Phytotelmata Might Account for the High Prevalence of Dengue Hemorrhagic Fever in Lampung, Indonesia

By Emantis Rosa



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Abstract It was revealed that phytotelmata is an importance breeding place for numerous types of insect including the vector mosquitoes of deadly diseases such as dengue and malaria. Yet, such small ponds tend to be neglected by the country health authorities in setting eradication program of the disease vectors. This study aimed to find out whether phytotelmata contribute in providing breeding place for vector mosquito of dengue hemorrhagic fever (DHF)- the Ades larvae. The survey carried out in four districts namely Pesawaran, Metro, Pringsewu and Bandar Lampung and lasted from March to August 2016. Phytotelmic criteria of the study were every structures present in plants which allow rain water to impound and showed the signs of life exist in it. The phytotelmata containing Aedes mosquito larvae were assessed for its physical parameters and the data resulted were analysed and presented descriptively. There were six types of phytotelmata found namely leaf axils, tree holes, tree stumps, fallen spathe, fruit shells, and fallen leaves. The Aedes larvae found belong to only two species, i.e. Aedes albopictus (n=373) and Aedes crysolineatus (n=26).

Keywords Phytotelmata, Mosquito breeding place, Dengue fever, Aedes larvae, Vector mosquito

1. Introduction

Dengue hemorrhagic fever (DHF) is one of the mosquito-borne diseases that is prevalent in the Indonesian provinces, including Lampung. It said prevalent due to the occurrence of the disease is seasonal in pattern and often be an outbreak in a certain area. In the period of 2000-2012, in the province of Lampung with a population of about 8 million, more than 14,174 people suffered from DHF and 151 of them died [1, 2].

Despite the government, both national and local, have been implementing a variety of strategies to eradicate the diseases, among others, through the eradication of the vector but in fact, this disease still occur annually. The question, then, why is it so hard to prevent the disease, especially in relation to vector control? As is known, most Indonesian people have been briefed by the authorities for vector control programs against deadly diseases such as dengue and malaria through the insertion of these materials into primary and secondary school curricula. In the context of DHF, one of the most info 10 tion known to the community is that this disease is transmitted by the mosquito Aedes aegypti [3].

One of the shortcomings of vector control programs in use today, particularly against mosquitoes, is ignoring that the breeding places of the vectors are not just limited to the

ponds, water container, or a variety of man-made waste items such as discarded tyres or used cans. In fact, various species of vector mosquitoes such as *Aedes* sp. were found to breed in a very small water bodies trapped in tree holes, leaf axils, bamboo joints, or papaya stumps [4, 5]. Such small water bodies found in or upon plants are called, by Ludwig Varga in 1928, as phytotelmata [6]. Unfortunately, in Indonesia, study on phytotelmata has not been done in earnest [7]. As a result, the importance of phytotelmata in the epidemiological fields and health management programs has not been put into consideration.

This study aimed to find out whether phytotelmata contribute in providing breeding place for mosquito vector of DHF in Lampung, an Indonesian province situated in the most southern part of Sumatra, by identifying mosquito larvae of *Aedes* inhabit the phytotelmata.

2. Method

Study Area

The study was carried out from the end of rainy season until the peak of dry season, March –July 2016. The survey was targeting four districts in Lampung Province namely Pesawaran, Metro, Pringsewu and Bandar Lampung. Pesawaran, is known as the district with the highest malaria endemicity in which one of the sub-district ever records the levels of annual parasite incidence (API) up to 51.8. Furthermore, Metro, Bandar Lampung and Pringsewu are the three districts that were ranked first, second and third highest

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for the DHF incident rate in 2012 [8].

Inclusion Criteria

The phytotelmic criteria of the study w 20 dopted from Kicthing [6] i.e. every structures present in terrestrial plants such as modified leaves, leaf axils, flowers, stem holes or depressions, tree stumps, open fruits and fallen leaves, which allow rain water to impound and in the impounded water the signs of life was seen visually. Plants that were observed include ornamental plants grown in backyard, wild plants and crop plants that grown naturally or cultivated in plantation areas.

Sampling

Each plant or its parts found to contained phytotelmata was photographed or sampled for further taxonomic identification. The impounded water in each phytotelmata was collected using a hose connected to a manual suction pump, but before being discharged the phytotelmic water 27 perature was measured. The phytotelmic fluid then placed separately in labeled vials and transported to the laboratory for further analysis. At the laboratory, the mosquito larvae we 26 orted and separated from trash and other debris. Then, the volume and pH of the phytotelmic water were assessed. The dead larvae were separated from the living larvae and put into bottles containing 70% alcohol for toxonomic identification. Furthermore, the living larvae were kept alive and reared until reachs mature stage. Both the dead and living larvae of the mosquitoes were examined under a stereo microscope Nikon Olympus SZ51. The taxonomic procedures for larval identification and determination of genus Aedes referring to the work of Farajollahi and Price [9].

Data Analysis

The data resulted from the study were presented and analysed using descriptive statistics.

3. Results

There were five types phytotelmata found in the study namely leaf axils, tree holes, fallen spathe, fallen leaves, and open fruit shells (Fig. 1). Plant species the complete list of plant species with parts that fulfil the inclusion criteria as phytotelmata found in the study are presented in Table 1. Of all the phytotelmic sample, 22 of them were found to contain larvae of Aedes mosquito. The larval count of the Aedes mosquitoes and the physical parameters of the phytotelmic water are presented in Table 2.

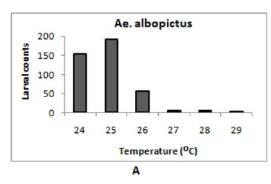
The data in **Table 2** can be elaborated as follows. There are two species of *Aedes* mosquito larvae managed to be identified in this study, i.e. *Ae.albopictus* and *Ae.crysolineatus*. The larvae can be found in the phytotelmata with a volume as low as of 8.5 ml, i.e. in the leaf axils of pineapple (*Ananas comosus*) and banana (*Musa paradisiaca*). In average, a single larvae of *Ae. albopictus* can even live in the water column with a volume of only 0.31 ml, such as in the hole of bamboo stumps where 125 larvae inhabit water column with a volume of 38.8 ml. Phytotelmic water temperature that can be occupied by *Aedes* larvae is ranged from 24 to 29°C, but seemingly the most preferred temperature is 24-26°C (**Fig. 2**). Phytotelmic water pH range where *Aedes* larvae are found is 5.5 - 9, with the most preferred pH by both species is 6 (**Fig. 3**).



Figure 1. Samples of plant parts that impound water serve as phytotelmata found in Lampung

Table 1. The plant species and plant parts that impound water serves as phytotelmata in Lampung

	Plant species		Plant parts that impound water					
No		Plant family	Leaf axils	Tree holes	Tree stumps	Fallen spathe	Fruit shells	Faller leaf
1	Colocasia esculenta	Araceae	√					
2	Alocasia macrorrhiza	Araceae	\checkmark					
3	Alocasia indica	Araceae	\checkmark					
4	Alocasia cupra	Araceae	\checkmark					
5	Dieffenbachia amoena	Araceae	\checkmark					
6	Dracaena fragrans	Asparagaceae	\checkmark					
7	Sanseviera trifasciata	Asparagaceae	\checkmark					
8	Ananas comosus	Bromeliaceae	\checkmark					
9	Guzmania sanguinea	Bromeliaceae	\checkmark					
10	Neoregelia sp.	Bromeliaceae	\checkmark					
11	Saccharum officinarum	Poaceae	\checkmark					
12	Pandanus amaryllifolius	Pandanaceae	\checkmark					
13	Musa paradisiaca	Musaceae	\checkmark			\checkmark		
14	Cocos nucifera	Arecaceae				\checkmark	$\sqrt{}$	
15	Areca catechu	Arecaceae				\checkmark		
16	Gigantochloa apus	Poaceae			\checkmark			
17	Gigantochloa atroviolacea	Poaceae			$\sqrt{}$			
18	Theobroma cacao	Malvaceae		\checkmark			\checkmark	
19	Artocarpus sp.	Moraceae						$\sqrt{}$
20	Mangifera indica	Anacardiaceae		$\sqrt{}$				
21	Cerbera manghas	Apocynaceae		$\sqrt{}$				
22	Manihot glaziovii	Euphorbiaceae		$\sqrt{}$				
23	Jatropha curcas	Euphorbiaceae		$\sqrt{}$				
24	Hevea brailiensis	Euphorbiaceae		$\sqrt{}$				
25	Leucaena leucocephala	Fabaceae		$\sqrt{}$				
26	Senna siamea	Fabaceae		\checkmark				
27	Acacia mangium	Fabaceae		$\sqrt{}$				
28	Archidendron pauciflorum	Fabaceae		$\sqrt{}$				
29	Gnetum gnemon	Gnetaceae		\checkmark				
30	Hibiscus tiliacius	Malvaceae		\checkmark				
31	Averrhoa carambola	Oxalidaceae		\checkmark				
32	Pyracantha angustifolia	Rosaceae		$\sqrt{}$				
33	Nephelium lappaceum	Sapindaceae		\checkmark				
34	Filicium decipiens	Sapindaceae		V				



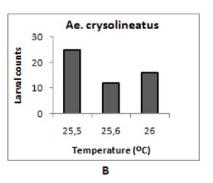
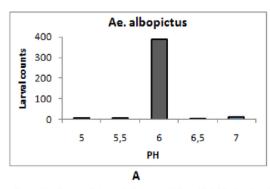


Figure 2. Range of phytotelmic water temperature that are occupied and preferred by Ae albopictus (A) and Ae crysolineatus (B)

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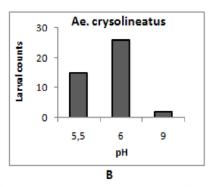


Figure 3. Range of phytotelmic water pH found inhabited and preferred by Ae albopictus (A) and Ae crysolineatus (B)

Table 2. The physical parameters of phytotelmata in which Aedes mosquito larvae are found

Campla	Phytotelmata		Physic	Physical parameters			Larval counts (n)	
Sample No.		Plant species	Volume (mL)	pН	Temp (°C)	Ae. albopictus	Ae. crysolineatus	
1	Leaf Axils	Colocasia esculenta	15.7	5.5	25.1	9		
2	Leaf Axils	Alocasia macrorrhiza	34.9	6	26	40	4	
3	Fruit shells	Cocos nucifera	152	5	26	5		
4	Fruit shells	Cocos nucifera	387.5	6	25.6	107		
5	Fallen spathe	Cocos nucifera	29.7	6	26	10	3	
6	Fallen spathe	Cocos nucifera	109	6	26		7	
7	Fallen spathe	Areca catechu	36	6	28	3		
8	Leaf Axils	Ananas comosus	10.8	6	25	2		
9	Leaf Axils	Ananas comosus	8.5	6.5	24.7	2		
10	Leaf Axills	Musa paradisiaca	22	6	28	3		
11	Leaf Axills	Musa paradisiaca	8.5	6	29	2		
12	Tree stumps	Gigantochloa apus	21	5	27	4		
13	Tree stumps	Gigantochloa apus	29.8	6	25.6	61	12	
14	Tree stumps	Gigantochloa sp	38.8	6	24.7	125		
15	Tree stumps	Gigantochloa sp	127	6	25.6	13		
16	Tree stumps	Gigantochloa sp	93	5.5	25.5		15	
17	Tree holes	Manihot glaziovii	14.5	7	26	2		
18	Tree holes	Leucaena leucocephala	79	6	24	16		
19	Tree holes	Senna siamea	40	7	24	6		
20	Tree holes	Theobroma cacao	126	6	27	2		
21	Fruit shells	Theobroma cacao	45.3	7	25	3		
22	Fruit shells	Theobroma cacao	29	9	26		2	

4. Discussion

This research was conducted in 2016, the year in which 4.523 people in the studied province found suffered from DHF and 15 of them were died. In the surveyed district namely Pesawaran, Metro, Pringsewu and Bandar Lampung, the DHF patients consecutively were 324 (one died), 222 (two died), 684 (three died) and 826 (one died) [10]. This study has confirmed the findings of previous studies that in nature, mosquito larvae, including the larvae of *Culex* spp. [11], *Ochlerotatus notoscriptus* [12] and *Ae albopictus* can breed in various small water bodies including the dried

fallen leaves and tree holes [13]. Interestingly, besides Ae. albopictus and Ae. crysolineatus, this study did not find any Ae. aegypti mosquito larvae in each phytotelmata that was investigated. The question is why?

The first possibility is *Ae aegypti* mosquito prefers to breed indoor in a small water body such as in the open masonry tanks used for water storage, or out door in a small water body trapped in the discarded goods such as old tyres and cans [14, 15].

The second possibility is the DHF in the area surveyed is not only transmitted by the *Ae aegypti* mosquitoes but also, or mostly, by the *Ae albopictus*. It is very likely for the Ae 10

albopictus, commonly called as the Asian tiger mosquito, is known as a secondary vector of dengue [16]. In Indonesia alone, precisely in Banjarnegara, Central Java, the role of Ae albopictus in dengue disease transmission is indicated by Rahayu and Ustiawan [17]. Of course, it should be recognized that the morphological appearance of both species is very resemble to each other. A strong evidence that Ae albopictus can contribute to the spread of dengue demonstrated by Rosa et al. [18]. By using RT-PCR methods for detecting dengue virus on Ae. albopictus larvae inhabit phytotelmata they found two serotypes of dengue viruses DEN-1 and DEN-4 and suggested that the mosquito has the potential to transmit and spread DHF.

The third possibility which account for Ae. aegypti are not found in phytotelmata is the larvae allegedly did not manage to compete with Ae. albopictus larvae. As shown by a study carried out in Malaysia, A 21 gypti has lower competitive advantage compared to Ae. albopictus in situation of limited resources. This situation allegedly the cause of the dominancy of Ae. albopictus mosquito in Penang Island [19]. The high adaptability of Ae albopictus 17 illustrated by the extent of the spread of mosquitoes that originally indigenous to South-east Asia and islands of the Western Pacific and Indian Ocean, to Africa, the mid-east Europe and the Americas (north and south) [20, 21].

Lastly, the conditions that may lead to the absence of Ae aegypti in the phytotelmata samples found in this study are the difference of physical condition suitable for the larvae. As revealed from an experimental study, the best water temperature for the Ae aegypti mosquito larvae to grow is 30°C [22], while the temperature of the phytotelmic water in where the larvae of Ae albopictus found in this study is 24 - 29°C.

5. Conclusions

This research was conducted in the year in which the incidence rate of DHF was high, however the study did not manage to find *Ae aegypti* in addition to Ae albopictus and Ae crysolineatus. Given Ae albopictus was revealed to contain viruses DEN 1 and DEN 4, and has been long known as a secondary vector of dengue fever, then it can be assumed 15. *Ae albopictus* could potentially be the DHF vector and phytotelmata might account for the high prevalence of the disease in Lampung, Indonesia.

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