II Perhimpunan Fitopatologi Indonesia Komisariat Daerah Yogyakarta, Solo, dan Semarang, Yogyakarta, Indonesia, (2017^b), pp. 354-360.
- [12] Mahfut, “Indonesia Darurat Konservasi: Sudah Amankah Kebun Raya Kita?”, Prosiding Seminar Nasional Biodiversitas Indonesia, Jurusan Biologi, Fakultas Sains dan Teknologi, UIN Alauddin Makassar, (2019). pp. 1-6.
- [13] Mahfut and B.S. Daryono, “Variation Symptoms and Resistance Response of Different Types on Orchids (Orchidaceae) Against Odontoglossum ringspot virus (ORSV) Infection”, International Series on Interdisciplinary Science and Technology, vol. 4, no. 2, (2019). pp: 246–249.
- [14] Mahfut, B.S. Daryono, A. Indrianto and S. Somowiyarjo, “Plant-Virus Interaction on Orchids Infected Odontoglossum ringspot virus (ORSV) in Bogor Botanical Garden, Indonesia”, The 1st International Conference on Science and Technology (ICoST), Makassar, Indonesia (2019^a).
- [15] Mahfut, B.S. Daryono, A. Indrianto, and S. Somowiyarjo, “Effectiveness Test of Orchid Mycorrhizal Isolate (Ceratorhiza and Trichoderma) Indonesia and Its Role as a Biofertilizer”, Annual Research & Review in Biology, vol. 33, no. 4, (2019^b). pp. 1-7.
- [16] Mahfut, “Effectiveness Test of Orchid Mycorrhizal Isolate (Ceratorhiza and Trichoderma) Indonesia and Its Role as a Biofertilizer: Critical Overview”, Current Research Trends in Biological Science Vol. 1., Book Publisher International, India & United Kingdom, (2020^a). pp. 139-145.

- [17] Mahfut, “Variation of Resistance Responses on Indicator Plants Against Odontoglossum ringspot virus (ORSV) Infection”, International Journal of Advanced Science and Technology, vol. 29, no. 3, (2020^b). pp. 11780-11785.
- [18] Mahfut, A. Indrianto, S. Somowiyarjo, and B.S. Daryono, “ Molecular Phylogeny Of Orchids Mycorrhiza Isolated From Native Tropical Orchids In Indonesia”, Malaysian Journal of Microbiology, vol. 16, no. 1, (2020). pp. 68-72.
- [19] M.C. Lee, S.F. Cheng, D.C.N. Chang, Y.J. Chang, and Y.S. Chang, “Specific Detection Of Mycorrhizal Colonization In Orchid Roots By Fluorescence Microscopy”, Journal of Biotechnology, vol. 150, (2010). pp. 496.
- [20] R.J. Bayer, D.E. Soltis, and P.S. Soltis, “Phylogenetic Inferences in Antennaria (Asteraceae; Gnaphalieae; Cassiniinae) Based on Sequences From Nuclear Ribosomal Dna Internal Transcribed Spacers (ITS)”, American Journal of Botany, vol. 83, no. 4, (1996). pp. 516-527.
- [21] R.S. Currah and C.D. Zelmer, “A Key and Notes For The Gen Genera of Fungi Mycorrhizal With Orchids, and a New Species in The Genus Epulorhiza”, Rep. Tottori Mycol Inst., vol. 30, (1992). pp. 43–59.
- [22] T.F. Andersen and H.N. Rasmussen, “The Mycorrhizal Species Of Rhizoctonia, In: Rhizoctonia Species: Taxonomy, Molecular Biology, Ecology, Pathology And Disease Control”, B. Sneh, S. Jabaji-Hare, S. Neate, and G. Dijst. (eds.). KAP, London. (1996). pp. 379-390.
- [23] X.C. Shan, E.C.Y. Liew, M.A. Weatherhead, and I.J. Hodgkiss, “Characterization and Taxonomic Placement of Rhizoctonia-Like Endophytes From Orchid Roots”, Mycologia, vol. 94, no. 2, (2002). pp. 230–239.