Identification of Disease and Efforts to Protect Natural Orchid Plants Against Mixed Infections in the Liwa Botanical Garden

3 4

Mahfut^{1*}, Ketut Lestari¹, Sri Wahyuningsih¹, Tundjung Tripeni Handayani¹, Sukimin²

¹Department of Biology, Faculty of Mathematics and Natural Science, Universitas Lampung. Jl. Prof. Soemantri Brojonegoro 1, Lampung, Indonesia.

²Liwa Botanical Garden. Way Mengaku, Balik Bukit, Lampung Barat, Lampung, Indonesia. Email: mahfut.mipa@fmipa.unila.ac.id

10 ABSTRACT

Liwa Botanical Garden is located in Balik Bukit Subdistrict, West Lampung Regency, is a mountainous area surrounded by hills with a slope of 10% - 40%. This botanical garden has one of its superior collections namely native orchids. Based on the results of previous studies, it is known that several native orchid individuals at Liwa Botanical Garden show symptoms of being infected with a mixture of viruses, bacteria and fungi. Mixed infections will show more severe symptoms than single infections. This study was conducted to determine the identification of diseases and efforts to protect natural orchids against mixed infections at Liwa Botanical Garden through a collection of samples that showed symptoms of infection, analysis of disease symptoms, and analysis of disease resistance levels. The results showed the response of natural orchids at Liwa Botanical Garden to mixed infections showed symptoms in the form of necrosis, streak, chlorotic, soft rot, and fungal spots on the orchid Ascideria longifolia, Polisthacia sp., Bulbophyllum sp., Calanthe tripicata, and Eria sp. The type of orchid that shows the most symptoms is Bulbophyllum sp. a total of 8 samples. The type of natural orchid that is most susceptible to mixed infections is Eria sp. with a disease intensity of 32.7%. The results of this activity are expected to be basic information in efforts to protect plants against diseases to support the application of conservation of native orchids at Liwa Botanical Garden.

Key words: identification of diseases, mixed infections, orchid viruses, orchid fungi, orchid bacteria, liwa botanical garden

INTRODUCTION

Liwa Botanical Garden located in Balik Bukit Subdistrict, West Lampung Regency, is a mountainous area surrounded by hills with a slope of 10% - 40%. This botanical garden has an Indonesian ornamental plant concept with its superior collection of native orchids. Until November 2013, the total number of natural orchid collections was 805 specimens which were included in 425 collection numbers (60 genera, identified to species level of 214 collection numbers). Most collections are *Dendrobium*, *Eria* and *Bulbophyllum* with 48, 40 and 38 numbers, respectively (Saniyatun, 2015). Mahfut et al. (2019) reported several natural orchid individuals at Liwa Botanical Garden showing symptoms of being infected with a mixture of viruses, bacteria and fungi. Mixed infections will show more severe symptoms than single infections. Each infection of this disease can inhibit plant growth and resilience, and decrease the aesthetic value of orchids periodically Mahfut (2019).

To facilitate disease control, an inventory of data is first carried out as preliminary information about the infection. Recognition of symptoms as an infection response is the first data in disease identification. The data is used to determine disease intensity, plant resistance, and disease control efforts. Research on the identification of diseases and efforts to protect natural orchids against mixed infections at Liwa Botanical Garden has never been done. The results of this study are expected to be fundamental data in efforts to protect orchids to support the application of conservation of natural orchids at Liwa Botanical Garden. In addition, the results of this activity serve as a medium of good communication between researchers and managers of Liwa Botanical Garden in their disease control efforts.

Sample Collection. Sample collection was carried out on orchid leaves in Liwa Botanical Garden which 45 showed symptoms of being infected with a virus. All samples were tabulated and documented with photos. 46

47 48

Analysis of Disease Symptoms. This analysis is done by matching the sample documentation with the literature that has been previously reported. The literature used is Muharam et al., 2013; Mahfut and Daryono, 2014; Mahfut et al., 2017; Fery et al., 2018.

50 51 52

49

Disease Intensity Analysis. This stage is done to determine the severity of the disease (disease severity).

Disease intensity is the proportion of infected hosts to the total observed surface area of the host. Calculation of 53 54

disease intensity analysis is done using the method of Rahardjo and Suhardi (2008) and the scale of disease

intensity refers to Mahfut et al. (2019).

55 56 57

58

Plant Resistance Level. Determination of the level of resistance of orchids to disease follows the method of Mahfut et al. (2019).

59

60

61

62

63

64

65

RESULTS AND DISCUSSION

Sample Collection

Sample collection stage is conducted in the second week of January to the second week of February 2020. Samples are randomly selected from individuals who show mixed symptoms of infection at Liwa Botanical Garden green house. Based on the results of the collection obtained 24 samples from 5 types of orchids namely Ascideria longifolia, Polisthacia sp., Bulbophyllum sp., Calanthe tripicata, and Eria sp. The type of orchid that has the most infection is Bulbophyllum sp. as many as 8 samples, then Calanthe tripicata orchids as many as 5 samples. Overall data collection results are shown in Table 1.

66 67 68

Table 1. Collection of orchid leaf samples at Liwa Botanical Garden which shows a mixed infection

No	Species of Orchid	Number of Samples	Symptoms of Infection
1	Ascideria longifolia	3	All parts of the leaf wither and turn yellow, fungal spots, and chlorotic
2	Polisthacia sp.	1	Dry leaves and necrosis
3	Bulbophyllum sp.	8	Soft rot, chlorotic, and necrosis
4	Calanthe tripicata	5	Necrosis, streak, fungus spots
5	Eria sp.	4	Soft rot, chlorotic, fungal spots

69 70

71

72

73

74

75

76 77

78

79 80

81

82

83

84 85

86

Selection of sampling locations is based on previous research (Mahfut et al., 2019), who reported mixed disease infections in the collection of native orchids at Liwa Botanical Garden in August and December 2019. The type of orchid that was mixed was Calanthe sp. and Flicking eria with a mixture of viruses, fungi and bacteria. The study (Mahfut et al., 2019) also reported a mixed infection on Cymbidium sp. with symptoms of full necrosis, streak, and fungal spots (back), in the Corynborsis sp. with symptoms of necrosis, mosaic, fungal spots, and on orchid Flicking eria. with symptoms of large necrosis and mosaics at Liwa Botanical Garden.

In this study, Bulbophyllum sp. being the most mixed type of orchid. This shows that this type of orchid is the most vulnerable host infected with pathogens. Besides that are Ascideria longifolia, Polisthacia sp., Calanthe tripicata, and Eria sp. Mahfut et al., (2019) also reported that on Bulbophyllum sp. showed a fairly severe mixed infection at Liwa Botanical Garden.

Other studies have also reported viral infections with *Phalaenopsis amboinensis* (KRB2) and *Phalaenopsis* amabilis (KRB12) from Bogor Botanical Gardens, Phalaenopsis amabilis (KRP18) and Dendrobium salacence (KRP20) from the Purwodadi Botanical Gardens, and Phalaenopsis modesta J. Sm. (KRBp5) from Balikpapan Botanical Gardens (Mahfut et al., 2017). Muharam et al. (2013) reported the presence of CymMV infection on Phalaenopsis sp. and Dendrobium sp. in Java and Bali, there are symptoms of necrosis.

Other studies have also reported bacterial infections with symptoms of soft rot on Onchidium sp. and Paphiopedilum sp. in Bogor, West Java (Hanudin et al., 2011), Grammatophyllum, Dendrobium, and Catleya sp. in D. I. Yogyakarta (Joko et al., 2010). Research on fungal infections with symptoms of wilting was also reported to infect 3 types of *Dendrobium stratiotes*, *Dendrobium moschatum*, *Dendrobium concinnum*, and *Pteroceras unguiculatum* at Bogor Botanical Gardens (Anita, 2017); spot on *Phalaenopsis* sp. in Lembang, West Bandung (Devi, 2018).

Analysis of Disease Symptoms

Based on the analysis of mixed infectious diseases in orchid samples of *Ascideria longifolia, Polisthacia* sp., *Bulbophyllum* sp., *Calanthe tripicata*, and *Eria* sp. Overall samples showing symptoms of mixed infections namely necrosis, streak, chlorotic, soft rot, and fungal spots that infect native orchids at Liwa Botanical Garden are presented on **Fig 1**.

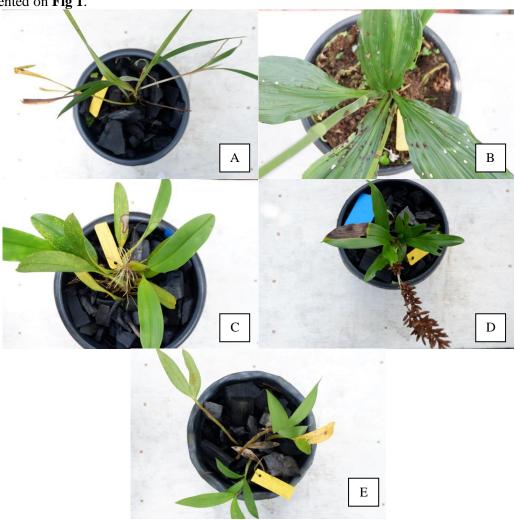
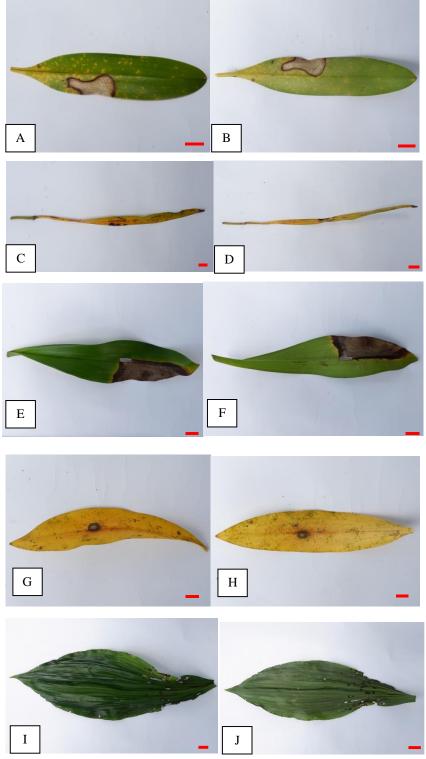


Figure 1. Symptoms of mixed infections with natural orchids at Liwa Botanical Garden: (A) *Ascideria longifolia* (B) *Calanthe triplicata* (C) *Bulbophyllum* sp. (D) *Polisthacia* sp. (E) *Eria* sp.

On Ascideria longifolia sample, a mixed infection causes the entire leaf to wilt and turn yellow, fungal and chlorotic spots. In the Polisthacia sp. Sample, the symptoms caused by a mixed infection were dry leaves and necrosis. In the sample of Bulbophyllum sp., The symptoms were soft rot, chlorotic and necrosis. In the Calanthe tripicata sample the symptoms were necrosis, streak and fungal spots. Whereas on Eria sp. sample, the symptoms were soft rot, chlorotic and fungal spots. A collection of samples of mixed disease symptoms that infect native orchids at Liwa Botanical Garden is presented on Fig. 2.



107

Figure 2. Collection of samples of mixed infection symptoms in native orchids at Liwa Botanical Garden:

(A, B): Soft rot, chlorotic, and necrotic symptoms on Bulbophyllum sp.

- (C, D): Symptoms of fungus, chlorotic spots and all parts of the leaves wither and change color turn yellow on Ascideria longifolia.
- (E, F): Symptoms of necrosis and leaf drying on *Polisthacia* sp.
- (G, H): Symptoms of soft rot, chlorotics, and mold spots on Eria sp.
- (I, J): Symptoms of necrosis, streak, and mold spots on Calanthe triplicate.
- (A, C, E, G, I): Top surface of the leaf, (B, D, F, H, J): Surface under the leaf. Bar: 1 cm

109 110 111

112 113 114

115 116

Necrotic is characterized by physical damage or death to cells or tissues. Some of the symptoms that include necrotic types are necrose, rot, die back, and cancer (dead bark dries with a clear border) (Purnomo, 2006). Chlorotic symptoms are symptoms caused by damage to chloroplasts which results in parts of the plant which are normally green to yellow. Chlorotic symptoms are often associated with necrotic where chlorotic surrounds necrotic called "hello". Whereas curling leaf is a leaf symptom that shows changes in the shape of the edges in the form of curling (Mahfut and Daryono, 2014).

Disease Intensity

The results of disease intensity analysis of mixed dinfection in native orchids in the Liwa Botanical Garden, obtained the percentage of intensity as showed on **Table 2**.

Table 2. Analysis of mixed diseases in natural orchids on Liwa Botanical Garden

No.	Species of Orchid	Disease Intensity
1	Ascideria longifolia	22,5%
2	Polisthacia sp.	8,57%
3	Bulbophyllum sp.	20%
4	Calanthe tripicata	16,6%
5	Eria sp.	32,7%

Eria sp. has the highest disease intensity. This is caused by several factors, including: the percentage of disease events is quite high at 90%, and as many as 4 samples from a total of 11 leaves show mixed infection symptoms. Three samples of Ascideria longifolia showed mixed infection symptoms and the percentage of disease was 90%, but the intensity of the disease in this orchid was smaller when compared with Eria sp. The lower intensity of disease on Ascideria longifolia is caused by the total number of leaves in one plant that is 12 leaves, so that the ratio of the number of diseased and healthy leaves in this orchid is lower when compared to Eria sp. The lower the total ratio of the number of diseased and healthy leaves and the lower percentage of disease events will cause the results of the analysis of disease intensity to be lower (Rahardjo and Suhardi, 2008). Then the percentage of disease intensity of 20% occurred in Bulbophyllum sp., 16.6% in Calanthe tripicata, and the lowest percentage was 8.57% on Polisthacia sp.

Plant Resistance

The results of the analysis of the level of resistance to mixed infection on native orchids in Liwa Botanical Garden are showed on **Table 3.**

Table 3. Analysis of the level of plant resistance to mixed infections on Liwa Botanical Garden

No.	Species of Orcid	Plant Resistance Level
1	Ascideria longifolia	Resistance
2	Polisthacia sp.	Resistance
3	Bulbophyllum sp.	Resistance
4	Calanthe tripicata	Resistance
5	Eria sp.	Resistance

Overall the sample has a level of resistance rather resistant to mixed infections because it has a disease incidence of less than 40%. Mahfut et al. (2019) explain that the level of resistance is somewhat resistant is the level of resistance in orchids infected with the disease but the incidence of the disease is not more than 40%. This shows that the overall collection of natural orchid samples at Liwa Botanical Garden is capable of being infected by pathogens but a small portion of the cells supports the growth and development of pathogens so that they cause less disease.

166 167

172

173

174

175

176

177 178

183

184

192

Thank you to Liwa Botanical Garden who facilitated the research. This activity is a continuation of the collaboration program with Universitas Lampung.

157 **REFERENCES**

- Anggraeni, D. N. dan Sumiati, E. 2019. *Antagonistic Endosymbion Bacteria against Soft-Rot Bacteria in Phalaenopsis* sp. ICOST 2019. Makassar.
- Anita, D.P. 2017. Inventaris Penyakit Pada Tanaman Anggrek di Kebun Raya Bogor.(*Skripsi*). Institut Pertanian Bogor.
- Arditti, J., T. Kull, dan S. M. Wong. 2002. Orchid Biology: *Reviews and Perspective VIII*. Kluwer Academic Publisher. Dordrecht, Belanda.
- Badan Penelitian dan Pengembangan Pertanian. 2017. Pengendalian Organisme Pengganggu Tanaman (OPT)
 pada Budidaya Cabai. Pusat Penelitian dan Pengembangan Holtikultura. Jakarta.
 - Cronquist, A. 1981. An Integrated System of Classification of Flowering Plants. Columbia University Press. New York.
- Djatnika, I., Hanudin, W., Nuryani, dan Silvia, E. 2011. Kemangkusan Formula Jenis Bakteri Antagonis (Hasil Isolasi 2010) terhadap Intensitas Serangan Penyakit Busuk Lunak (Pectobacterium carotovorum pv. Carotovorum) pada Anggrek Phalaenopsis. Laporan Hasil Penelitian Badai Penelitian Tanaman Hias Tahun Anggaran 2010.
 - Edwin, N. 2009. Teknik Pengendalian Hama Dan Penyakit Pada Anggrek Di Widoro Kandang Yogyakarta. Universitas Negeri Yogyakarta. Yogyakarta.
 - Fery A, Tutung H, dan Erlina E.P. 2018. *Identifikasi Penyakit Yang Disebabkan Oleh Virus Pada Tanaman Anggrek Cattleya sp. di Malang, Jawa Timur*. Universitas Brawijaya. Malang.
 - Hanudin, W. Nuryani, K. Budiarto, A. A. Nawangsih, and B. Tjahjono. 2011. Identification of Soft Rot Bacterial Disease on Orchids Collected from West Java and DKI Jakarta. *Journal of The International Society for Southeast Asian Agricultural Sciences* (In Press).
- Harsono, A. 2017. *Pengenalan dan Pengelolaan Gulma pada Kedelai*. Balai Penelitian Tanaman Kacang-Kacangan dan Umbi-Umbian. Malang.
- Hartati, S. dan Darsana, L. 2015. Karakterisasi Anggrek Alam secara Morfologi dalam Rangka Pelestarian Plasma Nutfah. *Jurnal Agronomi Indonesia*. 43(2): 133-139.
 - . Huang, T. C. 2008. *The Occurrence and Control of Fungi andd Bacterial Orchid Disease*. Taichung Branch Office, Bureau of Animal and Plant Health Inspection and Quarantine.
- Hull R. 2002. Matthews Plant Virology. Ed ke-4. San Diego: Academic Press.
- Kartikaningrum, S., Widiastoety, D., dan Effendie, K., 2004. Panduan Karakterisasi Tanaman Hias: Anggrek dan Anthurium. Sekretariat Komisi Nasional Plasma Nutfah, Bogor.
- Mahfut, dan Daryono., 2014. *Deteksi Odontoglossum ringspot virus Terhadap Anggrek Alam di Hutan Wonosadi, Gunung Kidul.* Universitas Lampung. Lampung.
- Mahfut, Wahyuningsih, S., dan Handayani, T.T. 2019. *Konservasi Anggrek Alam di Kebun Raya Liwa* (Laporan Kemajuan Penelitian). Universitas Lampung.
 - Matthews REF. 1992. Plant Fundamental of Plant Virology. California: Academic Press Inc.
- Muharam A, Sulyo, Y, Rahardjo, IB, Diningsih, dan Suryanah. 2013. Studi penyebaran Tobacco Mosaic Virus

 Strain Orchid dan Cymbidium Mosaic Virus dengan Metode DAS ELISA pada Tanaman Anggrek

 Komersial di Pulau Jawa dan Bali serta Teknologi Pembebasannya. Balai Besar Pengkajian dan
 Pengembangan Teknologi Pertanian. Bogor.
- Nuryani, W dkk. 2012. Kemangkusan Biobakterisida terhadap Penyakit Busuk Lunak (*Pseudomonas viridiflava*) pada *Phalaenopsis. Jurnal Hortikultura*. 22(4): 392-399.
- Pearson, MN & Cole, JC. 2008. 'The effects of cymbidium mosaic virus and odontoglossum ringspot virus on the growth of Cymbidium orchids'. J. Phytopathol. 3(19): 193-197
- 201 Prihatman, K. 2000. Anggrek. Budidaya Pertanian. Jakarta.
- Purnomo B. 2006. Konsep Ilmu Penyakit Hutan. Fakultas Pertanian Univeristas Brawijaya. Hal: 1-9.

- Pusat Konservasi Tumbuhan Kebun Raya. 2016. Kebun Raya Liwa. Diakses pada http://kebunrayadaerah.krbogor.lipi.go.id/kebun-raya-liwa.html
- 205 Rahardjo, I. B. dan Suhardi. 2008. Insidensi dan Intensitas Serangan Penyakit Karat Putih pada Beberapa Klon Krisan. *Jurnal Holtikultura*. 18(3): 312-318.
- 209 Rifai, M. A. 1976. Sendi-sendi Botani Sistematika. Lembaga Biologi Nasional LIPI. Bogor.
- Sastrahidayat, I. R. 2011. Fitopatologi (Ilmu Penyakit Tumbuhan). UB Press. Malang.
- Sjahril, R., Chin, D. P., Khan, R. S. Yamamura, S., Nakamura, I., Amemiya, Y., and Mii, M. 2006. Transgenic *Phalaenopsis* Plants with Resistance to *Erwinia carotovora* Produced by Introducing Wasabi Defensing Gene Using Agrobacterium Method. Plant Biotechnology. 23: 191-194.
- Soelistijono, R. 2015. Kajian Efektifitas *Rhizoctonia* SP Mikoriza Dataran Rendah dan Sedang pada Tingkat Keparahan Penyakit (Dsi) Anggrek *Phalaenopsis amabilis* terhadap *Fusarium* sp. Jurnal Biosaintifika. 7(2): 112-119.
- Solihah, S. M. 2015. Koleksi, Status, dan Potensi Anggrek di Kebun Raya Liwa. *Warta Kebun Raya*. 13(1): 14-218 23.
- Supriadi, N. I. dan Taryono. 2002. Karakterisasi *Erwinia chrysanthemi* Penyebab Penyakit Busuk Bakteri pada Daun Lidah Buaya (*Aloe vera*). *Jurnal LITTRI*. 8(2): 45-48.
- 221 Sutiyoso, Y. dan Sarwono, B. 2005. Merawat Anggrek. Penebar Swadaya. Jakarta.
- Suyono, Yoyon dan Salahudin, Farid. 2011. Identifikasi dan Karakteristik Bakteri *Pseudomonas* pada Tanah yang Terindikasi Terkontaminasi Logam. *Jurnal Biopropal Industri*. 2(1): 8-13.
- Timper P, Minton NA, Johnson AW, Brenneman TB, Culbrreat AK, Burton GW, Baker SH, Gascho GJ (2001)
 Influence of cropping system on stem rot (*Sclerotium rolfsii*), *Meloydogyne arenaria*, and the nematode antagonist *Pasteuria penetrans* in peanut. Plant Disease. 85: 767-772.
- Wheeler H. 1975. *Plant Pathogenesis*. Berlin: Springer-Verlag.
- FW Zettler, NJ Ko, GC Wisler, S Elliot, Mark, SM Wong. 1990. Viruses of orchids and control. *Plant Dis.* pp. 621-626.