

EJSR

EUROPEAN JOURNAL OF
**SCIENTIFIC
RESEARCH**

Vol 38 No 4
December, 2009

USD \$25.00
CAD \$30.00



ISSN: 1450-216X

HALAMAN PENGESAHAN

Judul : Morusin, a Bioactive Compound from the Root Bark of *Artocarpus dadah*
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Sebagai : Anggota (Kelompok)
NIP : 197631082003121003
Program Studi : Pendidikan Dokter
Fakultas : MIPA
Buku/Jurnal/Prosiding : European Journal of Scientific Research
Halaman : 643 – 648
ISBN : 1450-216X Vol. 38 No. 4

Bandar Lampung, 8 Maret 2010
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DOKUMENTASI LEMBAGA PENELITIAN
UNIVERSITAS LAMPUNG

TGL.	10 Maret 2010
	173 / 1116 / 8 / pc / f-mipa / 2010
	jurnal

European Journal of Scientific Research

ISSN: 1450-216X

Volume 38, No 4 December, 2009

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Morusin, a Bioactive Compound from the Root Bark of *Artocarpus dadah*

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Morusin, a Bioactive Compound from the Root Bark of *Artocarpus dadah*

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Abstract

Morusin, a flavonoid prenylated at C-3 compound, has been isolated from the root bark of *Artocarpus dadah* which grown in Lampung. The structure of this compound has been identified by physical and spectroscopy methods. This compound has high cytotoxicity against murine leukemia cell P-388, IC_{50} 3,1 $\mu\text{g/mL}$.

Keywords: *Artocarpus dadah*, morusin, leukemia P-388.

1. Introduction

The last report of research to *Artocarpus dadah* plant which belong to Indonesian endemic plant (Lemmens *et al.*, 1995; Heyne, 1987; Jones and Luchsinger, 1987), from the root wood has been isolated a derivative compound of stilben, oxyresveratrol (Suhartati *et al.*, 2009). Previous researcher, Su *et al.* (2002) and Ersam (2001) have investigated the bark of *A. dadah*.

In our further research, from the root bark of *A. dadah* has successfully been isolated morusin (I), a prenylated (at C-3) flavonoid compound, which represent the first report this flavonoid found in *A. dadah*. The structure of this compound has been identified by physycal data as well as UV-Vis, IR and $^1\text{H-NMR}$ spectroscopis. By the finding of morusin in this plant, it has proven the Nomura hypothesis who stated that the marker compound from *Artocarpus* is a prenylated flavon compound at C-3 (Nomura *et al.*, 1998). In the cytotoxycity test using murine leukemia P-388 cell, morusin has shown high activity with IC_{50} value of 3.1 $\mu\text{g/mL}$.

2. Materials and Methods

2.1. Plant Material

The root bark *A. dadah* were collected from Wonoasri village, North Metro, Lampung, in March 2008 and were identified in Herbarium Bogoriense, Research Center for Biology, Indonesian Institute of Sciences, Bogor, Indonesia and a voucher specimen has been deposited at the herbarium.

2.2. General Experimental Procedures

Thin layer chromatography (TLC) analysis was carried out on pre-coated Si-gel plates (Merck Kieselgel 60 F254) and the UV lamp of Spectroline, ENF-240 C/F model was used to see the spot in TLC. VLC was carried out using Merck Si-gel 60. Melting point were determined on a Fisher Johns micro-melting point apparatus and were uncorrected. UV-Vis and IR spectra were measured with Beckman DU-7000 and Varian 2000 FTIR spectrophotometer respectively. $^1\text{H-NMR}$ spectrum was recorded with JEOL ECA 500 spectrometer, operating at 500.00 MHz.

2.3. Isolation and Purification of the Compounds

2.4 kg of root bark powder of *A. dadah* was macerated with methanol for 3 x 24 hours, with a maceration was 200 g. The methanol extract obtained was filtered and then evaporated by rotavapor at 45-50°C with velocity of 120-150 rpm. To the concentrate methanol extract was added 1% NaCl solution by proportion 1:4 to methanol extract, and then was partitioned with dichloromethane (DCM)-ethyl acetate 20%, to afford 151.28 g extract.

This extract was fractionated by VLC over Si gel, eluted with gradient mixture of methanol-DCM to afford four main fractions (A-D). The main fractions B (2.1675 g) and C (47 g) were fractionated with VLC over further Si gel using gradient mixture of ethyl acetate-*n*-hexane respectively. After passing the several ranks of VLC, CC, and flash chromatography using *n*-hexane, DCM, and ethyl acetate solvents with several concentration variation, the fractions which the same R_f in TLC were combined, then further purified by CC and flash CC. From this combined fraction was obtained the brown-yellow crystals (**1**) (25 mg), mp 118-123°C (crystallization in DCM-*n*-hexane). Chromatogram TLC of compound (**1**) using three eluent systems were showed one main spot R_f 0.20; 0.31; and 0.63 respectively using ethyl acetate-DCM 5%, ethyl acetate-*n*-hexane 30%, and ethyl acetate-DCM-*n*-hexane 3:3:4 eluent mixtures.

2.4. Bioactivity Test on the Pure Compound

The bioactivity test done includes the cytotoxicity test of compound (**1**) based on the method of Alley *et al.* (1988).

2.5. Structure Determination

The structure of pure compound was determined based on physical data and spectroscopy techniques, namely melting point, test with some specific reagent, spectra analysis of UV-Vis, IR and NMR.

3. Results and Discussions

3.1. The Analysis of Spectrometry

The UV-Vis spectrum obtained for brown-yellow crystal is shown in Figure 1, with absorption at maximum wavelengths 204, 279 and 328 nm. This UV spectrum indicates a flavonoid (Markham, 1980) which prenylated at C-3 on flavon skeleton (Suhartati, 2006), as shown in band I at λ_{max} 328 nm has lower intensity than band II at λ_{max} 279 nm. In NaOH addition, the spectrum showed bathochromic effect of band I 40 nm, which informed the presence of free OH group at C-4' on flavon skeleton. IR

spectrum of this compound (Figure 2) showed absorptions at 3365 cm^{-1} for OH group and conjugated carbonyl group at 1655 and 1620 cm^{-1} , while the presence of aromatic system was shown by absorption at $1597\text{--}1467\text{ cm}^{-1}$.

The ^1H NMR spectrum of compound (1) (Figure 3) confirmed the existence of aromatic skeleton and hydroxyl group in this compound, that is signals at (Figure 3) (aseton- D_6 , 500 MHz) δ (ppm): 13.57 and 8.85 singlet respectively for proton OH group at C-5 dan C-4', while aromatic proton ABX system were shown at 7.19 (1H, d, $J = 8\text{ Hz}$), 6.56 (1H, d, $J = 1.85\text{ Hz}$), and 6.52 (1H, dd, $J = 1.85$ and 8 Hz) on B ring; and 6.27 (1H, s) on A ring. Isoprenyl substituent at C-3 was shown by protons of two CH_3 groups at δ (ppm) 1.42 (3H, s) and 1.56 (3 H, s); and 3 proton of ABX system, that is chemical shift at 3.10 ppm (2 H, d, $J = 7\text{ Hz}$) and 5.11 ppm (1 H, t, $J = 7\text{ Hz}$). While 2,2-dimethylchromen from isoprenyl substituent at C-8 was shown by protons from two CH_3 group with chemical shift at 1.45 ppm (6 H, s) and two protons of a vinyl group which bound to C-8 on A ring at 6.67 ppm (1 H, d, $J = 10\text{ Hz}$) and 5.74 ppm (1 H, d, $J = 10\text{ Hz}$). Based on the ^1H -NMR data spectrum can be concluded that compound (1), was a prenylated flavon at C-3 containing two hydroxyl groups at C-2' and C-4', and 2,2-dimethylchromen which belong to isoprenyl at C-8. The flavon compound possessing spectrum data equivalent with compound (1) was morusin (Figure 4). The comparison of ^1H NMR data of compound (1) and morusin was shown in Table 1. By the finding of morusin in *A. dadah*, it is strengthened the hypothesis by Nomura *et al.* (1998) that this plant is part of *Artocarpus* genus, which contains flavon compound prenylated at C-3.

Figure 1: Ultraviolet spectrum of compound (1): (a) in MeOH (b) MeOH + NaOH

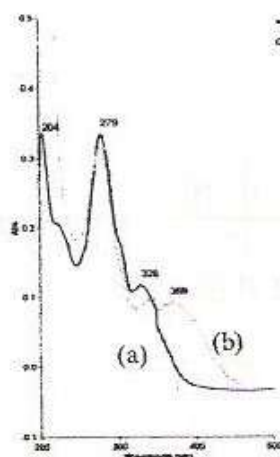
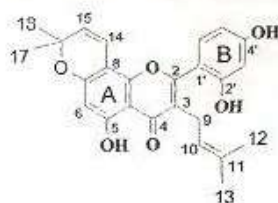


Figure 4: Molecular structure of morusin



3.2. Bioactivity Test

The bioactivity test of compound (1) using murine leukemia P-388 cells, compound (1) showed high cytotoxicity with IC_{50} value of 3.1 $\mu\text{g/mL}$. The possibility of high activity of compound (1) is due to the fact that compound (1) has two free hydroxyl groups at B ring and a prenyl group at C-3 similar to those of artonin E and artocarpin (Suhartati, 2001)

Conclusions

In this research, it has been successfully isolated compound (1) which was a prenylated flavonoid at C-3, morusin, which was the first reported from *A. dadah*. Compound (1) has high activity against murine leukemia P-388 cells with IC_{50} value of 3.1 $\mu\text{g/mL}$.

Acknowledgement

The authors would like to thank to The Directorate of Research and Community Services, Directorate of Higher Education, The Ministry of National Education of Republic of Indonesia that provide fund for this project to be undertaken through *Hibah Kompetitif Penelitian Sesuai Prioritas Nasional* Tahun Anggaran 2009, with contract number of 321/SP2H/PP/DP2M/VI/2009, 16 June 2009.

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