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An Evaluation of Deep Neural Network Performance on Limited Protein Phosphorylation Site Prediction Data

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Abstract

One of the common and important post-translational modification (PTM) types is phosphorylation. Protein phosphorylation is used to regulate various enzyme and receptor activations which include signal pathways. There have been many significant studies conducted to predict phosphorylation sites using various machine learning methods. Recently, several researchers claimed deep learning based methods as the best methods for phosphorylation sited prediction. However, the performance of these methods were backed up with the massive training data used in the researches. In this paper, we study the performance of simple deep neural network on the limited data generally used prior to deep learning employment. The result shows that a deep neural network can still achieve comparable performance in the limited data settings.

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Keywords: Phosphorylation Site Prediction; Protein Phosphorylation; Deep Learning; Deep Neural Network

1. Introduction

One of the most important post-translational modifications [PTMs] is phosphorylation. With protein kinase, protein phosphorylation occurs when a phosphate group is added to an amino acid. These amino acids are Serine (S), Threonine (T), dan Tyrosine (Y)[1]. It is also the most used post-translational modification in eukaryotes[2 3] and play crucial

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roles in many cellular behaviour, such as metabolism[4], DNA repair[5], environmental stress response[6], regulation of transcription[7], and other important processes[8]. Therefore abnormality in protein can affect these cellular processes which may lead to many kinds of diseases. Because of that reason, it is important to identify and learn more about phosphorylation in the cell.

In general, there are two approaches to predict phosphorylation sites. The first approach is the kinase-specific phosphorylation prediction site. This approach requires information about the protein sequences, which are phosphorylated kinase enzymes. However, the main constraint of this approach is that kinase enzyme information for the public is limited[9]. The second approach is non-kinase-specific phosphorylation prediction site. This approach only requires the information of the phosphorylated protein sequences to predict the phosphorylated site. The differences and comparisons between these two approaches are explained by Xue et al. in their paper[10].

The typical studies of phosphorylation site prediction heavily employ machine learning algorithm as a site predictor. Following the recent trend in machine learning, recent studies of phosphorylation site prediction use deep learning with massive training dataset[11,12]. However, prior to 2017, the training dataset used is relatively small and limited to only 9 amino acids per sequence. Therefore, we cannot fairly compare the performance of deep learning based method with other well-established methods. In this study, we explore the performance of a simple deep neural network in the limited data settings typically used before 2017. We argue that, with the recent invention of various techniques in deep learning, a simple deep neural network is capable to achieve powerful performance even with limited training data.

2. Related Works

Since the advent of deep learning, the trend of machine learning research is shifted to the utilization of dataset with massive size. The emergence of this trend is due to the exceptional performance of deep learning given a massive training dataset. This trend is also apparent in phosphorylation site prediction. For instance, Musite Deep[11] and Deepphos[12] used dataset with 913,623 and 335,622 sites respectively.

The studies prior to Deep Musite typically used P.ELM[13] and PPA[14], which has only 4,750 and 852 sites. These studies generally employed popular machine learning models at that time, such as SVM[15,16,17,18] and Random Forest[15,19,20]. These models have limited performance on large data, thus they reduce the data further by cropping the protein sequences to 9 amino acids instead of using full sequence.

3. Materials and Methods

3.1. Materials and Datasets

The datasets used in this study are composed of polypeptide sequences, where each sequence consisting of 9 amino acid. We define this fixed length with 9 amino acid as window 9 sequence. The fifth amino acid or the amino acid in the middle of the sequence is the amino acid with the possible location for phosphorylation, Serine (S), Threonine (T), or Tyrosine (Y). Each sequence is labelled as a positive or negative sequence, where positive means that a phosphorylation event occurs on that location.

The window 9 sequence is generated from P.ELM database version 9[13] and PPA database[14]. The sequences are grouped according to their database source and phosphorylatable residues (Serine, Threonine, or Tyrosine). To reduce redundancy, sequences with the similarity of 0% to 20%, gap opening set the value to 10, and also the value of gap extension to 0.5, were removed using skipredundant[21]. We used the exact same datasets that were used by Lumbanraja et al[15] in their study. Table 1 below shows the size of each dataset.

3.1.1. Neural Network Architecture

The neural network model used in this study consists of 4 fully-connected layers. Prior to each fully-connected layer, we use Batch Normalization[22] layer to stabilize the training of the model. Every Dense layer has 32 neurons and exponential linear unit (ELU)[23] as the activation functions. We use an embedding model as a feature representation for each amino acid. The embedding model is inspired by word2vec model[24]. We modify word2vec model to encode each amino acid as a vector instead of words. The architecture of our neural network is illustrated in Figure 1.

Table 1: Datasets size.

Datasets	Serine		Threoninie		Tyrosine	
	Positive	Negative	Positive	Negative	Positive	Negative
P.ELM	1554	1543	707	453	267	226
PPA	307	307	68	68	51	51

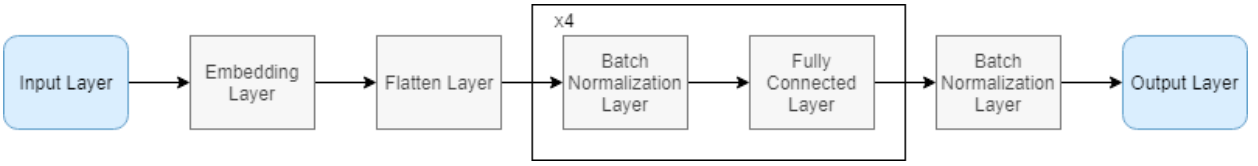

Fig. 1: Neural Network Diagram

To optimize the model, we use Adam optimizer[25] with standard configuration to optimize the model. The model is trained for 100 epochs with a scheduled learning rate decrease. The learning rate starts at 0.001, then it is reduced to 0.0005, 0.0002, and 0.0001 subsequently at 10th, 40th, and 70th epoch. To decrease overfitting, we utilize Dropout[26] with a drop rate of 0.1 and l2 regularization with a rate of 0.0001.

3.1.2. Evaluation

To evaluate our model, 10-folds cross-validation method is used to score our phosphorylation site prediction algorithm. Our datasets are split into 10 folds proportionately. Afterwards, we train and evaluate the model ten times, with each fold is used as validation split in turns.

To measure the performance of our method, we use the same metrics as in the study done by Lumbanraja et al[15]: Accuracy, Sensitivity, Specificity, F1 score, Area Under Curve (AUC)[27], and Matthews Correlation Coefficient (MCC). The metrics are formulated as follow:

$$Accuracy = \frac{TP + TN}{TP + TN + FP + FN} \tag{1}$$

$$Sensitivity = \frac{TP}{TP + FN} \tag{2}$$

$$Specificity = \frac{TN}{TN + FP} \tag{3}$$

$$F1score = \frac{TP}{TP + FP + FN} \tag{4}$$

$$MCC = \frac{(TP * TN) - (FP * FN)}{\sqrt{(TP + FP)(TP + FN)(TN + FP)(TN + FN)}} \tag{5}$$

After the neural network is trained and validated using the metrics above, we took the average score of the 10 validation scores to compare with the previous methods.

4. Result and Discussion

The performance of our proposed method is shown in Table 2 below. It can be seen that the performance is generally proportional to the size of the dataset. We can see that the performance of the neural network on P.ELM Serine dataset which is the biggest dataset, gave the best result. On the other hand, our deep neural network gave the lowest result

Table 2: Proposed Method's Performance.

Metrices	P.ELM			PPA		
	Serine	Threonine	Tyrosine	Serine	Threonine	Tyrosine
Dataset Size	3097	1160	493	614	136	102
Accuracy	0.9146	0.8733	0.7564	0.8109	0.8242	0.6409
AUC	0.9185	0.8708	0.7602	0.8104	0.8288	0.6565
Sensitivity	0.9305	0.8768	0.7272	0.8247	0.8034	0.6536
Specificity	0.9065	0.8648	0.7933	0.7962	0.8542	0.6595
F1-Score	0.9197	0.8939	0.7609	0.8111	0.8076	0.6339
MCC	0.8385	0.7362	0.5186	0.6211	0.6597	0.3120

on the PPA Tyrosine dataset, which is the smallest dataset. Therefore, we can conclude that, despite the limited data setting, the dataset size still plays an important role in deep neural network performance.

Currently, the best method for phosphorylation site prediction in limited data is feature extraction and feature selection developed by Lumbanraja et al [15]. Therefore, we compare our deep neural network to the Lumbanraja et al. method along with other well-established phosphorylation site prediction methods: Netphos K [28], GPS 2.1 [10], Swaminathan et al. method [17], Netphos [29], PPRED [30], PHOSFER [20], Musite [18], Phospho SVM [16], and RF-Phos [19]. We do not compare our method with another deep learning based method such as Musite Deep [11] and Deepphos [12], because these methods are tailored for full sequence setting instead of window 9 sequence setting. The comparison is shown in Table 3 and 4 for both datasets P.ELM and PPA. The best performance is marked with ***bold-italic*** font, while for the second best is marked with **bold** font. We can see that our simple deep neural network can deliver a comparable performance among the other methods. It is even the second best method overall behind Lumbanraja et al. method [15]. This suggests that a simple deep neural network can still deliver a powerful performance despite using limited training data.

Table 3: Result Comparison for P.ELM Datasets.

Method	Serine				Threonine				Tyrosine			
	AUC	Sens	Spec	MCC	AUC	Sens	Spec	MCC	AUC	Sens	Spec	MCC
Netphos K	0.63	0.51	0.68	0.08	0.6	0.62	0.57	0.07	0.6	0.39	0.74	0.08
GPS 2.1	0.73	0.33	0.93	0.2	0.7	0.38	0.93	0.2	0.61	0.34	0.79	0.08
Swaminathan et al.	0.7	0.31	0.89	0.13	0.72	0.28	0.92	0.14	0.62	0.60	0.57	0.09
Netphos	0.7	0.34	0.87	0.12	0.66	0.34	0.84	0.09	0.65	0.35	0.84	0.13
PPRED	0.75	0.32	0.92	0.17	0.73	0.30	0.91	0.13	0.7	0.43	0.83	0.17
Musite	0.81	0.41	**0.94**	0.25	0.78	0.34	***0.95***	0.22	0.72	0.384	**0.87**	0.18
Phospho SVM	0.84	0.44	**0.94**	0.3	0.82	0.38	***0.95***	0.25	0.74	0.42	**0.87**	0.21
Rf-Phos	0.88	0.84	0.85	0.65	**0.9**	0.83	**0.94**	0.7	***0.91***	**0.83**	***0.88***	***0.7***
Lumbanraja et al.	***0.96***	***0.97***	***0.96***	***0.93***	***0.92***	***0.93***	0.92	***0.84***	**0.8**	***0.84***	0.76	**0.6**
Our Method	**0.92**	**0.93**	0.91	**0.84**	0.87	**0.88**	0.86	**0.74**	0.76	0.73	0.79	0.52

5. Conclusions

In this paper, we show that a simple deep neural network can achieve comparable performance to other state-of-the-art models for phosphorylation site prediction with limited data. This study suggests that the remarkable performance of deep learning in phosphorylation site prediction is not only due to the massive dataset used for training. Therefore, the use of more complex deep learning method is suggested for future study in phosphorylation site prediction.

Table 4: Result Comparison for PPA Datasets.

Method	Serine			Threonine			Tyrosine		
	Sens	Spec	MCC	Sens	Spec	MCC	Sens	Spec	MCC
Netphos K	0.80	0.39	0.10	0.69	0.51	0.06	0.25	**0.83**	0.04
GPS 2.1	***0.95***	0.29	0.14	***0.96***	0.21	0.07	***0.98***	0.21	0.09
Netphos	0.77	0.54	0.16	0.54	0.77	0.12	0.65	0.67	0.13
PHOSFER	0.75	0.66	0.22	0.78	0.65	0.14	0.63	0.59	0.08
Musite	0.56	***0.87***	0.31	0.49	***0.94***	0.26	0.47	***0.89***	0.20
Phospho SVM	0.64	0.81	0.29	0.71	0.82	0.19	**0.82**	0.64	0.18
Rf-Phos	0.72	0.70	0.41	0.79	0.70	0.50	0.61	0.62	**0.29**
Lumbanraja et al.	**0.89**	**0.86**	***0.76***	**0.88**	***0.94***	***0.82***	0.53	0.63	0.16
Our Method	0.82	0.80	**0.62**	0.80	**0.85**	**0.66**	0.65	0.66	***0.31***

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